In vivo characterization of coronary plaques: novel findings from comparing greyscale and virtual histology intravascular ultrasound and near-infrared spectroscopy

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Aims
To test the hypothesis that near-infrared spectroscopy (NIRS) combined with intravascular ultrasound (IVUS) would provide novel information of human coronary plaque characterization.

Methods and results
Greyscale-IVUS, virtual histology (VH)-IVUS, and NIRS were compared in 131 native lesions (66 vessels) that were interrogated during catheterization by all three modalities. Greyscale-IVUS detected attenuated and echolucent plaques correlated with NIRS-detected lipid-rich areas. Attenuated plaques contained the highest NIRS probability of lipid core, followed by echolucent plaques. By VH-IVUS, 93.5% of attenuated plaques contained confluent necrotic core (NC) and were classified as VH-derived fibroatheromas (FAs). Although 75.0% of echolucent plaques were classified as VH-FAs, VH-NC was seen surrounding an echolucent zone, but not within any echolucent zone; and echolucent zones themselves contained fibrofatty and/or fibrous tissue. All calcified plaques with arc >90° contained >10% VH-NC (range 16.0–41.2%) and were classified as calcified VH-FAs, but only 58.5% contained NIRS-detected lipid core. A positive relationship between VH-derived %NC and NIRS-derived lipid core burden index was found in non-calcified plaques, but not in calcified plaques.

Conclusion
Combining NIRS with IVUS contributes to the understanding of plaque characterization in vivo. Further studies are warranted to determine whether combining NIRS and IVUS will contribute to the assessment of high-risk plaques to predict outcomes in patients with coronary artery disease.

Keywords
Coronary disease • Intravascular imaging • Near-infrared spectroscopy

Introduction
Greyscale-intravascular ultrasound (IVUS) is widely used for quantifying plaque distribution and severity, but, beyond the assessment of calcification, is limited to tissue characterization, especially for lipid-containing plaque; and the meanings of the greyscale-IVUS findings of echolucency and attenuation are not clear.1 Intravascular ultrasound radiofrequency data analysis, known as virtual histology IVUS (VH-IVUS), has been developed to improve on the plaque characterization of greyscale-IVUS and colour-coded plaque as either white (dense calcium, DC), red (necrotic core, NC), light-green (fibrofatty, FF), or dark-green (fibrous tissue, FT).2

Near-infrared spectroscopy (NIRS) is routinely used to characterize chemical composition of biological tissue.3 The ability of NIRS to discriminate lipid-rich atherosclerotic plaque with high sensitivity and specificity in vitro4 provides the possibility that NIRS can be used...
to detect lipid-rich atheromas in vivo. The major limitation of NIRS is that it provides compositional, but not structural, information.

The purpose of the present study was to use NIRS and VH-IVUS in comparison with greyscale-IVUS to better understand plaque morphology, morphometry, and composition. In particular, we evaluated three greyscale-IVUS findings that have been related to coronary disease progression: echolucent plaques and attenuated plaques have been associated with a more unstable clinical course, whereas calcified plaque has been associated with a more stable course.

**Methods**

**Study population**

From October 2008 through February 2010, we conducted a study to compare the in vivo characterization of coronary plaques among greyscale and VH-IVUS and NIRS. Patients were recruited from the population undergoing elective percutaneous coronary intervention (PCI) of native de novo coronary lesions for stable coronary artery disease or acute coronary syndrome (ACS). Each lesion selected for imaging was a culprit or target lesion undergoing planned PCI. Patients were excluded if they were pregnant or nursing, if life expectancy was <2 years, or if target vessels were coronary artery bypass grafts. Data of angiograms, NIRS, and IVUS were prospectively entered into the study database. Overall, greyscale and VH-IVUS and NIRS were performed in 78 consecutive patients (91 vessels). Sixteen vessels (12 patients) required pre-dilation prior to imaging and were excluded; and 9 vessels (7 patients) were excluded because of poor quality of IVUS ($n = 3$) or NIRS ($n = 4$) images. Thus, 66 vessels from 59 patients were included in the final analysis. No patient developed angina or had any complication (i.e. distal embolization, no-reflow, or thrombus formation) during IVUS and NIRS imaging, both of which were performed after intracoronary administration of nitroglycerin (100–200 μg). The greyscale-IVUS analysis, the VH analysis, and the NIRS analysis were performed without knowledge of the findings obtained from the other two methods.

This study was performed in accordance with the Code of Federal Regulations and the Declaration of Helsinki. The study protocol was approved by the Institutional Review Board. Informed consent was obtained from all patients before the procedure.

**Intravascular ultrasound image acquisition and analysis**

An Eagle Eye 20 MHz, 3.2Fr phased-array catheter (Volcano Corporation, Rancho Cordova, CA, USA) was advanced into the distal coronary artery and withdrawn to the aorta by motorized pullback (0.5 mm/s). During pullback, greyscale-IVUS was recorded, raw radiofrequency data were captured at the peak R-wave, and reconstruction of the colour-coded map by a VH-IVUS data recorder was performed. Image data were archived onto DVD and sent to the Cardiovascular Research Foundation, NYC, NY, for off-line analyses.

Greyscale analyses were performed using the validated planimetry software (EchoPlaque, INDEC Systems, Inc., Mountain View, CA, USA). Criteria for inclusion of a plaque in the study database were maximal plaque thickness ≥0.5 mm and plaque burden ≥40%, at least 1.5 mm in length, on greyscale-IVUS. Quantitative measurements were done according to criteria from the American College of Cardiology (ACC) consensus statement on IVUS. Qualitative assessments of greyscale-IVUS plaque morphology were based on the echogenic characteristics of the plaque on the cross-sectional view, and the adventitia was used as a reference for echoreflectivity. We emphasized three greyscale-IVUS plaque types: intraplaque echolucent zone (echolucent plaque); plaque with backward attenuation despite the absence of calcification (attenuated plaque); and calcified plaque. Echolucent plaque was defined as a large echolucent zone (thickness >0.3 mm) surrounded by tissue of greater echogenicity that was closer to the luminal surface than to the adventitia (shallow location). Attenuated plaque was defined as plaques with >30° ultrasonic attenuation (attenuation of deeper arterial structures) despite the absence of bright calcium. Calcified plaque was defined as hyperechoic plaque occupying >90° of arc with acoustic shadowing. Control plaque was defined as plaques with echogenicity similar to the adventitia, but without an echoluent zone, attenuation, or calcification.

**Near-infrared spectroscopy image acquisition and analysis**

Using the same protocol as for IVUS imaging, a 3.2Fr InfraReDx (Burlington, MA, USA) NIRS catheter was advanced into the distal coronary vessel. Automated mechanical pullback was performed at a speed of 0.5 mm/s and 240 rotations per minute until the NIRS catheter entered the guiding catheter. Raw spectra were acquired at a rate of ~40 Hz (one spectrum every 25 ms). Spatial filtering and image processing of the raw data produced an image with data points every 0.1 mm and every 1°.

During catheter pullback, the measurement of the probability of lipid core is displayed as an NIRS ‘chemogram’ (Figure 1, top), a digital colour-coded map of the location and intensity of lipid core, with the X-axis indicating the pullback position in millimetres (every 0.1 mm) and the Y-axis indicating the circumferential position in degrees (every 1°) as if the coronary vessel has been split open along its longitudinal axis. Spectroscopic information at each pixel is transformed into a probability of lipid core that is then mapped to a 128 (7-bit) red-to-yellow colour scale, with the low probability of lipid shown as red and the high probability of lipid shown as yellow. If a pixel does not contain enough data (e.g. as caused by guidewire shadowing), it appears black—i.e. a ‘non-viable’ pixel.

The ‘block chemogram’ (Figure 1, bottom) is a summary metric that is computed to display the probability that a lipid core plaque (LCP) is present for all measurements using the top 10th percentile pixel information (i.e. the 90th percentile value) of the corresponding 2 mm NIRS ‘chemogram’ segment. If the probability of the top 10th percentile is ≥0.98, the entire block is assigned yellow; if the probability of the top 10th percentile is 0.84–0.98, the entire block is assigned tan; if the probability of the top 10th percentile is 0.57–0.84, the entire block is assigned orange; if the probability of the top 10 percentile is <0.57, the entire block is assigned red. The ‘block chemogram’ provides a summary of the data to enhance interpretation of the chemogram and does not indicate individual pixel data or the location of a measurement in the circumferential dimension.

Near-infrared spectroscopy image analyses were performed off-line using the in-house, Matlab-based software programmed at the...
Cardiovascular Research Foundation, which read the information of each pixel and counted total viable and non-viable pixels. The presence of lipid core within the region of interest required at least one yellow block (95% specificity that lipid core was present). Yellow pixels (pixels above the preset threshold for the detection of LCP) within the analysed segment were divided by all viable pixels in the ‘chemogram’ to generate the lipid core burden index (LCBI) per mille (‰). Total LCBI, the 2 mm length with the maximum LCBI, and mean and maximum angles of lipid core were computed using the entire spectroscopic information within the region of interest in the ‘chemogram’ and exported by the NIRS software automatically.

Registering intravascular ultrasound and near-infrared spectroscopy images

Important angiographic landmarks during the pullback were imprinted on the chemogram—i.e. starting NIRS position, starting position of target lesion, side branches, and distal edge of the guiding catheter, etc. Those same landmarks were also bookmarked on the IVUS images. Near-infrared spectroscopy images were matched to IVUS images using known landmarks on both modalities and with the aid of interpolated lengths calculated using catheter pullback speeds. Because each chemogram block measured 2 mm in length, separate lipid cores (on NIRS) required three consecutive non-yellow chemogram blocks between yellow chemogram blocks, whereas separate greyscale and VH-IVUS lesion types required 6 mm separation between them.

Statistical analysis

Data analyses were performed using SPSS version 12.0 (SPSS, Inc., Chicago, IL, USA) and SAS version 9.1 (SAS Institute, Inc., Cary, NC, USA). For patient-level data, categorical data were expressed as absolute values and percentages and compared using χ² or Fisher’s exact test, and continuous data were reported as median (inter-quartile range, IQR) and compared using Kruskal–Wallis/Wilcoxon rank-sum tests. For lesion-level data, a model with generalized estimating equation (GEE) approach was used to compensate for any potential cluster effect of multiple lesions in the same individual. The correlation between VH-IVUS-derived parameters vs. NIRS-derived parameters was analysed using Spearman correlation coefficients. P-values were adjusted with the GEE method for repeated measures. Inferential statistical tests were conducted at the significance level of 0.05.

Results

Patient characteristics

Baseline patient characteristics are shown in Table 1. The length of the artery imaged was 93.4 ± 20.8 mm in the left anterior descending artery (LAD) to the ostium of the left main coronary artery (LMCA), 67.9 ± 17.5 mm in the left circumflex (LCX) to the ostium of the LMCA, and 94.4 ± 26.4 mm in the right coronary (RCA) to its ostium.
Table 1  Clinical characteristics (patients = 59; vessels = 66)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>61 (57–68)</td>
</tr>
<tr>
<td>Male, n (%)</td>
<td>54 (91.5)</td>
</tr>
<tr>
<td>Medical history, n (%)</td>
<td></td>
</tr>
<tr>
<td>Current smoker</td>
<td>22 (37.3)</td>
</tr>
<tr>
<td>Hypertension</td>
<td>55 (93.2)</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>25 (42.4)</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td>53 (89.8)</td>
</tr>
<tr>
<td>Prior myocardial infarction</td>
<td>24 (40.7)</td>
</tr>
<tr>
<td>Prior coronary bypass</td>
<td>11 (18.6)</td>
</tr>
<tr>
<td>Family history</td>
<td>28 (47.5)</td>
</tr>
<tr>
<td>Clinical presentation, n (%)</td>
<td></td>
</tr>
<tr>
<td>Non-ST-elevation myocardial infarction</td>
<td>12 (20.3)</td>
</tr>
<tr>
<td>Unstable angina</td>
<td>14 (23.7)</td>
</tr>
<tr>
<td>Stable angina</td>
<td>31 (52.5)</td>
</tr>
<tr>
<td>Atypical chest pain</td>
<td>2 (3.4)</td>
</tr>
<tr>
<td>Target vessel, n (%)</td>
<td></td>
</tr>
<tr>
<td>Left anterior descending</td>
<td>32 (48.5)</td>
</tr>
<tr>
<td>Left circumflex</td>
<td>16 (24.2)</td>
</tr>
<tr>
<td>Right coronary artery</td>
<td>18 (27.3)</td>
</tr>
</tbody>
</table>

Data are presented as median (IQR) or number (%).

Overall greyscale and virtual histology-intravascular ultrasound findings

A total of 131 plaques in 66 vessels that met all inclusion criteria were included into the study database. From the database, we identified 16 echolucent plaques in 13 vessels (6 LAD, 3 LCX, and 4 RCA); 31 attenuated plaques in 26 vessels (11 LAD, 6 LCX, and 9 RCA); 41 calcified plaques in 31 vessels (17 LAD, 6 LCX, and 8 RCA), and 30 control plaques in 30 vessels (15 LAD, 8 LCX, and 7 RCA). Virtual histology-IVUS findings are summarized in Table 2. Overall, VH-derived %NC was 23.5% (IQR 17.4–29.6%) with a maximum %NC of 34.5% (IQR 27.4–38.6%). The mean NC angle was 62.5° (IQR 27.3–106.8°) with a maximum NC angle of 101.0° (IQR 54.5–191.5°).

Overall near-infrared spectroscopy findings

Near-infrared spectroscopy findings are summarized in Table 2. Overall, lipid core was detected in 75 lesions (57.3%) in 41 vessels (20 LAD, 10 LCX, and 11 RCA). In the ‘chemogram’, the overall LCBI was 101.2 (IQR 31.4–237.5) with a mean lipid core angle of 39.3° (IQR 16.3–89.3°). In the ‘block chemogram’, there were 15.1% yellow blocks (highest probability of lipid core) and 60.1% red blocks (lowest probability of lipid core). The proportion of tan and orange blocks—indicating that lipid core was present, but with a probability less than that of yellow—were 9.2 and 15.7%, respectively. Overall, there was no significant correlation between LCBI and plaque burden (Rho = 0.26, P = 0.103).

Attenuated plaques

As shown in Figure 2, attenuated plaques had a greater remodelling index and were more eccentric compared with control plaques. As shown in Figure 3 and compared with control plaques, attenuated plaques had more %NC (P < 0.001) as well as more VH-derived FAs (93.5 vs. 60.0%, P = 0.002) and VH-derived TCFAs (45.2 vs. 20.0%, P = 0.041). However, there were similar amounts of NC [26.0% (22.1–31.3%) vs. 27.8% (22.3–34.5%), P = 0.762] when comparing calcified vs. attenuated plaques. Qualitatively, most (93.5%) attenuated plaque contained confluent NC when analysed by VH-IVUS; an example is shown in Figure 4.

On NIRS analysis, 90.3% of attenuated plaques contained lipid core as identified by the presence of at least one yellow block on the block chemogram (Figure 5). As shown in Figure 6, compared with control plaques, attenuated plaques had a higher LCBI, a larger maximum 2 mm LCBI, and a greater mean and maximum angles of lipid core (all P < 0.001). Similarly, there were a significantly higher LCBI, maximum 2 mm LCBI, and mean angle and maximum angles of lipid core in attenuated vs. calcified plaques (all P < 0.001). As shown in Figure 7, the probability of lipid core was highest in attenuated plaques, as indicated by the greatest percentage of yellow blocks.

Echolucent plaques

As shown in Figure 2, there was a higher eccentricity index in echolucent plaques when compared with control plaques (P = 0.005).

Table 2  Virtual histology-intravascular ultrasound and near-infrared spectroscopy findings (plaques = 131)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Median (IQR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>VH-IVUS parameters</td>
<td></td>
</tr>
<tr>
<td>Mean NC area (mm²)</td>
<td>0.9 (0.4–1.9)</td>
</tr>
<tr>
<td>Mean FF area (mm²)</td>
<td>0.7 (0.4–1.4)</td>
</tr>
<tr>
<td>Mean FT area (mm²)</td>
<td>3.7 (1.7–5.7)</td>
</tr>
<tr>
<td>Mean DC area (mm²)</td>
<td>0.3 (0.1–0.6)</td>
</tr>
<tr>
<td>%NC</td>
<td>23.5 (17.4–29.6)</td>
</tr>
<tr>
<td>%FF</td>
<td>10.8 (6.5–16.1)</td>
</tr>
<tr>
<td>%FT</td>
<td>49.6 (42.7–60.9)</td>
</tr>
<tr>
<td>%DC</td>
<td>10.7 (5.5–19.1)</td>
</tr>
<tr>
<td>Mean angle of NC (degree)</td>
<td>62.5 (27.3–106.8)</td>
</tr>
<tr>
<td>Maximum angle of NC (degree)</td>
<td>101.0 (54.5–191.5)</td>
</tr>
<tr>
<td>NIRS parameters</td>
<td></td>
</tr>
<tr>
<td>LCBI</td>
<td>101.2 (31.4–237.5)</td>
</tr>
<tr>
<td>Maximum 2 mm LCBI</td>
<td>157.9 (57.5–322.3)</td>
</tr>
<tr>
<td>Mean angle of LCP (degree)</td>
<td>39.3 (16.3–89.3)</td>
</tr>
<tr>
<td>Maximum angle of LCP (degree)</td>
<td>83.0 (35.0–137.0)</td>
</tr>
</tbody>
</table>

Data are expressed as median (IQR). DC, dense calcium; FF, fibrofatty; FT, fibrotic tissue; LCBI, lipid core burden index; LCP, lipid core plaques; NC, necrotic core; NIRS, near-infrared spectroscopy; VH, virtual histology.
Figure 2  Baseline morphometric greyscale-intravascular ultrasound results of different groups. Lumen area (A), vessel area (B), plaque and media area (C), plaque burden (D), remodelling index (E), and eccentricity index (F) in echolucent plaque (EP), attenuated plaque (AP), calcified plaque (CP) groups were compared with those in the control plaque (CTRL) group.

Figure 3  Relationship between greyscale-intravascular ultrasound plaque types and virtual histology findings. (A–E) Mean percentage of the four virtual histology-intravascular ultrasound plaque components, including necrotic core (NC, A), fibrofatty (FF, B), fibrous tissue (FT, C), and dense calcium (DC, D) as well as the maximum %NC (Max NC%, E) in echolucent plaque (EP), attenuated plaque (AP), calcified plaque (CP) groups were compared with those in control plaque (CTRL) group. (F) Percentages of virtual histology-derived lesion phenotypes in EP, AP, and CTRL groups.
Figure 4  Representative greyscale-intravascular ultrasound and corresponding virtual histology-intravascular ultrasound images. (A) Echolucent plaque (EP) appears as ‘black area’ (arrow) in the greyscale-intravascular ultrasound image. The corresponding virtual histology-intravascular ultrasound shows mixture of fibrofatty (light green) and fibrous tissue (dark green) (white triangle) surrounded by necrotic core (red) indicating thin-cap fibroatheroma. Note that the necrotic core surrounds, but is not part, of the echolucent zone. (B) Attenuated plaque (AP) shows non-calcified plaque with attenuation (arrow) in the greyscale-intravascular ultrasound image. The corresponding virtual histology-intravascular ultrasound shows confluent necrotic core abutting the lumen, indicating virtual histology-thin-cap fibroatheroma. (C) Calcified plaque (CP) shows hyperechoic plaque with acoustic shadow (arrow). The corresponding virtual histology-intravascular ultrasound shows thick calcified cap with confluent necrotic core behind dense calcium (white), indicating calcified thick-cap FA.
Although 75.0% of echolucent plaques were classified as VH-FAs, VH-NC was seen surrounding an echolucent zone, but not within any echolucent zone (Figure 4); and echolucent zones themselves contained mostly FT (6.2%), FF (37.5%), or a combination (56.3%). Echolucent plaques had significantly more VH-derived TCFAs (56.3 vs. 20.0%, \( P = 0.023 \)) and a slightly higher maximum %NC [26.8% (15.9–34.8%) vs. 20.7% (14.3–25.6%), \( P = 0.090 \)] when compared with control plaques.

On NIRS analysis, 75.0% of echolucent plaques contained lipid core; however, because NIRS did not provide structural information, it was not clear whether NIRS-detected lipid was within or surrounding the area that appeared echolucent on greyscale-IVUS imaging. Echolucent plaques had significantly higher NIRS-derived LCBI (\( P = 0.008 \)) and mean angle (\( P = 0.009 \)) when compared with control plaques.

On NIRS analysis, 75.0% of echolucent plaques contained lipid core; however, because NIRS did not provide structural information, it was not clear whether NIRS-detected lipid was within or surrounding the area that appeared echolucent on greyscale-IVUS imaging. Echolucent plaques had significantly higher NIRS-derived LCBI (\( P = 0.008 \)) and mean angle (\( P = 0.009 \)) when compared with control plaques (Figure 6).

**Calcified plaques**

VH analysis showed that in the calcified plaque group, all confluent DC had confluent ‘NC’ (range 16.0–41.2%) coexisting with calcium (Figure 4) with a significant positive correlation between the volume of DC vs. %NC (Rho = 0.91, \( P < 0.001 \)), %DC vs. %NC (Rho = 0.66, \( P < 0.001 \)), or maximum %DC vs. maximum %NC (Rho = 0.70, \( P < 0.001 \)). The %NC of calcified plaques was greater than that of control plaques (Figure 3, \( P < 0.001 \)). All 41 greyscale-IVUS calcified plaques (arc > 90°) were classified as calcified VH-FA with NC that measured 26.0% (IQR 22.1–31.3%). On NIRS analysis, we found that 58.5% of them contained lipid core. Lipid core burden index of calcified plaques was similar to that of control plaques (Figure 6).

**Virtual histology-intravascular ultrasound vs. near-infrared spectroscopy findings**

Since both VH-IVUS and NIRS were developed to detect lipidic necrotic regions within atherosclerotic plaques, we further compared VH-IVUS-derived vs. NIRS-derived parameters. Overall, the relation between VH-derived %NC vs. NIRS-derived LCBI was not significant (Rho = 0.16, \( P = 0.110 \)). However, when lesions were separated according to greyscale-IVUS morphology,
a significant positive relationship was found between VH-derived %NC vs. NIRS-derived LCBI (Rho = 0.50, P = 0.006) and between VH-derived maximum %NC and NIRS-derived maximum 2 mm LCBI (Rho = 0.40, P = 0.037) in attenuated plaques. Within the echolucent plaque group, there was a borderline correlation between VH-derived %NC vs. NIRS-derived LCBI (Rho = 0.42, P = 0.076) and a significant positive correlation between VH-derived maximum %NC vs. NIRS-derived maximum 2 mm LCBI (Rho = 0.49, P = 0.048). We did not find any correlations between VH-derived %NC vs. NIRS-derived LCBI (Rho = −0.18, P = 0.316) or between maximum %NC vs. maximum 2 mm LCBI (Rho = −0.06, P = 0.681) within the calcified plaque group. The correlation between VH-IVUS %NC vs. NIRS-derived LCBI in overall non-calcified plaques was significant (Rho = 0.51, P = 0.001) (Figure 8). Similar results were seen when comparing the mean and maximum angles of the NC (VH-IVUS) and lipid core (NIRS) (data not shown).

**Plaque characteristics in acute coronary syndrome vs. non-acute coronary syndrome patients and among the three coronary arteries**

Echolucent plaques (18.5 vs. 7.8%, P = 0.065) and attenuated plaques (34.6 vs. 18.4%, P = 0.033) were more common in ACS than in stable angina, with no difference in the frequency of calcified plaques between ACS and stable angina (31.5 vs. 31.2%, P =
The percentage of NC [23.9% (21–30.4%) vs. 20.1% (11.3–29.1%), P = 0.028] was higher in ACS than in stable angina, with no difference in the percentage of FT [48.5% (42.0–57.4%) vs. 49.5% (40.4–64.6%), P = 0.243], FF [13.2% (7.1–20.7%) vs. 9.9% (6.9–15.6%), P = 0.081], and DC [9.0% (2.8–20%) vs. 11.7% (6.5–19.6%), P = 0.115] between ACS and stable angina. Near-infrared spectroscopy-derived LCBI [131.6 (31.9–278.8) vs. 87.0 (21.4–152.2), P = 0.020], maximum 2 mm LCBI [204.9 (57.5–400.9) vs.135.9 (67.8–249.3), P = 0.056], and mean angle [57.0° (15.9–102.6°) vs. 33.8° (13.5–64.9°), P = 0.018] and maximum angle [94.2° (32–160°) vs. 65.5° (35.3–101.3°), P = 0.042] of lipid core were higher in ACS than in stable angina.

There was only a weak trend for a correlation between VH-derived %NC and NIRS-derived LCBI when analysis was performed in ACS patients (Rho = 0.24, P = 0.097), but there was no correlation in stable angina patients (Rho = 0.08, P = 0.270). However, the correlation between VH-IVUS %NC vs. NIRS-derived LCBI in non-calcified plaques was significant in both ACS (Rho = 0.56, P = 0.001) and stable angina subgroups (Rho = 0.41, P = 0.006). The correlation between VH-IVUS %NC vs. NIRS-derived LCBI disappeared in calcified plaques in both ACS (Rho = −0.22, P = 0.534) and stable angina subgroups (Rho = −0.08, P = 0.232). Similarly, when data were analysed by artery, the correlation between VH-IVUS %NC and NIRS-derived LCBI was significant in non-calcified plaques in the LAD (Rho = 0.56, P = 0.001), LCX (Rho = 0.50, P = 0.015) and RCA (Rho = 0.45, P = 0.008), but was not significant in calcified plaques in the LAD (Rho = −0.26, P = 0.102), LCX (Rho = −0.15, P = 0.583) and RCA (Rho = −0.08, P = 0.642).

**Reproducibility of plaque type classification**

Interobserver variability for plaque type classification was validated in 101 lesions from 51 randomly selected vessels. Blinded analyses were repeated by the first observer after an interval of 12 weeks.
components such as smooth muscle cells in other studies.\textsuperscript{16,17} 

Intraobserver variability yielded good concordance for echolucent plaque (\(k = 0.772\), 95% CI 0.580–0.964); attenuated plaque (\(k = 0.914\), 95% CI 0.865–0.963); calcified plaque (\(k = 0.960\), 95% CI 0.932–0.988); and VH-IVUS phenotype [VH-TCFA (\(k = 0.883\), 95% CI 0.703–0.961), ThCFA (\(k = 0.840\), 95% CI 0.734–0.946), and PIT (\(k = 0.935\), 95% CI 0.890–0.980)]. Intraobserver variability yielded good concordance for echolucent plaque (\(k = 0.772\), 95% CI 0.580–0.964); attenuated plaque (\(k = 0.914\), 95% CI 0.865–0.963); calcified plaque (\(k = 0.960\), 95% CI 0.932–0.988); and VH-IVUS phenotype [VH-TCFA (\(k = 0.883\), 95% CI 0.703–0.961), ThCFA (\(k = 0.840\), 95% CI 0.734–0.946), and PIT (\(k = 0.935\), 95% CI 0.890–0.980)]. 

**Discussion**

Our current knowledge about coronary plaque characteristics is largely obtained from ex vivo postmortem studies of lesions at the almost end of the coronary disease spectrum. In this in vivo comparison of tissue characterization of human coronary plaques using NIRS and greyscale and VH-IVUS, important findings were: (i) greyscale-IVUS-detected attenuated and echolucent plaques indicated the presence of NIRS-detected lipid core; (ii) most (93.5%) attenuated plaques contained confluent NC and were classified as VH-derived FA; (iii) although 75.0% of echolucent plaques were classified as VH-FAs, VH-NC was seen surrounding an echolucent zone, but not within any echolucent zone; (iv) all calcified plaques (arc >90°) contained >10% NC and were classified as VH-FA, whereas 58.5% contained NIRS-detected lipid core; (v) A positive relationship between VH-derived %NC and NIRS-derived LCBI was found in non-calcified plaques but not in calcified plaques.

**Echolucent and attenuated plaques**

Although conventional greyscale-IVUS is widely used during interventional procedures, several special greyscale-IVUS images—i.e. echolucent plaques and attenuated plaques—are still incompletely understood. Several small in vitro studies of echolucent and attenuated plaques yielded conflicting results.\textsuperscript{15–19} For example, echolucent components detected by greyscale-IVUS have been related to high lipid content in some histological studies,\textsuperscript{12,15} but to non-lipid components such as smooth muscle cells in other studies.\textsuperscript{16,17} Attenuated plaque has also been variously related to cholesterol clefts, microcalcification, or organized thrombus.\textsuperscript{18–20} In our study, tissue characterization by NIRS offered an in vivo opportunity to assess the chemical composition of echolucent and attenuated plaques. The NIRS findings suggested that these greyscale-IVUS characteristics were related to lipid content. In the NIRS ‘block chemogram’, the probability of lipid core was highest in attenuated plaques, followed by echolucent plaques.

These in vivo findings have clinical implications. Ultrasonic attenuation from non-calcified plaques (attenuated plaques) may indicate a large lipid/NC, whereas an intraplaque echolucent zone (echolucent plaques) may indicate a smaller lipid/NC.

**Assessment of lipidic/necrotic core**

A positive relationship between the NIRS-derived LCBI and VH-derived %NC was found in non-calcified plaques, but not in calcified plaques. The accuracy of VH-IVUS assessment of plaque composition behind calcium, especially NC, remains actively debated.\textsuperscript{20–22} Highly calcified lesions might be an anatomical limitation to IVUS radiofrequency data analysis. It is unclear how often the VH-derived signals behind calcium are mostly noise and how often they contain useful data. Several investigators have shown that VH-IVUS of calcium can include a surrounding red halo that is part of the calculation of lesion NC, but that is most probably an artefact as has been suggested by Shin et al.\textsuperscript{23,24}

Our results suggested that VH-IVUS might overestimate NC in the presence of heavy calcium. First, all calcified plaques (arc >90°) had >10% confluent VH-derived NC (range 16.0–41.2%) coexisting with calcium and were classified as VH-FAs. Second, significant and strong positive correlations between all parameters of DC and NC were found in the calcified plaque group, but not in other groups. Pathological studies have shown that not all calcified plaques contain NC. Calcium could deposit either within or around a necrotic region, resulting in the formation of a calcified FA (FA with calcified core); alternatively, calcium could deposit at sites of fibrous collagenous tissue, resulting in the formation of calcified fibrous plaque (calcified plaque without adjacent lipid/necrosis).\textsuperscript{4,13,25} Absolute or relative amounts of VH-derived NC have been used as a risk stratification indicator for patients with coronary artery disease.\textsuperscript{26} Our results suggested that caution should be taken in the VH-IVUS assessment of NC in vessels with heavy calcification, as has been demonstrated by a study using stent metal to simulate the addition of calcium to a lesion.\textsuperscript{23}

VH-IVUS overestimation of NC content in the presence of calcification could also explain the discrepancy between VH-IVUS and NIRS findings in our study. Similar mean and maximum percentages of NC were found by VH-IVUS, but significantly lower mean and maximum LCBI values were shown by NIRS in calcified plaques compared with attenuated plaques. In addition, there was no correlation between VH-derived %NC and NIRS-derived LCBI in calcified plaques. Near-infrared spectroscopy, unlike ultrasound, assesses chemical composition based on near-infrared light absorption and scatter. Necropsy validation data have demonstrated that NIRS can differentiate calcified plaques containing a lipid core, from calcified fibrotic plaques that do not contain a lipid core.\textsuperscript{4}

In the current *in vivo* study, 58.5% of greyscale-IVUS calcified plaques contained NIRS-derived lipid core while all of them had >10% VH-IVUS NC. A study performed at almost the same time as the current study also compared NIRS and VH and showed that the overall correlation between the relative VH-NC content and the values of the NIRS block chemogram was weak,\textsuperscript{27} similar to the current study. However, excluding calcified lesions from the analysis did not improve, but rather worsened the correlation. Several important differences between two studies might contribute to this disagreement. First, the ‘plaque’ in the study by Brugaletta et al.\textsuperscript{27} was defined according to the NIRS chemogram blocks, not greyscale-IVUS. Second, the ‘calcified plaque’ in the study by Brugaletta et al.\textsuperscript{27} was defined as >10% of average VH-DC content for a given VH segment matched to a
2 mm chemogram block and not based on the greyscale-IVUS criteria as in the current study. Third, in the study by Brugaletta et al., NIRS was compared with the VH-derived %NC using the ‘chemogram block probability’ as defined as the probability of LCP displayed in the chemogram block rather than using the LCBI. The NIRS ‘block chemogram’ provides only a summary of the data in every 2 mm segment and does not indicate individual pixel data or the location of a measurement in the circumferential dimension, and, therefore, correlating ‘chemogram block probability’ to the VH-derived %NC might not be the optimal approach.

Clinical implications
In the present study, we showed that echoluent and attenuated plaques were more frequent in ACS than in stable angina, with no difference in the frequency of calcified plaques between ACS and stable angina. Tissue characterization by NIRS and VH-IVUS in the present study suggested that echoluent and attenuated plaques were representative of lipid/NC-containing plaques. In previous studies, although the mechanisms were unclear, both echoluent plaques and attenuated plaques have been associated with distal embolization during PCI, especially PCI of patients presenting with ACS. Similarly, in seven studies, VH-derived NC has also been associated with distal embolization (using a variety of endpoints) during PCI, again especially PCI of patients presenting with ACS. Finally, in one published NIRS study, a large amount of lipid plaque has also been associated with distal embolization during PCI, leading to the randomized CANARY(Coronary Assessment by Near-infrared of Atherosclerotic Rupture-prone Yellow) trial comparing distal protection vs. no distal protection in patients with an LCBI >600. The current three-way greyscale-IVUS, VH-IVUS, and NIRS comparison links these observations.

Study limitations
First, although data in our study are prospectively collected and blindly analysed, the sample size (131 plaques in 66 vessels) was relatively small. Second, we did not assess very severe stenoses because NIRS and IVUS were usually performed after pre-dilation of such lesions. Third, we could not exclude the possibility of a mismatch in the analysis segments, leading to inaccuracies in the registration of segments between the different technologies. The development of NIRS/IVUS combination catheter, in part, addressed this issue. Fourth, the VH-IVUS identification of a TCFA is inferential because a thin-fibrous cap is below the resolution of IVUS; however, a VH-TFCF was a predictor of events in PROSPECT. In addition, because the NIRS system used in this study did not provide structural information, we were only able to compare entire cross-sections between spectroscopy and ultrasound and not individual regions of interest. Finally, our findings from the coronary arteries of in vivo patients lack a direct comparison with histopathology. Ex vivo histological studies in human coronary autopsy specimens are needed in order to further confirm such in vivo findings in living patients.

Conclusion
Combining NIRS with IVUS contributes to the understanding of plaque characterization in vivo. An NIRS/IVUS (optical/acoustic) combination catheter, which allows co-registration of coronary anatomy and chemical composition, has been developed and is currently undergoing pre-clinical and early clinical evaluation. Further studies are warranted to determine whether combining NIRS and IVUS techniques will enhance the assessment of high-risk plaque to predict outcomes in patients with coronary artery disease.

Conflict of interest: G.S.M. reports receiving consulting fees from Boston Scientific and Volcano and research/grant support from Boston Scientific, InfraRedx, and Volcano. G.W.S. reports being an advisory board for Boston Scientific and Abbott Vascular, and a consultant for Medtronic, Volcano, and InfraRedx. E.S.B. reports receiving speaker honoraria from St Jude Medical and Terumo, and research support from Abbott Vascular. S.B. reports receiving speaker honoraria from St Jude Medical, Medtronic, and Johnson & Johnson and research support from Boston Scientific and The Medicines Company. B.M. reports being an advisory board for InfraRedx, Abiomed, and Medtronic, and a consultant for Abbott Vascular and St Jude Medical. A.M. reports receiving research grants from Boston Scientific and lecture fee from J.P. reports receiving research grants from Boston Scientific China.

References
Plaque characterization by IVUS and NIRS


