The non-dystrophic myotonias: molecular pathogenesis, diagnosis and treatment

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The non-dystrophic myotonias are an important group of skeletal muscle channelopathies electrophysiologically characterized by altered membrane excitability. Many distinct clinical phenotypes are now recognized and range in severity from severe neonatal myotonia with respiratory compromise through to milder late-onset myotonic muscle stiffness. Specific genetic mutations in the major skeletal muscle voltage gated chloride channel gene and in the voltage gated sodium channel gene are causative in most patients. Recent work has allowed more precise correlations between the genotype and the electrophysiological and clinical phenotype. The majority of patients with myotonia have either a primary or secondary loss of membrane chloride conductance predicted to result in reduction of the resting membrane potential. Causative mutations in the sodium channel gene result in an abnormal gain of sodium channel function that may show marked temperature dependence. Despite significant advances in the clinical, genetic and molecular pathophysiological understanding of these disorders, which we review here, there are important unresolved issues we address: (i) recent work suggests that specialized clinical neurophysiology can identify channel specific patterns and aid genetic diagnosis in many cases however, it is not yet clear if such techniques can be refined to predict the causative gene in all cases or even predict the precise genotype; (ii) although clinical experience indicates these patients can have significant progressive morbidity, the detailed natural history and determinants of morbidity have not been specifically studied in a prospective fashion; (iii) some patients develop myopathy, but its frequency, severity and possible response to treatment remains undetermined, furthermore, the pathophysiological link between ion channel dysfunction and muscle degeneration is unknown; (iv) there is currently insufficient clinical trial evidence to recommend a standard treatment. Limited data suggest that sodium channel blocking agents have some efficacy. However, establishing the effectiveness of a therapy requires completion of multi-centre randomized controlled trials employing accurate outcome measures including reliable quantitation of myotonia. More specific pharmacological approaches are required and could include those which might preferentially reduce...
important largely unresolved questions in relation to morbidity and molecular pathogenesis of these disorders. We highlight caused by mutations in \( \text{SCN4A} \) the dominant feature \( \text{Venance et al.} \) myotonia in some cases, although episodic paralysis is usually channel gene \( \text{ders caused by point mutations in the skeletal muscle sodium sodium channel myotonias are allelic, autosomal dominant disor-

Introduction

The non-dystrophic myotonias are skeletal muscle ion channel disorders traditionally considered to be distinct from myotonic dystrophy because of the absence of progressive weakness and systemic features. The non-dystrophic myotonias are now known to be caused by dysfunction of key skeletal muscle ion channels and include myotonia congenita, paramyotonia congenita and the sodium channel myotonias. The worldwide prevalence of nondystrophic myotonia has been estimated to be \(~1\) in 100,000 (Emery, 1991). However, the prevalence seems to vary considerably between geographical regions. For example, myotonia congenita alone was estimated to have a prevalence of between 7 and 10 in 100,000 in Scandinavia (Baumann et al., 1998; Sun et al., 2001).

The major clinical manifestation of the non-dystrophic myotonias is muscle stiffness as a consequence of the myotonia. Additional common symptoms include pain, weakness and fatigue (Walsh et al., 2007; Trivedi et al., 2008; Wang et al., 2008c). Myotonia can be demonstrated on examination as delayed muscle relaxation following muscle contraction or following mechanical stimulation such as percussion. The underlying muscle membrane hyper-excitability manifests neurophysiologically as repetitive muscle fibre after-discharges on EMG. Recent studies have revealed a wide range of clinical phenotypes which may present diagnostic difficulty. Importantly, it has also been observed that patients with myotonic dystrophy type II may present with a clinical phenotype that is difficult to distinguish from myotonia congenita (Fialho et al., 2007). It is now clear that the clinical severity of these disorders can range from a neonatal life threatening presentation through to mild late-onset symptoms. The application of specialized electrophysiological protocols can reveal gene-specific patterns which can be used to direct DNA-based diagnosis (Fournier et al., 2004).

Myotonia congenita is caused by mutations in the skeletal muscle chloride channel gene \( \text{CLCN-1} \) and inherited in a domi-

Clinical features

Myotonia congenita

Myotonia congenita is the most common inherited skeletal muscle channelopathy. The autosomal dominant form was first described in the 19th century by the Danish physician Julius Thomsen in himself and his family (Thomsen, 1876). In the 1970s, the German Physician P.E. Becker fully documented the existence of the recessive form of myotonia congenita (Becker, 1977). In both forms muscle stiffness is most pronounced during rapid voluntary movements following a period of rest but improves with repeated activity—the so-called ‘warm-up’ phenomenon (Walsh et al., 2007; Trivedi et al., 2008; Wang et al., 2008c). Some clinical findings are more common in the recessive than in the dominant form but considerable overlap exists. Recessive myotonia congenita tends to be more severe, is more frequently associated with muscle hypertrophy and with depressed tendon reflexes (Becker, 1977; Fialho et al., 2007). Patients with recessive myotonia congenita typically experience a peculiar transient weakness on initiating an action, which is only rarely seen in dominant myotonia congenita. Although Becker found that most patients with recessive myotonia congenita presented between the ages 4 and 12 years while the dominant form usually manifested before the age of 3 years (Becker, 1977), we found no difference in the age of onset (Fialho et al., 2007).

Myotonic dystrophy types I and II can often be differentiated from myotonia congenita by the presence of systemic features. However, cases of myotonic dystrophy type II in which myotonia is the predominant complaint without any overt systemic features have been described (Fialho et al., 2007) and can lead to diagnostic difficulty.

Paramyotonia congenita

Eulenburg first used the term paramyotonia congenita in 1886 to describe a syndrome of episodic muscle cramps and paralysis profoundly exacerbated by cold and exercise in six generations of a German family (Eulenburg, 1886). The inheritance is autosomal dominant and symptoms usually manifest in the first decade of life. The facial, tongue, and hand muscles are predominantly affected and the lower limbs are generally only mildly affected (Miller et al., 2004). The myotonia can last seconds to minutes but the weakness may persist for hours and occasionally days. Paradoxical myotonia that worsens with exercise can be demonstrated at the bedside in most patients (Trivedi et al., 2008). Muscle hypertrophy is less frequent than in myotonia congenita.
but in our recent series we found it to be present in ~30% of patients (Matthews et al., 2008b).

**Sodium channel myotonia**

In 1987, prior to the availability of genetic testing, it was observed that there was a group of myotonic patients who seemed clinically distinct from either myotonia congenita or paramyotonia congenita. The first kindred reported exhibited autosomal dominant inheritance of a phenotype characterized by cold insensitive painful myotonia that was markedly exacerbated by potassium ingestion. None of the affected family members reported attacks of weakness but all experienced a significant improvement in myotonic symptoms with acetazolamide treatment. The term ‘acetazolamide-responsive’ myotonia congenita was coined to describe this family (Trudell et al., 1987; Ptacek et al., 1994). Subsequent reports described patients with a cold insensitive pure myotonic phenotype who did not experience weakness but whose myotonia fluctuated dramatically and was profoundly worsened by potassium ingestion. Notably the myotonia tended to occur with a more delayed (10–30 min) onset after exercise rather than with the initiation of movement after rest as seen in myotonia congenita, or within seconds of exercise as seen in paramyotonia congenita. This phenotype was classified as myotonia fluctuans (Ricker et al., 1990, 1994; Lennox et al., 1992). The term myotonia permanens was introduced to describe patients with a third clinical variant characterized by very severe persistent myotonia which significantly impaired respiration (McClellan et al., 1992b; Lerche et al., 1993). These three purely myotonic disorders shared the potassium aggravation and the absence of sensitivity to cold. Together they have become known as the potassium aggravated myotonias. Additional pure myotonic phenotypes have been described but these differ from the potassium aggravated myotonia phenotypes in that they have been reported to be cold-sensitive (Heine et al., 1993; Koch et al., 1995; Wu et al., 2001). All these pure myotonic phenotypes have now been shown to be caused by allelic point mutations in the gene encoding SCN4A.

We consider that from a practical clinical viewpoint a simplified classification of sodium channel myotonic disorders into two broad groups based on the presence or absence of episodic weakness is helpful:

**Group 1** Paramyotonia congenita—characterized by a marked worsening of myotonia by cold and by the presence of clear episodes of weakness;

**Group 2** Sodium channel myotonia—notable for the absence of episodic weakness but may have cold sensitivity. This includes all the pure myotonic phenotypes, including the potassium aggravated myotonias (Fournier et al., 2004, 2006).

Distinguishing chloride from sodium channel myotonias is often possible on clinical grounds alone as indicated in Table 1. However, difficulty may arise as some cases with sodium channel myotonia may have clinical features that are very similar to those seen in some cases of dominant myotonia congenita (see Table 1). For example, sodium channel myotonia may exhibit the presence of the warm up phenomenon, have minimal or absent sensitivity to cold, and have an upper limb/facial distribution of myotonia that is indistinguishable from dominant myotonia congenita (see Table 1).

In such cases the presence of transient weakness would point to dominant myotonia congenita whereas the presence of eyelid myotonia is more suggestive of sodium channel myotonia (Trip et al., 2009b). In addition specialized electrophysiological protocols can be helpful.

**Myopathy**

Myopathy may develop in some patients with non-dystrophic myotonia (Becker, 1977; Plassart et al., 1996; Nagamitsu et al., 2000). In a series of 49 genetically confirmed paramyotonia congenita cases, ‘myopathic biopsy findings’ were reported in 33% of those biopsied although full clinical details of the degree of weakness were not available (Miller et al., 2004). Permanent severe myopathy seems to be more common in patients with periodic paralysis than in the non-dystrophic myotonias (Miller et al., 2004). In periodic paralysis it has been postulated that the severity of myopathy may not relate to paralytic attack frequency (Buruma et al., 1978; Links et al., 1990) but the exact relationship remains unclear. There is some evidence that the severity of myopathy associated with periodic paralysis does correlate with increasing age (Links et al., 1990; Plassart et al., 1994). It is not known if a similar relationship between age and severity of myopathy exists in the non-dystrophic myotonias or if symptom frequency or severity has a direct influence on the development of myopathy. In periodic paralysis there is some evidence that the frequency of paralytic attacks may decline with age (Miller et al., 2004). However, it is not established if myotonia severity alters over time in patients with non-dystrophic myotonia. Importantly, there are no published studies to provide accurate detailed data on the natural history of the non-dystrophic myotonias in order to address the above questions. Such a large natural history study is currently in progress as part of the Consortium for Clinical Investigation of Neurological Channelopathies (http://rarediseasesnetwork.epi.usf.edu).

**Mechanisms of muscle degeneration**

Patients with periodic paralysis have been frequently reported to exhibit vacuoles and/or tubular aggregates on muscle biopsy. However, the myopathological findings in non-dystrophic myotonias are not defined well and are often reported to be non-specific (Miller et al., 2004). Furthermore, with the characteristic clinical history and examination findings coupled with the recent advances in electrophysiological techniques a diagnosis of non-dystrophic myotonia is usually apparent and it is now rare that a muscle biopsy will be performed in such patients other than as a research procedure.

It is clear muscle damage can occur in the non-dystrophic myotonias but its pathomechanism and frequency are unknown. It has been postulated that the abnormally prolonged intramuscular
influx of sodium that is known to occur via the mutant sodium channels may be responsible for muscle degeneration (Bradley et al., 1990). There is evidence for increased intracellular sodium contributing to cell necrosis in the mouse model of Duchenne muscular dystrophy (Hirn et al., 2008). One recent study has employed ultrasound to assess permanent muscle changes in the non-dystrophic myotonias. Using ultrasound measurements of eight muscles (four upper limb and four lower limb), in a group of 63 patients with genetically confirmed non-dystrophic myotonia an increase in the mean echo intensity compared with controls of 63 patients with genetically confirmed non-dystrophic myotonia was observed. The ultrasound changes were considered to indicate structural muscle damage such as fatty infiltration or fibrosis. This change was most marked in the forearm flexors where the increased echogenicity correlated negatively with muscle power. There was no positive correlation between echo intensity and age for individual muscles except the rectus femoris although the sum of the scores did show a significant positive correlation (Trip et al., 2009c).

Recently, a mouse model of hyperkalaemic periodic paralysis has been engineered by introducing the murine equivalent of the $SCN4A$ mis-sense mutation M1592V (Hayward et al., 2008). This mutation causes both myotonia and paralysis in humans (Rojas et al., 1991; Kelly et al., 1997) and was demonstrated to produce the same symptoms in the mouse indicating the validity of the model. At a few months of age the heterozygous mice displayed subtle myopathic changes. In homozygous mice significant muscle abnormalities were seen including an increase in fibre size variability, frequent internal nuclei and large scattered vacuoles. These changes were present before any spontaneous episodes of paralysis had been observed. Furthermore, they were shown to increase with age in the heterozygotes while muscle force generation declined (Hayward et al., 2008). These findings support the clinical observations that myopathy increases with age (Links et al., 1990; Plassart et al., 1994) and that it may be independent of paralytic attacks in the periodic paralyses (Buruma et al., 1978; Links et al., 1990).

It seems likely that future insights into muscle degeneration gained from the study of this model will also have implications for our understanding of paramyotonia congenita and sodium channel myotonia associated with the same $SCN4A$ gene. The possibility that myopathy develops independently of symptom frequency or severity may influence future approaches to therapy which is currently aimed at relieving symptoms. At present many patients with minimal or manageable symptoms decline pharmacological treatment but it is possible that treatment may have a role in preventing subsequent myopathy.

<table>
<thead>
<tr>
<th>Inheritance</th>
<th>Recessive myotonia congenita</th>
<th>Dominant myotonia congenita</th>
<th>Paramyotonia congenita</th>
<th>Sodium channel myotonia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Causative gene</td>
<td>Recessive CLCN-1</td>
<td>Dominant CLCN-1</td>
<td>Dominant SCN4A</td>
<td>Dominant SCN4A</td>
</tr>
<tr>
<td>Myotonia distribution</td>
<td>Lower limbs more than upper limbs</td>
<td>Upper limbs more than lower limbs</td>
<td>Upper limbs and face more than lower limbs</td>
<td>Upper limbs, face and extraocular, more than lower limbs</td>
</tr>
<tr>
<td>Myotonia cold sensitivity</td>
<td>None or minimal</td>
<td>None or minimal</td>
<td>Yes—often dramatic</td>
<td>Variable—ranging from none to severe</td>
</tr>
<tr>
<td>Warm up phenomenon</td>
<td>Present</td>
<td>Present</td>
<td>Absent</td>
<td>May be present</td>
</tr>
<tr>
<td>Paradoxical myotonia</td>
<td>Absent</td>
<td>Absent</td>
<td>Present</td>
<td>May be present</td>
</tr>
<tr>
<td>Delayed onset myotonia after exercise$^a$</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
<td>Characteristic of myotonia fluctuans Not reported</td>
</tr>
<tr>
<td>Episodic muscle weakness</td>
<td>Common, develops on initiation of movement but transient and improves rapidly</td>
<td>Uncommon</td>
<td>Common often exacerbated by cold and/or exercise and frequently prolonged for several hours</td>
<td>Common</td>
</tr>
<tr>
<td>Eyelid myotonia</td>
<td>Infrequent</td>
<td>Infrequent</td>
<td>Little or no decrement in CMAP (see Fig. 2)</td>
<td>No significant change of the CMAP from baseline$^b$ (see Fig. 3)</td>
</tr>
<tr>
<td>Short exercise test without cooling</td>
<td>Early decrement in CMAP with rapid recovery</td>
<td>Gradual and persistent reduction in CMAP enhanced by repetition (see Fig. 4)</td>
<td>Gradual and persistent reduction in CMAP enhanced further by cooling (see Fig. 4)</td>
<td>Gradual and persistent reduction in CMAP enhanced by repetition (see Fig. 4)</td>
</tr>
<tr>
<td>Short exercise test with cooling</td>
<td>Decrement reduces with repetition (see Fig. 1)</td>
<td>No significant change of the CMAP from baseline$^b$ (see Fig. 3)</td>
<td>No significant change of the CMAP from baseline$^b$ (see Fig. 3)</td>
<td>No significant change of the CMAP from baseline$^b$ (see Fig. 3)</td>
</tr>
<tr>
<td>Fournier pattern</td>
<td>II</td>
<td>II/III</td>
<td>I</td>
<td>III</td>
</tr>
</tbody>
</table>

$^a$ Myotonia typically develops after a short period of exercise e.g. 10 min or on resting after a period of exercise.

$^b$ This same pattern may be observed in dominant myotonia congenita. CMAP = compound muscle action potential.

Table 1 Common clinical features and electrophysiological patterns of the sub-groups of non-dystrophic myotonia
Morbidity in the non-dystrophic myotonias

Very little is known about the impact of non-dystrophic myotonias on quality of life and these disorders have often been regarded as benign. A single study has recently examined this in a group of 62 patients with genetically confirmed non-dystrophic myotonia and found painful myotonia and fatigue to be the best predictors of poor general health perception and physical functioning (Trip et al., 2009a). In this study painful myotonia was reported in 28% of those with myotonia congenita and 57% with sodium channelopathy. In addition, there are numerous case reports where severe pain is described (Ptacek et al., 1994; Rosenfeld et al., 1997; Vicart et al., 2004; Jorgensen et al., 2006; Colding-Fialho et al., 2007; Walsh et al., 2007; Wang et al., 2008c). This suggests that pain is a frequent symptom that may have been previously under-recognized and possibly undertreated in the non-dystrophic myotonias.

Clinical electrophysiology

Recently, specialized clinical neurophysiology protocols have aided precise diagnosis in muscle channelopathies by directing genetic testing based on channel-specific electrophysiological patterns. Sarcolemmal excitability can be measured indirectly as the variability of the compound muscle action potential (CMAP) following different stimuli. The CMAP size varies in skeletal muscle channelopathies in response to short (10–20 s) or long (3–5 min) exercise tests (Streib, 1982; McManis et al., 1986). Using these exercise protocols in combination with muscle cooling, distinct electrophysiological patterns—termed patterns I, II and III—are now recognized for each of the non-dystrophic myotonia groups (Fournier et al., 2004, 2006). For clinical diagnosis the repeat short exercise test with muscle cooling is of great value in the non-dystrophic myotonias.

Patients with chloride channel myotonia can show one of two patterns. The most common is pattern II (Fournier et al., 2004) in which at room temperature there is an immediate CMAP decrement after exercise which recovers quickly and diminishes with repetition, reflecting the transient weakness observed clinically (Fig. 1). This pattern is most frequently seen in recessive myotonia congenita but can be observed in any muscle ion channel disorder in which there is a loss of sarcolemmal chloride conductance. It is therefore also seen in dominant myotonia congenita and in both myotonic dystrophy types I and II. That this pattern may be seen in myotonic dystrophy types I and II is not unexpected as there is now clear evidence that the myotonia in these dystrophies is secondary to reduced chloride conductance (Charlet et al., 2002). In recessive myotonia congenita cooling has little further effect (Fig. 1). However, in dominant myotonia congenita, the CMAP decrement may be worsened or only seen with cooling (Fournier et al., 2006) making it essential to perform the short exercise test at both room temperature and with the muscle cooled (Fig. 2). Occasional patients with dominant myotonia congenita show a normal response [pattern III (Fournier et al., 2006)] to all provocative tests, even with muscle cooling which is indistinguishable electrophysiologically from sodium channel myotonia (Fig. 3).

Patients with paramyotonia congenita typically have a gradual and prolonged decrement in CMAP after exercise, termed pattern I (Fournier et al., 2004). This decrement is exacerbated with repeat testing and muscle cooling (Fig. 4) reflecting the clinically observed cold- and exercise-induced weakness. Some genotypes only display this typical pattern when the short exercise test is performed with the muscle cooled (Fournier et al., 2006) again emphasizing the importance in diagnosis of performing a short exercise test at both room temperature and with the muscle cooled.

The sodium channel myotonias are separated clinically from paramyotonia congenita by their lack of weakness. This is illustrated by pattern III, normal responses to all provocative tests (Fig. 3) with EMG myotonia as usually the only positive electrophysiological finding. This is the characteristic finding in sodium channel myotonia but is not absolute and there are some variations for certain genotypes (Fournier et al., 2006). It is notable that this is the same pattern observed in a minority of cases with dominant myotonia congenita.

The similarities between sodium channel myotonia and dominant myotonia congenita can lead to difficulty in prioritizing genetic testing. Clinical history and examination considered in...
conjunction with EMG findings (see Table 1) can improve the ability to distinguish between the two and guide genetic analysis, but in some cases screening of both the \textit{CLCN-1} and \textit{SCN4A} genes will be required. Variability exists in the response of the non-dystrophic myotonia subgroups to exercise testing and where muscle cooling has already proven useful in improving diagnosis, repetitive nerve stimulation may have a future role to play in distinguishing the sub-types of non-dystrophic myotonia. There is some evidence that a reduction in CMAP may be provoked by repetitive nerve stimulation in certain cases of recessive myotonia congenita where exercise testing even with the muscle cooled has failed to produce any such decrement (Michel et al., 2007). In this way, repetitive nerve stimulation may become an additional future tool to guide the genetic analysis towards recessive myotonia congenita, in cases that may otherwise be thought to be dominant myotonia congenita or sodium channel myotonia. There is currently no distinguishing electrophysiological test for myotonic dystrophy type II and this diagnosis should be considered for patients with a myotonic disorder in whom no mutations are found in \textit{CLCN-1} or \textit{SCN4A}.

### Genetics

**Skeletal muscle chloride channel**

Recessive and dominant myotonia congenita are caused by mutations in the voltage gated chloride channel gene on chromosome 7q35. (Koch et al., 1992; George et al., 1993). The functional chloride channel exists as a dimeric structure with two gating pores. To date, over 100 missense, non-sense, insertions, deletions and splice site mutations have been identified throughout \textit{CLCN-1}. Many patients carry ‘private’ mutations. It is a particular feature of myotonia congenita that several mutations have been reported to be inherited in both an autosomal dominant and autosomal...
recessive manner in different families (George et al., 1994; Meyer-Kleine et al., 1995; Zhang et al., 1996; Papponen et al., 1999; Sun et al., 2001). The advent of molecular genetic testing has demonstrated that familial non-dystrophic myotonia with dominant inheritance is often caused by missense mutations in SCN4A. Those CLCN-1 mutations that do cause dominant myotonia congenita seem to cluster in exon 8 of the gene (Fialho et al., 2007).

A recent report indicates that there is a higher frequency of recessive CLCN-1 mutations in myotonic dystrophy type II patients from Finnish and German populations. In the cases described, co-segregation of the CCTG expansion in the first intron of zinc finger protein-9 and a CLCN-1 mutation produced more severe myotonia than is commonly encountered in myotonic dystrophy type II. However, the number of cases was too small for this finding to be statically significant (Suominen et al., 2008). It is therefore not yet established if the presence of a CLCN-1 mutation is a genetic modifier of the myotonic dystrophy type II phenotype. This is an attractive hypothesis since both genetic changes would be predicted additively to impair chloride channel function. These findings also suggest that analysis of the myotonic dystrophy type II gene expansion should be considered in patients with myotonia only harboring a single recessive CLCN-1 mutation.

**Skeletal muscle sodium channel**

Hyperkalaemic periodic paralysis was the first of the sodium channel disorders to be linked to the SCN4A gene on chromosome 17 which encodes the skeletal muscle voltage gated sodium channel Na\textsubscript{v}1.4 (Fontaine et al., 1990; Koch et al., 1991b; Ptacek et al., 1991c). Given some of the shared clinical features of hyperkalaemic periodic paralysis and paramyotonia congenita and the recognition of abnormal sodium conductance in both (Lehmann-Horn et al., 1981, 1987a, b) it was proposed and subsequently confirmed that paramyotonia congenita and hyperkalaemic periodic paralysis were allelic disorders (Koch et al., 1991a; Ptacek et al., 1991a, b; Rojas et al., 1991; McClatchey et al., 1992a, b). Later the phenotypes grouped together as the potassium aggravated myotonia were also shown to be sodium channel disorders (Lerche et al., 1993; Ptacek et al., 1994; Ricker et al., 1994).

All the skeletal muscle sodium channelopathies are autosomal dominant conditions and de novo mutations can occur. In a proportion of patients with a phenotype typical for paramyotonia congenita no mutation has been identified in SCN4A raising the possibility of further genetic heterogeneity (Miller et al., 2004). Virtually all described mutations are missense with the only exception being a three base pair deletion (Michel et al., 2007).

Over 40 different mutations including those responsible for periodic paralysis have been reported in the SCN4A gene. Exons 22 and 24 are recognized as ‘hot spots’ for paramyotonia congenita, particularly the T1313M mutation and amino acid substitutions at the R1448 position (Mathews et al., 2008b). The most common sodium channel myotonia mutations are V1589M and those at the G1306 position (Vicart et al., 2005).

**Genetic diagnosis**

Although there are many clinical and electrophysiological indicators to help prioritize genetic testing in the non-dystrophic myotonias the difficulties described in distinguishing sodium channel myotonia from dominant myotonia congenita indicate that a proportion of patients will require screening of both the CLCN-1 and SCN4A genes (Trip et al., 2008). In some cases of recessive myotonia congenita, only one CLCN-1 mutant allele has been identified despite analysis of all coding exons (Trip et al., 2008). This indicates that mutations may be present in deeper intrinsic regions or possibly in promoter regions, although none have been reported. Another possibility is that there may be as yet unidentified large scale deletions in CLCN-1. Although it is recognized that some cases diagnosed clinically as non-dystrophic myotonia will have myotonic dystrophy type II, it is not known how frequent this is. Considering the importance of appropriate cardiac evaluation in myotonic dystrophy type II, we suggest that if CLCN-1 and/or SCN4A screening is negative, zinc finger protein-9 gene analysis should be undertaken.

**Genotype–phenotype correlations**

Marked phenotypic heterogeneity is common in the skeletal muscle channelopathies even in kindreds with the same mutation. Clinical and electrophysiological findings can help to distinguish between dominant and recessive myotonia congenita but since the same mutation can be inherited in a dominant or recessive manner this doesn’t necessarily narrow the possible genotypes. From a genetic point of view, non-sense mutations, small deletions and insertions leading to frameshift or mutations interrupting splice sites are usually associated with recessive myotonia congenita. However, missense mutations can lead to either recessive or dominant myotonia congenita depending on their location and the effect of the amino acid substitution on channel gating. Dominant mutations are clustered around the dimer interface of the channel (Duffield et al., 2003; Fialho et al., 2007) but are also found in other regions of the channel. R894X is the most studied mutation causing recessive and dominant myotonia congenita in different families. It is a non-sense mutation located within the C-terminus of the channel. Due to its location within the last exon of the CLCN-1 gene the mRNA does not undergo the usual non-sense mediated decay typically induced by earlier premature stop-codons. Duno et al. (2004) compared two families with dominant myotonia congenita and two families with recessive myotonia congenita carrying the R894X mutation. There was no direct relation between levels of CLCN-1 mRNA and inheritance type excluding differential allelic expression as an explanation for the varying inheritance mode. However, the most severely affected dominant case expressed more than twice the amount of mutant mRNA compared to the recessive families raising the possibility that this may contribute to phenotypic variability particularly within dominant pedigrees (Duno et al., 2004).
Paramyotonia congenita and sodium channel myotonia can usually be reliably distinguished from each other by the presence or absence of weakness from clinical and EMG findings. This narrows down the likely genotype to a certain degree but within each group there are still a number of possibilities. Supplementary Table 2 outlines the phenotypes reported for each mutation.

Now, there is evidence that certain SCN4A genotypes are associated with a severe neonatal phenotype. A fatal case of myotonia with significant respiratory muscle involvement was described in an infant with a de novo N1297K mutation (Gay et al., 2008). We observed the I693T mutation linked to spontaneously resolving neonatal hypotonia with variable feeding and respiratory difficulties in four unrelated families (Matthews et al., 2008a). The recognition that sodium channelopathies may present in such a way is important in order to provide appropriate prenatal advice for mothers known to carry these mutations and neonatal care for their children.

A further example of the phenotypic variability that can be observed with the same genotype is provided by G1306E mutation. The original phenotype reported with this was so severe the individual suffered permanent myotonia that included the respiratory muscles and which led to hypoxia and acidosis requiring ventilatory support (Lerche et al., 1993). In contrast, a more recent report observed that although affected individuals had relatively severe myotonia they did not exhibit respiratory involvement and were able to carry out daily activities including work without treatment (Colding-Jorgensen et al., 2006). See Supplementary Table 2 for more detail of phenotypes reported in different kindreds with the same SCN4A mutation.

One limitation in studying genotype-phenotype correlations has been the lack of availability of sufficient numbers of individuals carrying each mutation. The large multicentre natural history trial currently being run by the Consortium for Clinical Investigation of Neurological Channelopathies aims to address these issues.

Molecular pathophysiology

In a normal muscle fibre, a single nerve stimulus depolarizes the sarcolemma propagating a single action potential that results in a single muscle contraction followed by rapid relaxation. Myotonia results from an increased excitability of the muscle fibre membrane such that a single electrical stimulus triggers a repetitive train of action potentials.

In myotonia congenita, the enhanced excitability is due to reduced sarcolemmal chloride conductance and was initially demonstrated in muscles of myotonic goats (Lipicky et al., 1966; Bryant., 1969) and later in humans (Lipicky et al., 1971). Compared with other excitable cells, skeletal muscle has an unusually high chloride conductance, accounting for up to 85% of the resting membrane conductance (Bryant et al., 1971). The high chloride conductance is especially important in view of the large size of the muscle fibers which require the T-tubule system to propagate an action potential into the depth of the cell to initiate a synchronous contraction. Although T-tubules are directly connected to the extracellular space, they represent a significant diffusion barrier. Consequently with repeated membrane discharges, a build-up of potassium ions within the T-tubule system due to the repolarizing potassium ion currents increases the probability of additional spikes. The membrane depolarization as a consequence of potassium ion accumulation in the T-tubules is normally counteracted by the chloride conductance.

The chloride channel is an antiparallel assembled homodimer consisting of two identical subunits each with their own ion conducting pore (Fig. 5A). There are two main gating modes referred to as the fast gate, which can open and close the two pores independently, and a slow gating mechanism or ‘common gate’ which causes deactivation of both pores simultaneously. While all chloride channel mutations lead to loss of function, recessive mutations usually exert their effect by loss of function of the
mutated subunit, while the mutant subunit in dominant disease tends to have an adverse effect on the function of the co-expressed wild-type subunit, i.e. a dominant negative effect (Pusch et al., 1995). The majority of dominant myotonia congenita mutations shift the voltage dependence of CLCN-1 to more positive voltages (Pusch et al., 1995; Kubisch et al., 1998). Using a mathematical model, Barchi (1975) showed that decreasing the chloride conductance to 20% is sufficient to trigger myotonic discharges following a single stimulus. A similar hyperexcitability threshold of 25% was predicted by graded pharmacological inhibition of muscle CLCN-1 conductance (Kwiecinski et al., 1988). Clinically, a reduction to 50% does not seem to cause myotonia as evidenced by the majority of asymptomatic carriers of recessive myotonia congenita mutations. However, this assumes a 1:1 allelic expression in these cases, which may not always be the case (Chen et al., 1997). Supplementary Table 3 outlines details of known functional effects of all reported CLCN-1 mutations.

In contrast to the chloride channel, the voltage gated skeletal muscle sodium channel comprises a single ion conducting pore formed by the interaction between four homologous domains (Fig. 5B). All of the mutations associated with paramyotonia congenita and sodium channel myotonia produce ‘gain of function’ defects either by impaired inactivation or enhanced activation of the Nav1.4 channel (see Table 2 Supplementary Data). Impaired inactivation can either involve delayed inactivation or incomplete inactivation. Delayed inactivation of the skeletal muscle sodium channel causes increased excitability of the muscle fibre membrane and myotonia (Yang et al., 1994). The increased availability of sodium channels immediately after an action potential renders the fibre susceptible to sustained trains of repetitive discharges (Cannon, 2000), the electrophysiological hallmark of myotonia. In contrast to the repetitive firing seen in myotonia, the paralytic attacks experienced in paramyotonia congenita and in the allelic disorder hyperkalaemic periodic paralysis, are caused by episodic loss of fibre excitability. This sustained depolarization of the resting potential is due to sodium channels that do not inactivate completely, thereby conducting a persistent inward sodium current that depolarizes the fibre. (Bendahhou et al., 2002; Cannon, 2006).

Sodium channels also undergo a second, mechanistically distinct form of inactivation on a much slower time scale of seconds termed slow inactivation. Defects of slow inactivation increase the propensity for depolarization-induced attacks of weakness, and missense mutations of SCN4A that disrupt slow inactivation always result in a paralytic phenotype (Hayward et al., 1999).

Animal models

There are several animal models of myotonia congenita. The myotonic goat was bred by Dr H. H. Mayberry in Tennessee in the 1800s. This myotonic goat has been shown to harbour an alanine to proline substitution in the carboxyl terminus of the chloride channel (Beck et al., 1996). Other animal models include the arrested development of righting mouse which is caused by an insertion of a transposon element (Steinmeyer et al., 1991). More recently two different myotonic canine models have been described; the miniature Schnauzer and the Australian cattle dog with recessive missense mutations in CLCN-1 (Rhodes et al., 1999; Finnigan et al., 2007). The small size of the myotonic mouse makes it a difficult model upon which to undertake in vivo electrophysiology. Muscles from myotonic goats were very useful in the early experiments to elucidate the pathophysiology of myotonia congenita. However, in view of the substantial resource implications of goat care, goats are not considered ideal as a potential model for therapeutic trials. The maintenance of the myotonic dog models is also a major undertaking, but since many other canine models of disease have been extensively studied, greater technical experience is available indicating that myotonic dogs may be a more attractive model for future studies including therapeutic trials.

Hyperkalaemic periodic paralysis has been described in quarter horses. A single point mutation has been identified in the equine skeletal muscle sodium channel gene that substitutes a phenylalanine for a leucine in the DIV/S3 segment of the protein (Rudolph et al., 1992). Recently a mouse model of hyperkalaemic periodic paralysis was successfully engineered by introducing the equivalent of the common human M1592V mutation into the murine SCN4A gene (Hayward et al., 2008). The mouse was shown to display similar clinical and biopsy findings to those seen in human cases of hyperkalaemic periodic paralysis validating it as a reasonable animal model. New insights into the beneficial effects of elevated extracellular calcium levels and detrimental effects of impairing the sodium/potassium pump in the pathophysiology of hyperkalaemic periodic paralysis have already been determined (Hayward et al., 2008). This knock-in mouse offers potential for developing an increased understanding of the pathophysiology of hyperkalaemic periodic paralysis and possibly the development of future therapies.

Although hyperkalaemic periodic paralysis is allelic to paramyotonia congenita and sodium channel myotonia, no such phenotypes have been described in animal models. Myotonia does occur in horses but in conjunction with multisystem defects; the phenotype seems more like one of equine myotonic dystrophy (Reed et al., 1988).

**Treatment**

For those patients with mild symptoms no specific drug treatment may be needed, although it is important to provide advice regarding the avoidance of precipitating factors such as cold exposure or strenuous exercise. In those patients with significant symptoms and disability from myotonia, a variety of agents have been suggested which we outline in chronological order.

In early studies, procainamide, quinine and glucocorticosteroids were employed. A small randomized double blind trial compared the efficacy of each of these treatments in relation to placebo in 20 individuals with myotonic disorder (16 myotonic dystrophy, 4 myotonia congenita). The diagnosis was made on a clinical basis without genetic confirmation. The trial lasted 12 weeks and all participants received each of the four treatments for a 3 week period with no washout period. An end-point of at least a 50% reduction in the duration of hand grip myotonia,
measured by EMG and timed clinically, was employed. Using this endpoint, 6/20 participants taking quinine, 15/20 taking procainamide, 15/19 taking prednisone (one patient did not receive prednisone) and 0/20 taking placebo showed improvement (Leyburn and Walton, 1959). This study, although imperfect, illustrated a low efficacy of quinine. Despite the suggested benefits of procainamide and prednisone the side effect profile of both these drugs restricts their use and they are no longer recommended as therapeutic agents in the non-dystrophic myotonias.

The carbonic anhydrase inhibitor acetazolamide is commonly used in the periodic paralyses and has been reported to be beneficial in the non-dystrophic myotonias (Trudell et al., 1987; Ferriby et al., 2006). In a small series of nine patients with myotonia, seven diagnosed clinically with myotonia congenita and two with paramyotonia congenita, all cases reported a subjective and objective (timed measurements of myotonia) improvement in myotonia with acetazolamide. However, one individual with paramyotonia congenita developed quadrapareses 12 h after the ingestion of acetazolamide (Griggs et al., 1978). Larger studies of acetazolamide use in the non-dystrophic myotonias have not been performed, and while there is evidence of some benefit, it is not generally considered as a first line agent for the treatment of myotonia.

Anti-convulsants, local anaesthetics and anti-arrhythmic drugs which block sodium channels are the most frequently used agents in the treatment of myotonia. There are currently no safe drugs which specifically act on the chloride channel CLCN-1 (Verkman et al., 2009). Phenytoin has been shown to improve the righting time of myotonic mice turned onto their backs (Aichele et al., 1985). Ricker et al. (1978) reported subjective improvement in muscle stiffness and an improved timed walk in one patient with myotonia congenita and a dose dependant improvement in isometric force in another. The lignocaine derivative tocainide gave encouraging results initially (Rudel et al., 1980; Streib, 1987) but was eventually withdrawn from the market due to the risk of potentially fatal agranulocytosis (Volosin et al., 1985). Ricker et al. (1978) reported subjective improvement in muscle stiffness and an improved timed walk in one patient with myotonia congenita and a dose dependant improvement in isometric force in another. The lignocaine derivative tocainide gave encouraging results initially (Rudel et al., 1980; Streib, 1987) but was eventually withdrawn from the market due to the risk of potentially fatal agranulocytosis (Volosin et al., 1985). Synthesis of tocainide analogues has been attempted in vitro and it may be of value for future study as anti-myotonic agents if the efficacy and side effect profiles are favourable (Catalano et al., 2008).

More recently class I anti-arrhythmics have offered potential for treatment. Flecainide, a class Ic anti-arrhythmic, has been shown to be effective in vitro (Aoike et al., 2006) although its use in clinical practice as an anti-myotonic agent is rarely reported (Rosenfeld et al., 1997). An improvement in clinical symptoms and cold induced EMG findings with propafenone, another class Ic anti-arrhythmic has been reported in a single case of paramyotonia congenita (Alfonsi et al., 2007).

The class Ib anti-arrhythmic mexiletine is generally considered to be the first-line treatment of choice by myologists but a randomized controlled trial is required. It is usually well tolerated with only minor side effects reported. Importantly it has pro-arrhythmic potential and therefore pre- and post-treatment ECGs are essential to ensure satisfactory QT interval. More extensive cardiac evaluation prior to commencement is important if there is an abnormal baseline ECG or a history of cardiac disease. Single case reports have shown that mexiletine is effective in treating myotonia in both sodium and chloride channel disorders (Ceccarelli et al., 1992; Jackson et al., 1994). However, a recent Cochrane review highlighted the lack of adequate randomized double blind placebo controlled trials to prove efficacy (Trip et al., 2006). The ability to conduct such trials is partly hampered by the difficulty in quantitating myotonia (Torres et al., 1983; Hammaren et al., 2005; Logigian et al., 2005; Moxley et al., 2007; Hogrel, 2009) and in recruiting adequate numbers of patients to achieve statistical power. A recent study employed trunk sway analysis to measure the warm up phenomenon in recessive myotonia congenita and proposed that with further evaluation this may offer an alternative potential end-point for therapeutic trials (Horlings et al., 2009).

Sodium MRI has also recently been proposed as a possible outcome measure in patients with sodium channel diseases. An increase in intramuscular sodium content was demonstrated to accompany muscle weakness following exercise of cooled muscles in paramyotonia congenita. In a small group of patients this increase was significantly reduced following treatment with mexiletine (Weber et al., 2006). It is possible this technique could be used to monitor response to treatment in both a clinical and research setting. The Consortium for Clinical Investigation of Neurologic Channelopathies is currently performing a double-blind, placebo-controlled cross-over study of mexiletine for non-dystrophic myotonias (see clinicaltrials.gov) and will study 60 subjects with paramyotonia congenita and myotonia congenita. The primary endpoint measurement is patient stiffness as reported in the interactive voice response system (Wang, 2008c). The interactive voice response was chosen as the primary endpoint for this study as quantification of myotonia using handgrip devices has not consistently demonstrated the delayed relaxation phenomenon in patients with non-dystrophic myotonia (Wang, 2008c). It is hypothesized that patients’ self-reported responses of stiffness will be a more consistent and reliable endpoint measurement in order to determine efficacy of drug response.

In vitro studies continue to identify pharmacological agents that preferentially block sodium channels in the open state, thereby targeting persistent sodium currents (Wang et al., 2008a, b). These studies may identify future therapies.

No drugs are available which specifically act on the CLCN-1 channel. A number of experimental approaches may have future implications for the treatment of myotonia congenita (Cleland et al., 2008). It has been shown that alternative splicing of the CLCN-1 gene contributes to the myotonia in myotonic dystrophy type I and that directed anti-sense morpholino oligonucleotides to skip exon 7a restores chloride channel function and abolishes myotonia in a mouse model of myotonic dystrophy type I (Wheeler et al., 2007). It is possible that the development of this technique and its delivery could be a potential treatment for those cases of myotonia congenita due to splice site mutations.

Trans-splicing is a natural form of RNA processing where exons from two separate RNA transcripts are joined together. This can be manipulated to restore normal RNA processing of a mutant transcript. Such a technique has been employed using a trans-splicing ribozyme to restore a mutant chloride channel transcript in a cellular model. Although a good recovery of chloride channel function was observed in individual cells the efficiency of RNA
repair in the cell culture as a whole was only 1.2% (Rogers et al., 2002).

Improving defective CLCN-1 channel protein transport from the endoplasmic reticulum to the Golgi apparatus is another potential therapeutic strategy. Defective transport has been demonstrated for the F413C and A531V CLCN-1 mutations associated with recessive myotonia congenita (Papponen et al., 2008). Furthermore, functional expression of the F413C mutation in vitro showed only a minimal shift in chloride conductance (Zhang et al., 2000). This suggests that if the protein could be restored to the muscle membrane, this may result in an at least partial restoration of chloride conductance. Pharmacological therapies need to be developed that promote correct trafficking and which also build on the partial success of trans-splicing techniques safely.

It remains to be established if more specific therapies will not only reduce myotonic symptoms but, if given prophylactically, will also reduce the probability of developing a fixed myopathy.

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**Supplementary material**

Supplementary material is available at Brain online.

**References**


Appendix 1

This report summarizes the findings presented at the International Conference on Non-Dystrophic Myotonias (Kansas, June 2007). The conference was generously supported by a National Institutes of Health conference grant [R13 NS057995].

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