Thus, the complete functional annotation of a protein is a time consuming, expensive and not always feasible (Slabinski et al., 2007). This motivated the development of proteome-scale automated methods for protein tertiary structure and molecular function prediction.

In recent years, we developed the protein structure prediction algorithm TASSER that employs a hierarchical approach consisting of template identification by threading, followed by tertiary structure assembly from continuous template fragments (Zhang and Skolnick, 2004). TASSER can often refine the threading templates generating final models closer to their native tertiary structure than the input templates (Zhang and Skolnick, 2004). Recently, TASSER was improved by incorporating information from consensus threading templates (METATASSER) (Zhou et al., 2007). The assessment of TASSER’s performance in CASP 6-8 shows that it is among the best structure prediction algorithms (Zhang et al., 2005; Zhou et al., 2007). Furthermore, to provide rapid results, while retaining TASSER’s ability to improve structure quality, we developed TASSER-Lite, a version applicable when the sequence identity between target and template is ≥25% (Pandit et al., 2006).

For function annotation, we have implemented FINDSITE (Brylinski and Skolnick, 2008) for ligand-binding site prediction, DBD-Hunter (Gao and Skolnick, 2008) for DNA-binding prediction and EFICAz² (Enzyme Function Inference by a Combined Approach) (Arakaki et al., 2009) for enzyme function inference. FINDSITE predicts ligand-binding pockets based on the binding site similarity among superimposed groups of template structures identified from threading. Here, threading acts as a filter to establish that the set of identified template structures are evolutionary related. FINDSITE also specifies the chemical properties of ligands that are likely to occupy the binding site and provides a collection of ligand templates for use in fingerprint-based virtual ligand screening. Furthermore, FINDSITE assigns Gene Ontology (GO) terms (Ashburner et al., 2000) with a probability that corresponds to the fraction of threading templates annotated with that molecular function (Brylinski and Skolnick, 2008). DBD-Hunter is a recently developed structure-based method for identifying DNA-binding proteins and associated binding sites (Gao and Skolnick, 2008). The method first selects potential DNA-binding proteins through structural comparison to known DNA-binding proteins, and further assesses DNA-binding propensity with a DNA–protein interfacial potential. Finally, EFICAz² is an enzyme function inference approach that employs machine learning techniques to combine predictions from six different methods developed and optimized to achieve high prediction accuracy (Arakaki et al., 2009).
2 PSIFR SERVER OVERVIEW

The Protein Structure and Function prediction Resource (PSIFR) server provides integrated tools for protein tertiary structure prediction and structure and sequence-based function annotation. Users can submit the protein sequence or structure with options for structure/function prediction (Fig. 1a) and a user-friendly output for visualizing the results is provided (Fig. 1b).

The results from tertiary structure prediction (TASSER/METATASSER) include the display of the threading alignments and protein models, visualized using Imol (Imol: an open-source Java viewer for chemical structures in 3D) (Fig. 1b). FINDSITE uses the top predicted model or the user’s submitted structure for function prediction. For each model, the results for the top five binding pockets are displayed. For each pocket, predicted ligand-binding residues and predicted GO terms are shown (Fig. 1b).

In addition, results of ligand-based virtual screening against the KEGG compound library are also displayed. DBD-Hunter reports whether or not the target protein is likely to bind DNA and, for putative DNA-binding proteins, shows the predicted DNA-binding residues. Similarly, results from EFICAz2 show whether the target protein is likely to be an enzyme, in which case the predicted three field or four field enzyme commission (EC) number is also reported. The important feature of PSIFR is that its component methods provide confidence measures of the prediction quality. This helps a user to assess the reliability of the predictions. The PSIFR web service is accessible via http://psifr.cssb.biology.gatech.edu/

Funding: National Institutes of Health (GM-48835, GM-37408 and RR-12255).

Conflict of Interest: none declared.

REFERENCES