Soybean leaf hydraulic conductance does not acclimate to growth at elevated [CO₂] or temperature in growth chambers or in the field

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INTRODUCTION

A strong interdependency of leaf water transport capacity and photosynthetic capacity is expected from the principles of leaf gas exchange. Under natural mesophytic conditions, leaves lose several orders of magnitude more water to the atmosphere than they acquire CO₂ from the atmosphere. Thus, leaves must re-supply water to the sites of evaporation within the leaf mesophyll to enable the maintenance of open stomata for photosynthetic CO₂ acquisition without desiccating the leaf. Leaf hydraulic conductance (K_leaf) is a measure of water flow efficiency through the leaf and is defined as the water flux through the leaf per unit water potential driving force (Sack and Holbrook, 2006). Across sets of angiosperm species, K_leaf was found to correlate positively with maximum stomatal pore area per leaf area, mid-day stomatal conductance, photosynthetic electron transport rate and light-saturated CO₂ assimilation (Sack et al., 2003; Brodribb et al., 2007), suggesting evolutionary co-ordination between hydraulic and photosynthetic capacities of the leaf. Indeed, dynamic coordination of leaf and plant hydraulic conductance with gas exchange and photosynthesis has been observed in numerous species in response to environmental perturbation. Leaf net photosynthetic rates (A) were limited by whole-plant hydraulics under sufficient soil moisture conditions in Pinus ponderosa (Hubbard et al., 1999), and stomatal conductance (g_s) was limited by whole-plant and shoot hydraulics in several different deciduous and evergreen tree species (Salleo et al., 2000; Nardini and Salleo, 2000). Within given species, photosystem II quantum yield correlated with K_leaf on a diurnal cycle and also during leaf senescence (Brodribb and Holbrook, 2003, 2004). K_leaf plasticity has also been observed in response to dynamic changes in temperature and light (Sack et al., 2004; Scoffoni et al., 2008; Nardini et al., 2010), in some cases in coordination with A during growth under different environmental conditions (Brodribb and Jordan, 2011).

Atmospheric [CO₂] is expected to exceed 550 μmol mol⁻¹ (ppm) by mid-century and to drive increases of global temperature by 1–6 °C (Meehl et al., 2007). Elevated [CO₂] almost always leads to a reduction of g_s, lowering leaf- and canopy-level transpiration (Ainsworth and Long, 2005; Bernacchi et al., 2007). A lower transpiration rate should permit the reduction of K_leaf with no penalty to photosynthetic rate at elevated [CO₂]. Consistent with this expectation, whole-plant hydraulic conductance was reported to decrease in response to short-term exposure to elevated [CO₂] in chamber-grown Amaranthus hypochondriacus and Zea mays, as well as to long-term growth at elevated...
[CO₂] for chamber-grown *Glycine max* (soybean) and *Medicago sativa* (Bunce, 1996; Bunce and Ziska, 1998). Further, *K*_leaf decreased in *Pinus taeda* needles grown at elevated [CO₂] with free-air concentration enrichment (FACE) (Domenc et al., 2009).

The effects of elevated [CO₂] on soybean have been studied extensively because of its importance as the world’s third most economically valuable agricultural crop (Food and Agriculture Organization of the United Nations, 2010). With approx. 75 million acres of soybean planted annually in the USA, a thorough understanding of how soybean water relations respond to climate change is crucial to predicting how climate change will affect environmental processes and global food security. A generally increases at elevated [CO₂]. However, in field-grown soybean, the maximum velocity of carboxylation by ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco, *V*_c,max) tends to decrease during acclimation to growth at elevated [CO₂], thus restricting the degree to which elevated [CO₂] increases A (D. M. Rosenthal, USDA-ARS, Urbana, IL, USA, unpubl. res.). *Leaf* *g*ₜ consistently decreases at elevated [CO₂], but no acclimation of *g*ₜ to elevated [CO₂] has been observed across a decade of study at the SoyFACE field site (Leakey et al., 2006). The consistent decrease in upper canopy *leaf* *g*ₜ during growth at elevated [CO₂] is accompanied by decreased canopy evapotranspiration for field-grown soybean (Bernacchi et al., 2007). Despite the larger canopy area for elevated [CO₂]-grown soybean, the reduction in *g*ₜ significantly decreased water flow through the canopy, coupled with only a slight and inconsistent water potential (Ψ*leaf*) decrease, suggesting that *K*_leaf could be lower for soybean grown at elevated [CO₂] without limiting A (Bernacchi et al., 2007). That hypothesis was also supported by a recent field study in which intrinsic water use efficiency (A/Δ*g*ₜ) was observed to increase for soybeans grown at elevated [CO₂] (Ruiz-Vera et al., 2013).

Temperature and vapour pressure deficit (VPD) are major determinants of evapotranspiration. Global temperature increases throughout the 21st century will result in increased evaporative demand, but projections for VPD are less certain (Meeth et al., 2007). As temperatures rise with climate change, evapotranspiration (E) is likely to increase on a leaf area basis, as has been measured in soybean and *Z. mays* (Yang et al., 2012). With higher transpiration demand, *K*_leaf could become limiting to A if it has insufficient capacity to adjust, thereby causing reduction of *g*ₜ. A recent study with soybean grown at increased temperature in the field indeed found that *g*ₜ declined at higher temperatures, but intrinsic water use efficiency increased at higher temperatures due to biochemical properties of Rubisco (Ruiz-Vera et al., 2013). However, the stimulatory effect of temperature on photosynthesis declines at peak summer temperatures in many soybean-growing regions (D. M. Rosenthal, USDA-ARS, Urbana, IL, USA, unpubl. res.), and A will decline even as E continues to rise. There is evidence of a fast, reversible increase of *K*_leaf with rising temperature in *Acer saccharum*, *Aesculus hippocastanum* and *Quercus rubra* relating to changes in the viscosity of water and of membrane properties and/or aquaporin activity (Sack et al., 2004; Nardini et al., 2010). It is not known, however, if long-term growth at elevated temperature induces a more permanent increase in *K*_leaf.

Our objective in this study was to investigate the responses of *K*_leaf to growth at elevated [CO₂] and temperature in soybean. We hypothesized that for leaves grown at elevated [CO₂], *K*_leaf will decrease to reduce costly investment in water transport capacity while maintaining Ψ*leaf*, *g*ₜ and A. Furthermore, we hypothesized that for leaves grown at elevated temperature, *K*_leaf will increase to match higher E driven by an increase in VPD. These hypotheses were tested in two growth chamber experiments, one a factorial CO₂ × temperature experiment, and the second focused on temperature responses alone. Gas exchange parameters were measured along with *K*_leaf in these experiments. These chamber experiments were validated using soybean grown under FACE for [CO₂] and open-air, infrared temperature elevation in the field. Because the leaf is a critical component in the transpiration pathway, knowledge of leaf hydraulic responses and limits is necessary to be able to predict the extent to which gas exchange can adjust under increasingly extreme environmental conditions.

**MATERIALS AND METHODS**

**Plant material and growth conditions**

For both chamber experiments, seeds of soybean (*Glycine max* cultivar ‘93B15’ (Pioneer Hi-Bred, Johnston, IA, USA), a variety with indeterminate growth, were planted in 14.5-L pots with LC-1 Sunshine mix (SunGro Horticulture Canada Ltd, Bellevue, WA, USA). All seeds emerged from the soil within 4 d of planting. For the CO₂ × temperature factorial experiment, soybeans were germinated and grown in eight temperature- and CO₂-controlled growth chambers inside a greenhouse; four plants were grown per chamber. The chambers were constructed with aluminium frames and enclosed with clear acetate to allow entry of greenhouse light (Maherali and DeLucia, 2000). Natural light was supplemented with overhead lighting to reach approx. 750 μmol m⁻² s⁻¹ of photosynthetically active radiation (PAR) at plant level inside the growth chambers. The experiment was conducted from January to May 2010, with two separate sets of plants grown in succession. Measurements were taken on the youngest fully expanded leaves at 31, 36 and 43 d after planting in the first run of the experiment, and at 36, 38 and 43 d after planting in the second run of the experiment, so leaf developmental stage and leaf age were consistent for all measurements while plant age varies. Daylength varied greatly over the course of the experiment, but supplemental lighting was provided for 12 h d⁻¹ [CO₂] was continuously monitored in the centre of each chamber and fumigation was automatically adjusted to sustain the target. Treatments were applied in a 2 × 2 factorial design with [CO₂] treatments of 400 (ambient) and 700 μmol mol⁻¹ (ppm) (elevated) and daytime temperatures of 27 °C (ambient) and 31 °C (elevated). Elevated [CO₂] and temperature treatments began at seed planting, so plants experienced their assigned treatment conditions for their entire life span. Plants were randomly rotated among chambers weekly to reduce chamber effects, watered every other day, and fertilized twice weekly with 50 % Long Ashton solution supplemented with 10 mM NH₄NO₃ (Hewitt, 1966). In the CO₂ × temperature experiment, we were unable to control humidity, but it was high enough in all treatments to produce condensation on the walls of the growth chambers occasionally. High humidity may have obscured a possible response to temperature via VPD in the experiment.

For the temperature-only experiment, plants were grown in eight controlled environment chambers (GC-15, Environmental
Growth Chambers, Chagrin Falls, OH, USA). Twelve plants were grown in each chamber, and ambient and elevated temperature treatments were replicated in four chambers each. Daytime temperatures were 25 °C for ambient plants and 30 °C for the elevated treatment; night-time temperature was 22 °C and [CO₂] was 400 ppm for all plants. Elevated temperature treatment began at seed planting, so plants experienced their assigned treatment temperature for their entire life span. Measurements were taken on the youngest fully expanded leaves at 32, 39 and 41 d after planting. Plants were randomly rotated within chambers every 2 d and among chambers every 4 d to reduce chamber effects. Light levels were approx. 1200 μmol m⁻² s⁻¹ PAR at plant height. Plants were fertilized every other day with 50 % Long Ashton solution supplemented with 10 mM NH₄NO₃. Dataloggers were placed inside each chamber to record temperature and humidity at 15 min intervals (HOBO U12-012, Onset Computer Corporation, Inc., Pocasset, MA, USA). In this chamber experiment, relative humidity was carefully controlled at 60 % for both temperature treatments and continuously recorded, with an average VPD increase of 0.4 kPa for plants grown at elevated temperature (Fig. 1).

In 2010 and 2012, the same soybean cultivar ‘93B15’ (Pioneer Hi-Bred) was planted at the SoyFACE facility in Champaign, IL, USA in 0.38 m row spacing. Planting occurred on 27 May in 2010 and 15 May in 2012. A detailed description of the field site and SoyFACE CO₂ fumigation method can be found in Rogers et al. (2004). The [CO₂] experiment was conducted in a completely randomized block design. Each block consisted of two 20 m diameter rings, one at ambient [CO₂] and one fumigated with pure CO₂ to an elevated target [CO₂]. In 2010, ambient [CO₂] was 385 ppm and target elevated [CO₂] was 585 ppm; in elevated plots, [CO₂] was within 10 % of the target 75 % of the time. In 2012, ambient [CO₂] was 390 ppm. Elevated [CO₂] treatment began by the time seedlings emerged from the soil and continued for the full growing season, so all leaves developed entirely under their assigned treatment conditions. Measurements were taken at 32 and 50 d after planting.

In 2012, temperature elevation of 3.5 °C above ambient was achieved by placing infrared heaters above the canopy as detailed in Ruiz-Vera et al. (2013). Temperature treatment was applied continuously from 7 d after planting, with two exceptions from 10–15 July and 12–16 August, when electrical power was not available. After power was restored, measurements were not taken until at least one new leaf had fully developed under the elevated temperature treatment. Measurements were taken at 64 and 86 d after planting.

**Gas exchange**

Photosynthesis (A) and stomatal conductance (gₛ) were measured on the uppermost, fully expanded leaf on a plant using an open-path gas analysers equipped with a leaf chamber fluorometer (LI-6400, LI-COR Biosciences, Lincoln, NE, USA). Gas exchange was measured between 1200 h and 1400 h central standard time, as this typically corresponds to daily peak photosynthetic rates. Plants were briefly removed from growth chambers for gas exchange measurement, but [CO₂] temperature and PAR were set equal to growth conditions for each plant. Relative humidity in the gas exchange cuvette was maintained between 60 and 70 %. Different LI-6400s were used for the CO₂ × temperature chamber experiment and the temperature-only chamber experiment, but comparisons are only made between measurements taken with the same instrument.

**Leaf water potential**

Discs of 1.5 cm diameter were cut from uppermost mature leaves and sealed inside a stainless steel thermocouple psychrometer chamber within 15 s of cutting (Wescor C-30, Wescor, Inc., Logan, UT, USA). The psychrometer temperature was maintained at 22 °C in a controlled-environment chamber for 3 h until equilibrium temperature was achieved, and then the water potential of the leaf discs was recorded using a datalogger (Campbell CF-1000, Campbell Scientific, Logan, UT, USA). When pre-dawn Ψ_leaf was measured in the CO₂ × temperature experiment, leaves were collected between 0500 h and 0700 h. In this experiment, 12 leaf discs were sampled from the youngest, fully expanded leaf on each of two plants in each growth chamber. These discs were divided evenly among four psychrometer chambers for water potential measurement. Pre-dawn Ψ_leaf is assumed to be equal to soil water potential and was used to confirm that the treatments did not result in large soil moisture differences. There were only small differences in pre-dawn Ψ_leaf, with values ranging from −0.44 to −0.73 MPa across both experiments. When mid-day Ψ_leaf was measured in the temperature-only chamber experiment, three leaves were sampled per growth chamber, and three leaf discs were punched per leaf and measured in one psychrometer chamber. In this experiment, discs were sampled between 1300 h and 1400 h.

**Leaf hydraulic conductance**

K_leaf was measured using the evaporative flux method (Sack et al., 2002), with modifications as described below. Leaves

![Fig. 1. Average vapour pressure deficit (VPD) calculated from the four control and four elevated temperature chambers on every day of the experiment, beginning at planting. Temperature and relative humidity were logged at 15 min intervals; VPD was calculated for each of those intervals and then averaged over each 14 h daylight period. Ambient temperature chambers (25 °C) and elevated temperature chambers (30 °C) are as indicated. Relative humidity for all chambers was set at 60 %, although chamber dehumidification was insufficient to maintain this level as plants grew. Thus, relative humidity increased slightly and VPD decreased slightly over time.](image-url)
were cut at the base of the petiole immediately prior to turning on supplemental lighting for the growth chamber experiments or just prior to sunrise for the field experiments. Sampling at that time ensured that all leaves were fully hydrated and any emboli acquired during the previous day were refilled, so maximum \( K_{\text{leaf}} \) could be measured for each treatment. The youngest, fully expanded leaves were sampled for all measurements, so leaf age is consistent among all measurements, while plant age varied with the repeated sampling within experiments. Leaves of this soybean cultivar fully matured in approx. 8 d, so sampled leaf ages were within 4 d of each other, between the time the leaf becomes fully expanded and the time when it becomes shaded by a younger leaf above.

Evaporative flux measurements were made in the lab at ambient \([\text{CO}_2]\) and about 25 °C air temperature, so persistent but not quickly reversible effects of treatments were measured. Cut petioles were submerged immediately in water and re-cut under water. Crevices in the soybean petioles were blocked with petroleum jelly, and petioles were wrapped in Parafilm (Pechiney Plastic Packaging Company, Chicago, IL, USA) to ensure a tight seal with tubing that supplied water to the leaf (Tygon R-3603, Saint-Gobain Performance Plastics Corporation, France). This tubing was connected to a reservoir of ultrapure, partially degassed water situated on a high-precision balance (XS 250, Mettler Toledo, Columbus, OH, USA). The leaf was then placed under halogen lighting, with a glass tray of water placed between the lamp and the leaf to absorb infrared radiation, allowing approx. 700 m\(^2\)mol m\(^{-2}\) s\(^{-1}\) PAR to reach the leaf. A box fan was used to disrupt the leaf boundary layer, and water flow through the leaf was allowed to stabilize for at least 30 min. When water flow reached steady state, top and bottom leaf surface temperatures were measured (Fluke 574 Precision Infrared Thermometer, Fluke Corporation, Everett, WA, USA; calibrated using a black body calibrator, BB701, Omega Engineering, Inc., Stamford, CT, USA). Leaf discs were subsequently removed for \( \Psi_{\text{leaf}} \) determination using thermocouple psychrometers as described above; 12 discs were measured per leaf in four psychrometer chambers. Leaves were then photographed and leaf area was measured with ImageJ software (NIH, http://rsbweb.nih.gov/ij/). \( K_{\text{leaf}} \) was calculated as flow rate/leaf water potential and normalized for leaf area and the effect of temperature on water viscosity (Yang and Tyree, 1993). To increase throughput, four identical calibrators, BB701, Omega Engineering, Inc., Stamford, CT, USA; calibrated using a black body calibrator, BB701, Omega Engineering, Inc., Stamford, CT, USA) which was linked to a data logger (CR1000, Campbell Scientific, Inc.) which was linked to a data logger (CR1000, Campbell Scientific, Inc.) which was linked to a data logger (CR1000, Campbell Scientific, Inc.). This allowed water flow data from four leaves on four balances to be recorded in a single file and viewed simultaneously on a computer in real-time. Approximately 15 % of leaves wilted during evaporative flux measurement, and these values were not included in the analyses.

Statistical analyses

The SAS MIXED procedure was used for all statistical analyses (SAS 9.1, SAS Institute, Cary, NC, USA). For each experiment, data from multiple measurements days were pooled and analysed by repeated measures. This accounted for variation due to measurement day, as measurements were taken from the same replicate chambers or plots on multiple days in each experiment. \([\text{CO}_2]\) and temperature were always considered as fixed effects. In chamber experiments, each group of plants within a chamber was a replicate, and plants within these groups were treated as sub-samples. Because groups of plants were randomly rotated among chambers every 4 d, chamber effects were considered to be evenly distributed and were not included in the model. In field experiments, each treatment plot was a replicate, and plants within plots were considered as sub-samples. Plots were spatially blocked in the field, with one replicate of each treatment per block.

Optimizing \( \alpha \)

To avoid unnecessarily high rates of Type II error, \( \alpha \) values were optimized for hypothesis testing. Instead of minimizing only Type I or Type II error, this approach minimizes the average of Type I and Type II error, and therefore the overall error rate, which is optimal for a study in which Type I and Type II errors are considered to have equal consequence (Mudge et al., 2012). Degrees of freedom and Cohen’s \( \text{f}^2 \) were inputs for R code provided by Mudge et al. (2012). Degrees of freedom were taken from the data sets, and Cohen’s \( \text{f}^2 \) of 0.35 was chosen \textit{a priori} as representing a large effect size and corresponding 26 % of variance explained (Cohen, 1988). Based on the degrees of freedom in our study, the \( \alpha \) values generated for hypothesis testing are higher than the standard \( \alpha = 0.05 \) that is used to interpret most plant physiological data (Table 1).

### RESULTS

**\( K_{\text{leaf}} \) did not acclimate to growth at elevated \([\text{CO}_2]\)**

The \( K_{\text{leaf}} \) values obtained using the evaporative flux method were comparable with values observed in past studies with pot-grown soybeans and other herbaceous crop species (Sober, 1997; Tsuda and Tyree, 2000; Sack and Holbrook, 2006). In the \([\text{CO}_2] \times \) temperature experiment, growth at elevated \([\text{CO}_2]\) did not significantly affect \( K_{\text{leaf}} \) \( (P = 0.5466, \text{ d.f.} = 39) \) (Fig. 2A). Measurements with field-grown soybean showed similar results to the chamber experiment. Elevated \([\text{CO}_2]\) did not lead to a significant change in \( K_{\text{leaf}} \) for field-grown soybean under free-air \([\text{CO}_2]\) enrichment \( (P = 0.9852, \text{ d.f.} = 3) \) (Fig. 3).

**Elevated \([\text{CO}_2]\) increased carbon gain at both ambient and elevated temperature**

Elevated \([\text{CO}_2]\) increased \( A \) by 23 % in the \([\text{CO}_2] \times \) temperature experiment \( (P = 0.0009, \text{ d.f.} = 12) \) (Fig. 2C).

| Table 1. Optimal \( \alpha \) and \( \beta \) values used for hypothesis testing |
|-----------------|-----|-----|
| Degrees of freedom | Optimal \( \alpha \) | Optimal \( \beta \) |
| 2                | 0.38 | 0.40 |
| 7                | 0.25 | 0.30 |
| 12               | 0.18 | 0.21 |
| 39               | 0.04 | 0.05 |

Values were calculated according to Mudge et al. (2012); inputs were degrees of freedom from the data set and Cohen’s \( \text{f}^2 \) of 0.35, chosen \textit{a priori}. Degrees of freedom for each data set can be found in the figures.
[CO₂] decreased gs by 42% compared with ambient [CO₂] (P = 0.0093, d.f. = 12) (Fig. 2D).

**Growth at elevated temperature did not alter K_leaf**

In the CO₂ × temperature experiment, increased temperature did not change K_leaf (P = 0.4213, d.f. = 39) (Fig. 2A). Similarly, in the temperature-only chamber experiment, growth temperature did not alter K_leaf (P = 0.9542, d.f. = 4) (Fig. 3). Elevated temperature also did not affect K_leaf for field-grown plants in 2012 (P = 0.8002, d.f. = 7) (Fig. 5). Additionally, in the growth chamber experiment, mid-day V̇_leaf was not significantly altered by elevated temperature (P = 0.6731, d.f. = 2) (Fig. 4B).

**Temperature did not consistently affect gas exchange**

In the CO₂ × temperature experiment, temperature did not have a significant effect on A (P = 0.8691, df = 12) or gs (P = 0.7828, d.f. = 12) (Fig. 2B, C). Growth at elevated temperature also did not affect A (P = 0.4589, d.f. = 2) or gs (P = 0.8128, d.f. = 2) in the temperature-only chamber experiment (Fig. 4).

For each experiment, data for individual measurement days were also analysed individually. K_leaf across treatments was statistically different for measurement days in the temperature-only chamber experiment (P = 0.01, d.f. = 2) and in the [CO₂] field experiment (P = 0.06, d.f. = 13). Some variation was present in these data which was not consistent or significant across experiments; these results are presented separately (Supplementary Data Tables S1, S2, Figs S1–S4).

**DISCUSSION**

To our knowledge, we present the first measurements of K_leaf and its co-ordination with gas exchange in response to temperature and [CO₂] for field-grown soybean. K_leaf in field-grown soybean was maintained within a stable range of values for plants grown under open-air CO₂ fumigation or temperature elevation (Figs 3 and 5). Although there was some variation between measurement days for pooled data sets in the temperature chamber experiment and the [CO₂] field experiment (P = 0.01, d.f. = 2) and in the [CO₂] field experiment (P = 0.06, d.f. = 13). Some variation was present in these data which was not consistent or significant across experiments; these results are presented separately (Supplementary Data Tables S1, S2, Figs S1–S4).
hypothesis that \( K_{\text{leaf}} \) would be reduced at elevated [CO2] to match the decline of \( g_s \) and transpiration (Figs 2 and 3). Our data also suggested that the 46% decrease in whole-plant hydraulic conductance at elevated [CO2] previously reported from a chamber experiment with soybean (Bunce, 1996) probably did not involve a contribution from the leaves. Although that previous study did not measure \( K_{\text{leaf}} \), it reported a much greater [CO2] effect on stem hydraulic conductance than on root hydraulic conductance. In our study, gas exchange parameters responded as expected to elevated [CO2], with \( g_s \) decreasing and \( A \) increasing. That these responses were not reflected in \( K_{\text{leaf}} \) measurements indicates that soybean leaf gas exchange and leaf hydraulics are not closely coupled in their ability to acclimate to environmental conditions, and that \( K_{\text{leaf}} \) itself was not mechanistically influenced by growth [CO2] in the ways that have been recently hypothesized (Flexas et al., 2012), for example, due to developmental acclimation of aquaporin/CO2-porin activity. Notably, the high \( K_{\text{leaf}} \) relative to \( g_s \) in plants grown at high [CO2] could contribute to drought tolerance. Plants undergoing the onset of soil drying, or increases in VPD, can better maintain open stomata given high \( K_{\text{leaf}} \) relative to \( g_s \) (Brodribb and Jordan, 2008; Osborne and Sack, 2012). Furthermore, this insensitivity of \( K_{\text{leaf}} \) to growth [CO2] suggests that \( K_{\text{leaf}} \) was not limiting gas exchange under either the ambient or elevated [CO2] conditions tested in this study, which included field conditions.

\( K_{\text{leaf}} \) was similarly unresponsive to growth at elevated temperature (Figs 4A and 5). \( K_{\text{leaf}} \) has consistently been observed to increase with temperature in other studies, both at the time scale of minutes during measurement for A. hippocastanum, A. saccharum and Q. rubrum (Sack et al., 2004; Nardini et al., 2010) and with varying in situ leaf temperatures in Tilia cordata (Sellin and Kupper, 2007). While \( K_{\text{leaf}} \) did not show acclimation to growth at increased temperature in this study, evapotranspiration has been observed to increase for soybeans grown at elevated temperature in a chamber study (Allen et al., 2003). This finding suggests a lack of co-ordination of hydraulic and stomatal plasticity in soybean leaves.
Since the CO2 and temperature treatment differentials were not maintained during Kleaf measurements in this study, it is possible that soybean in a high temperature environment does have a higher Kleaf, but that this effect is transient and fully reversed when steady state was reached during the evaporative flux measurement. If so, this would suggest a lack of phenotypic plasticity for response to temperature in the structural components of the leaf which influence Kleaf, such as vein density (Sack and Frol, 2006; Brodribb et al., 2007). A previous study of the effect of growth [CO2] on Quercus petraea (350 ppm vs. 700 ppm) found no effects on vein density although stomatal density was reduced at high CO2 (Uhl and Mosbrugger, 1999), results analogous to our findings for Kleaf and gs in soybean. In contrast, elevated growth temperature was reported to increase Kleaf in Populus tremula when measurements were taken at a constant temperature (Aasamaa et al., 2005), and vein density is often found to increase in leaves grown under higher temperatures (Uhl and Mosbrugger, 1999; Sack and Scoffoni, 2013). Beyond vein density, the limited capability to adjust leaf hydraulic capacity observed in soybean may relate to its being a herbaceous annual bred under strong artificial selection. Greater Kleaf plasticity could also be more adaptive in tall plants than in short, herbaceous species. In Sclerocystis paniculatum, taller individuals had lower maximum Kleaf and lower Kleaf vulnerability than shorter individuals (Zhang et al., 2009). As cultivated soybean is short with ancestors that were vines, there may be less penalty for overall lack of hydraulic plasticity in the shoot. In summary, our data suggest a lack of phenotypic plasticity in soybean Kleaf during growth at elevated [CO2] and temperature. The responses of Kleaf and gas exchange to [CO2] and temperature do not appear to be mechanistically co-ordinated in soybean. This independence allows a shift in hydraulic supply relative to demand, such that plants grown at high [CO2] have high Kleaf relative to gs, and thus would be able to sustain higher gs and A during declines in soil water potential or high VPD. As gas exchange and leaf hydraulic conductance appear to be acting independently of each other in these studies, it is likely that Kleaf is not limiting to gas exchange under the conditions tested in these experiments. However, if Kleaf cannot be increased under elevated temperature, then it is possible that Kleaf could limit the delivery of water to points of evaporation within the leaf and thereby lead to a decline in leaf water potential, and a reduction of gs, that would limit A under such extreme weather conditions as are projected to become more frequent during this century (Meiell et al., 2007). Such a hydraulic limitation could be responsible for an increase in stomatal limitation to A at high temperatures, as had been previously observed in a study with field-grown soybean (D. M. Rosenthal, USDA-ARS, Urbana, IL, USA, unpubl. res.).

SUPPLEMENTARY DATA

Supplementary data are available online at www.aob.oxfordjournals.org and consist of the following. Table S1: degrees of freedom and P-values for each measurement day from all experiments, with measurement days analysed individually by ANOVA in SAS PROC MIXED. Table S2: optimal alpha and beta values used for hypothesis testing. Figure S1: leaf hydraulic conductance, photosynthesis and stomatal conductance for the CO2 × temperature experiment. Figure S2: leaf hydraulic conductance, midday leaf water potential, photosynthesis and stomatal conductance for the temperature-only chamber experiment. Figure S3: leaf hydraulic conductance for field-grown soybean under free-air [CO2] enrichment (FACE). Figure S4: leaf hydraulic conductance (Kleaf) for field-grown soybean. Data are shown for individual measurement days.

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LITERATURE CITED


**Hewitt EJ. 1966.** *Soybean leaf hydraulic conductance at elevated CO₂*.