

Single-copy genes as molecular markers for phylogenomic studies in seed plants

- Supplementary Materials -

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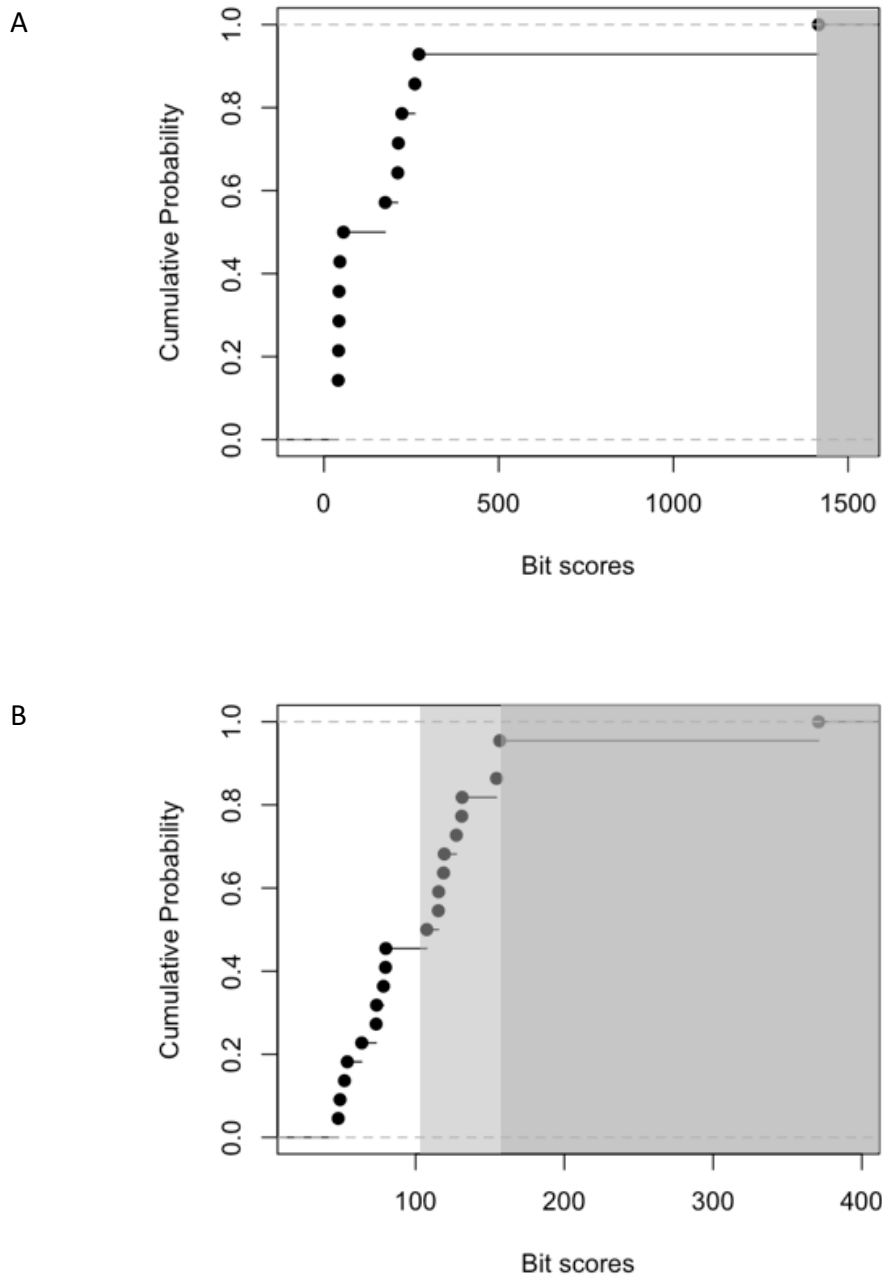
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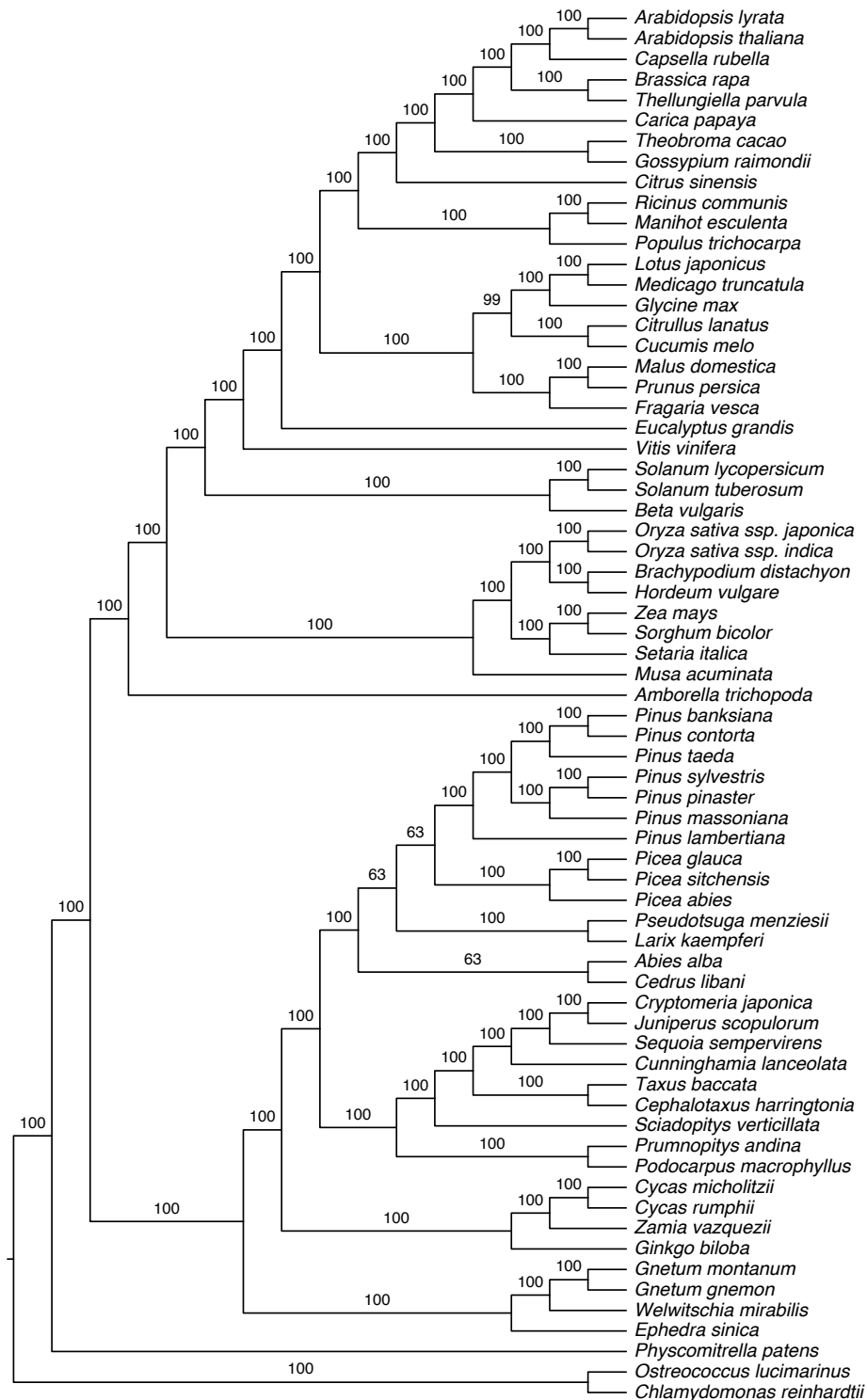
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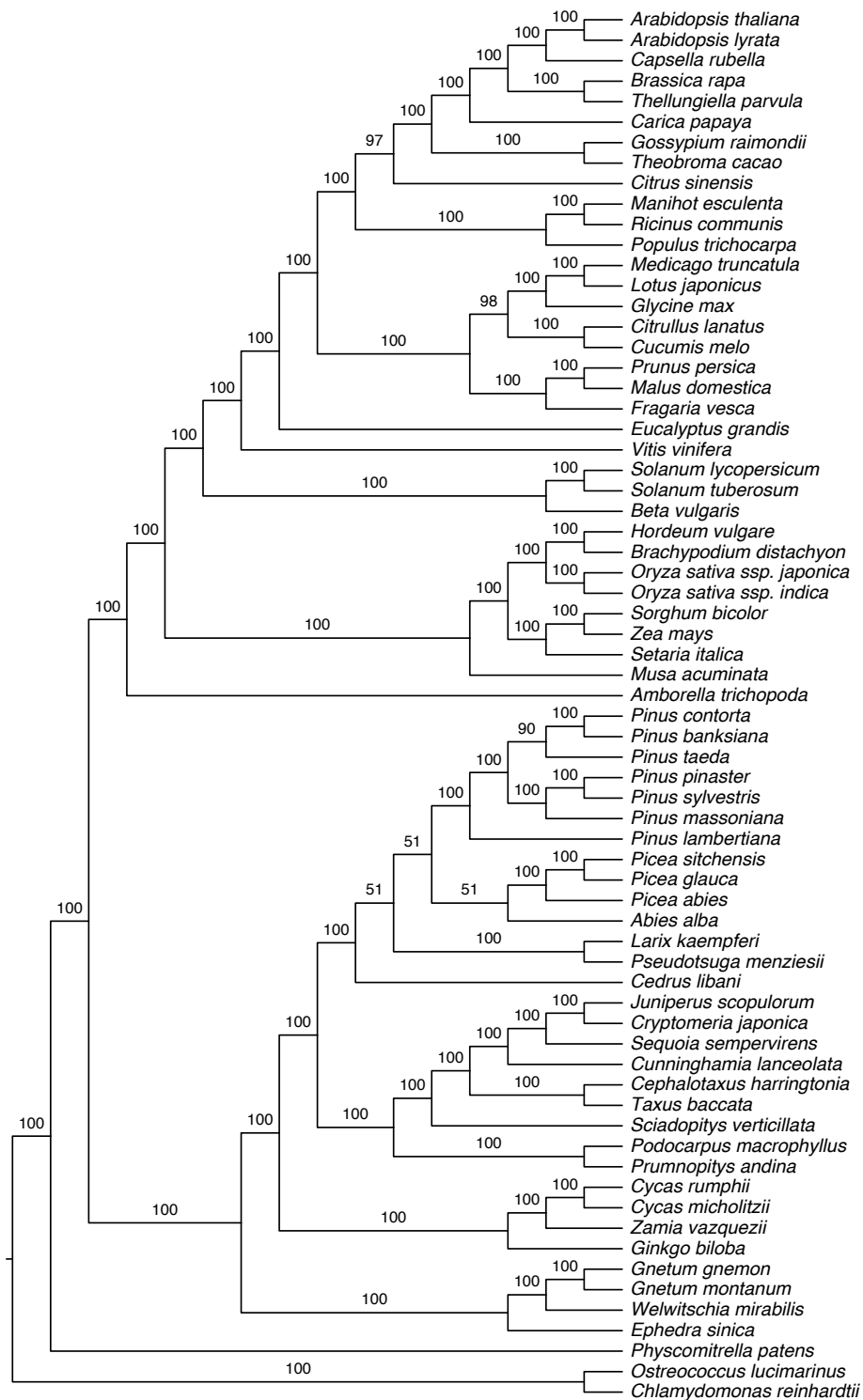
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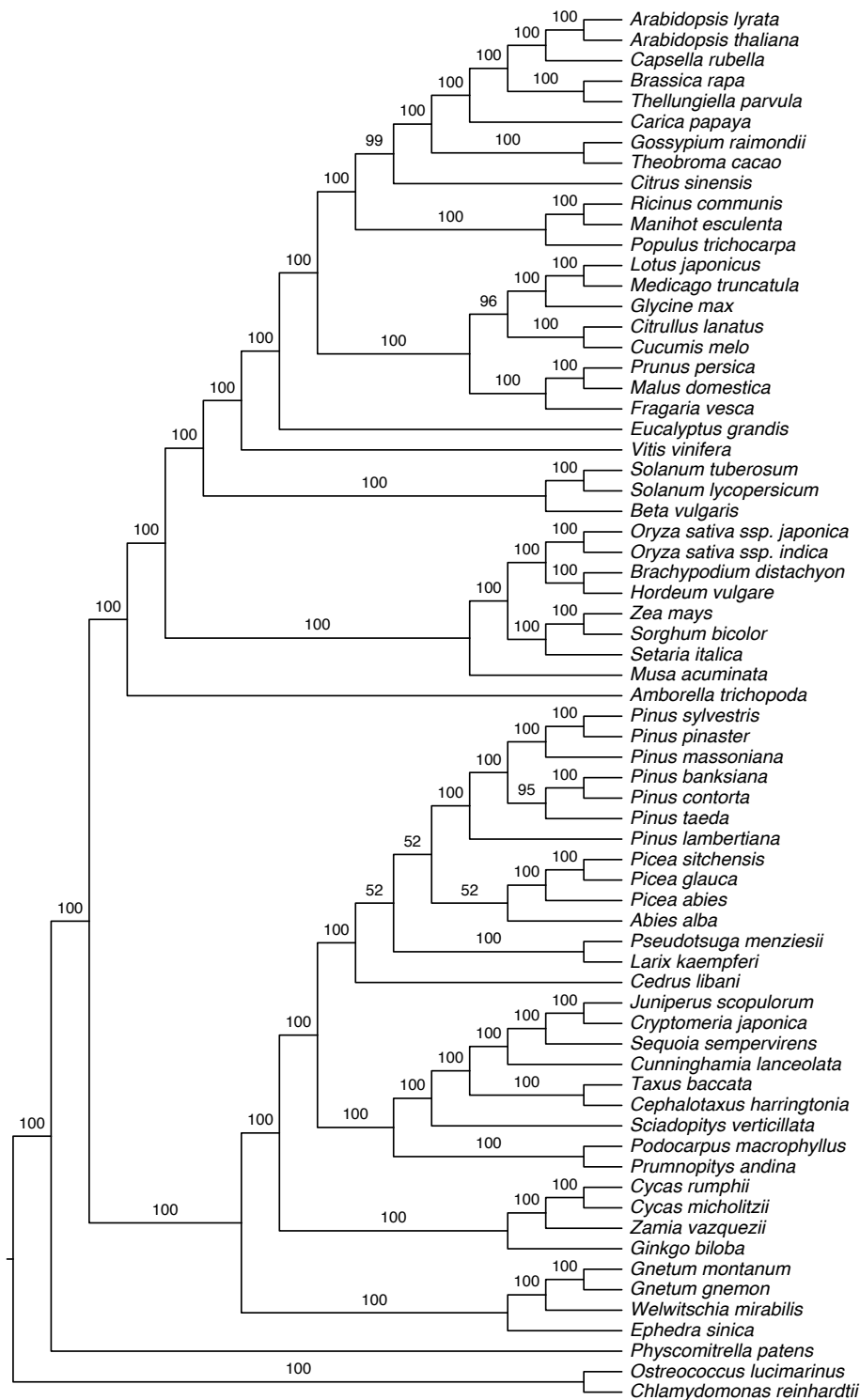
Supplementary Figure 1. Example showing the process to assign genes to a gene family accounting for 95% of the cumulative probability of bit-scores. The dark grey blocks in both A and B show regions with over 95% cumulative probability. Dots (i.e. hits) falling in the dark grey region are allocated to the gene family. In B, the light grey block denotes a region where hits have similar E-values ($\Delta E\text{-value} < 10 \times 10^{20}$) to the hit with the smallest Bit-score at the 95% border.



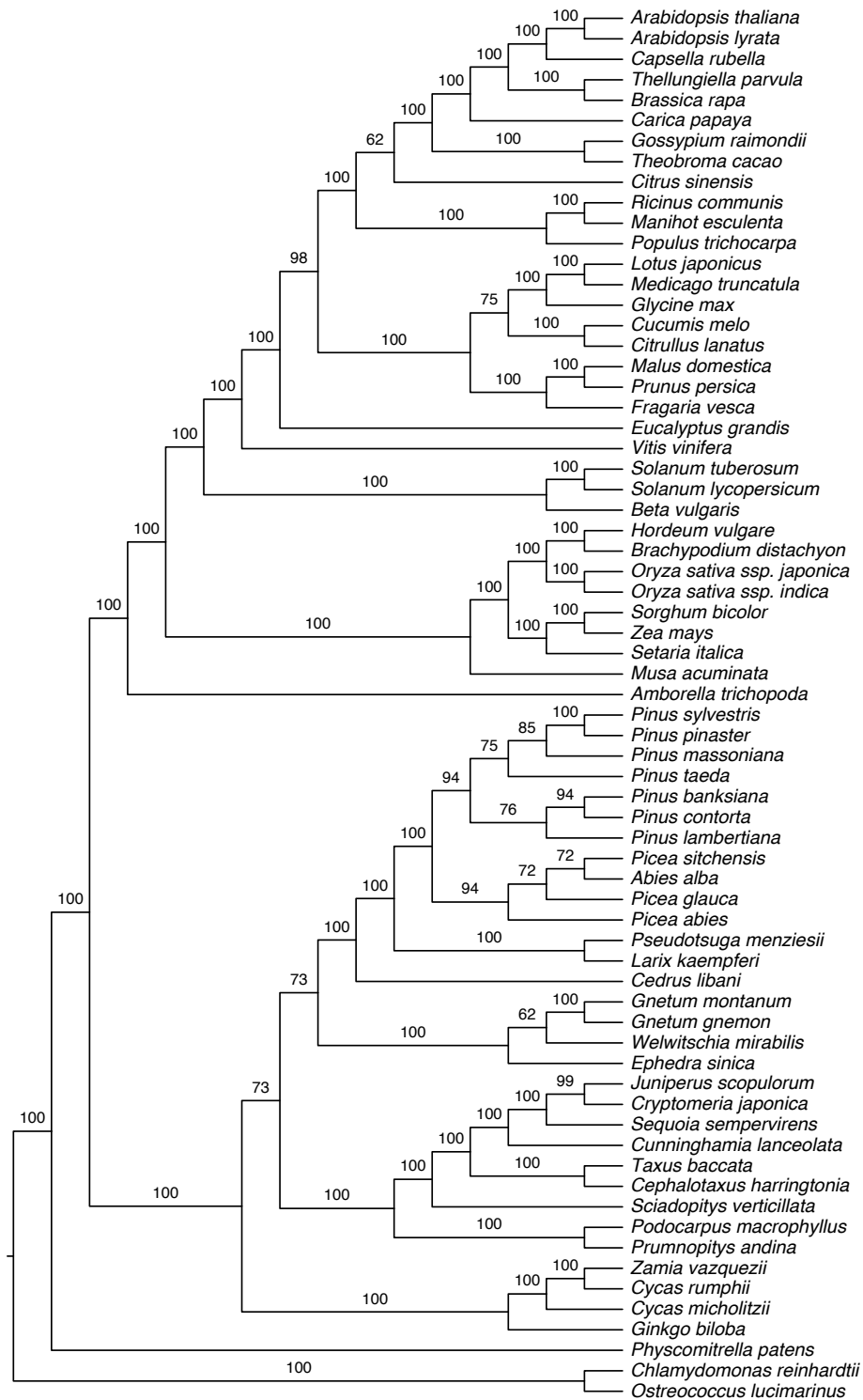
Supplementary Figure 2. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants including 3rd codon positions with one partition. Numbers on branches represent bootstrap values.



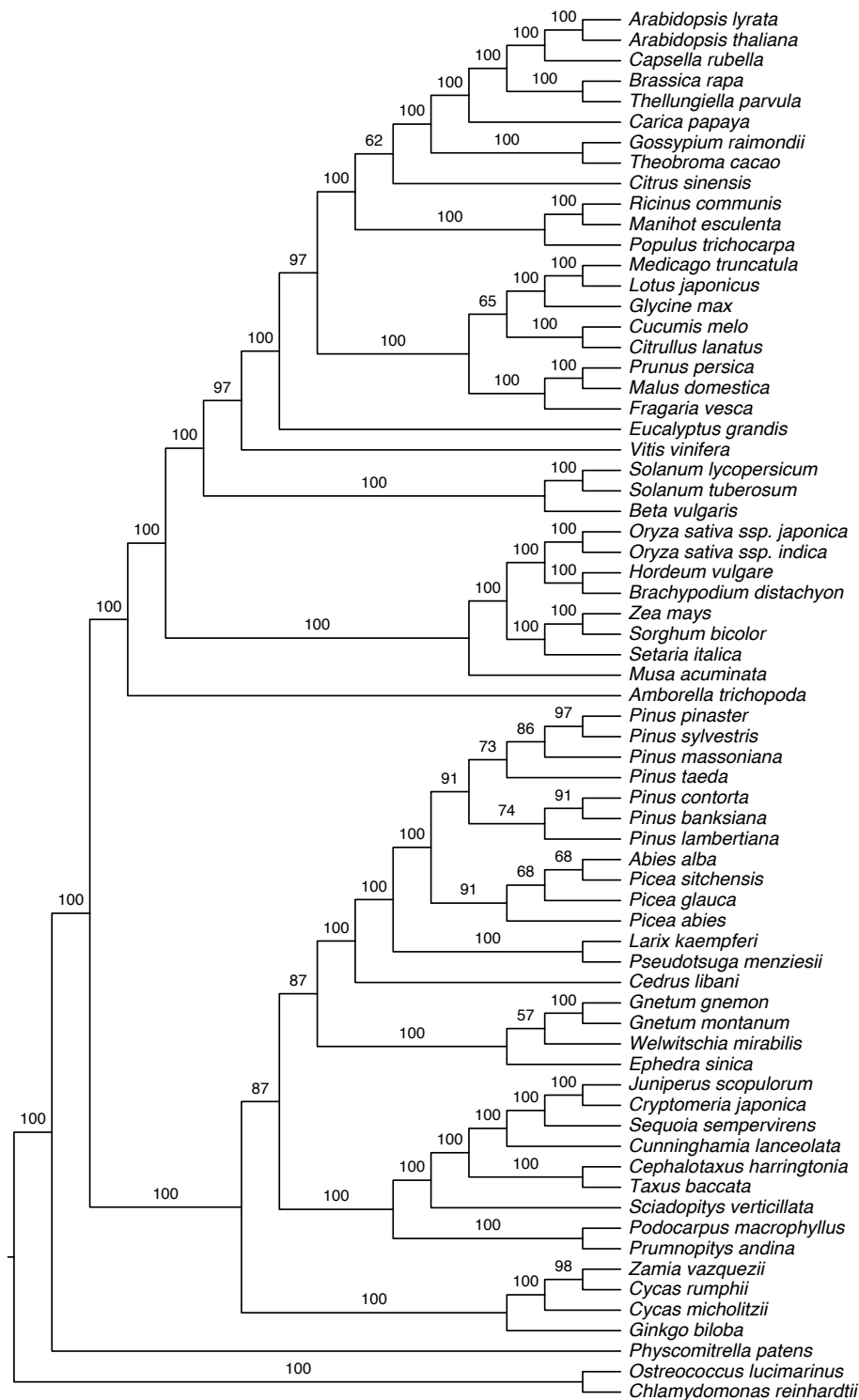
Supplementary Figure 3. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants including 3rd codon positions, but with 1st and 2nd codon partitions as one partition and 3rd codon partition as another. Numbers on branches represent bootstrap values.



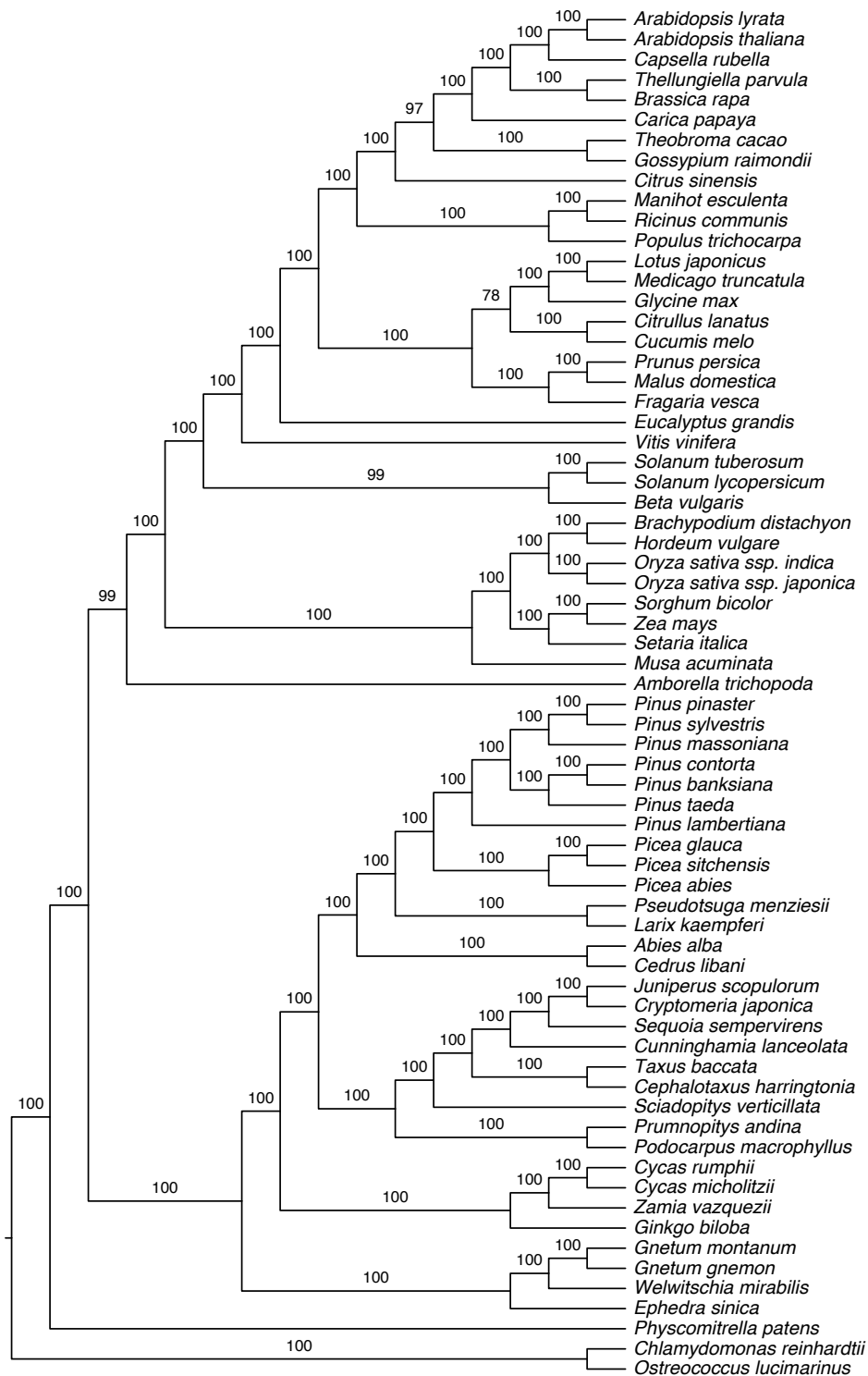
Supplementary Figure 4. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants including 3rd codon positions but with three partitions, i.e. one for each codon position. Numbers on branches represent bootstrap values.



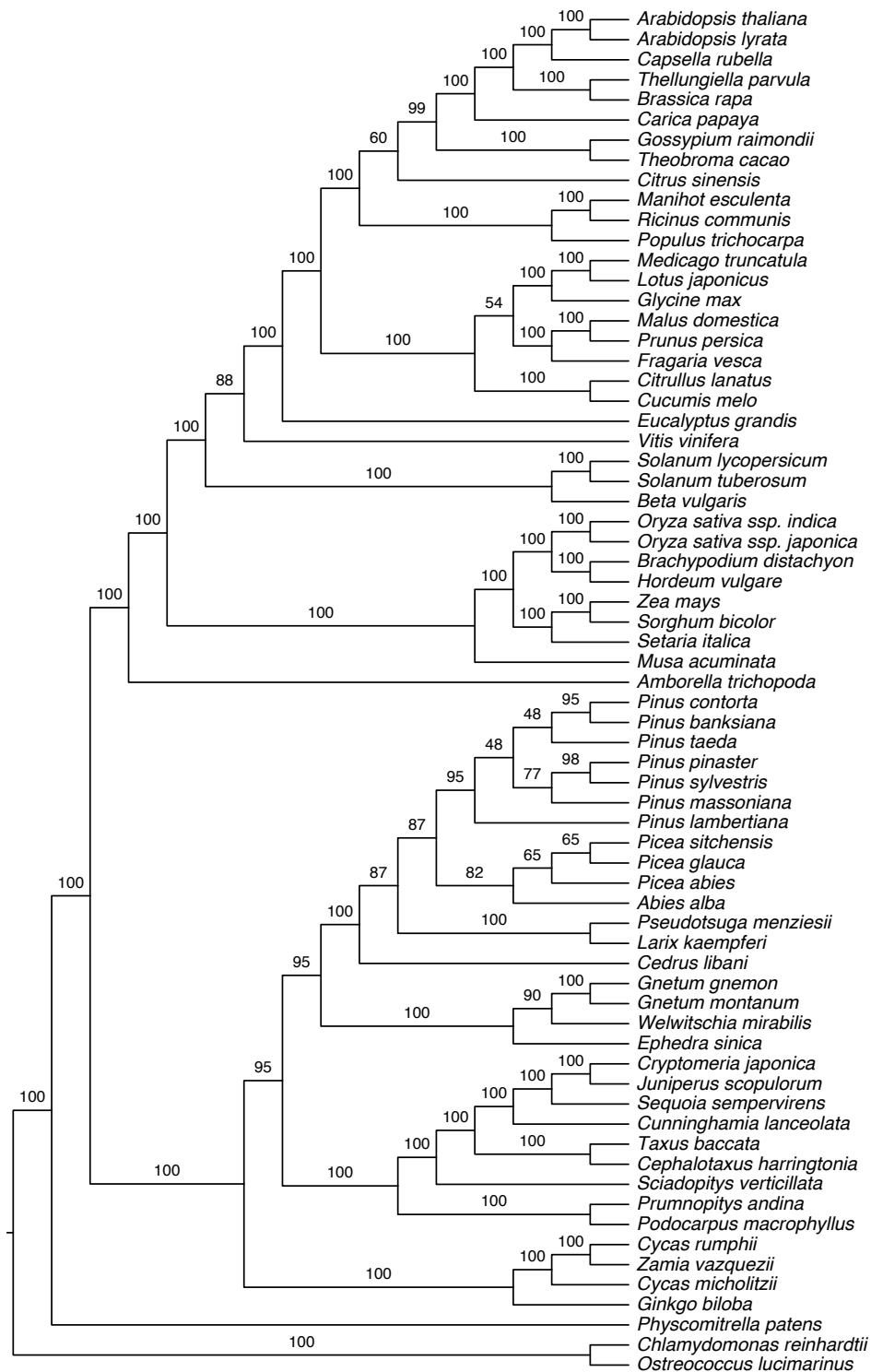
Supplementary Figure 5. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants with 3rd codon positions removed and with position 1 and 2 as one partition. Numbers on branches represent bootstrap values.



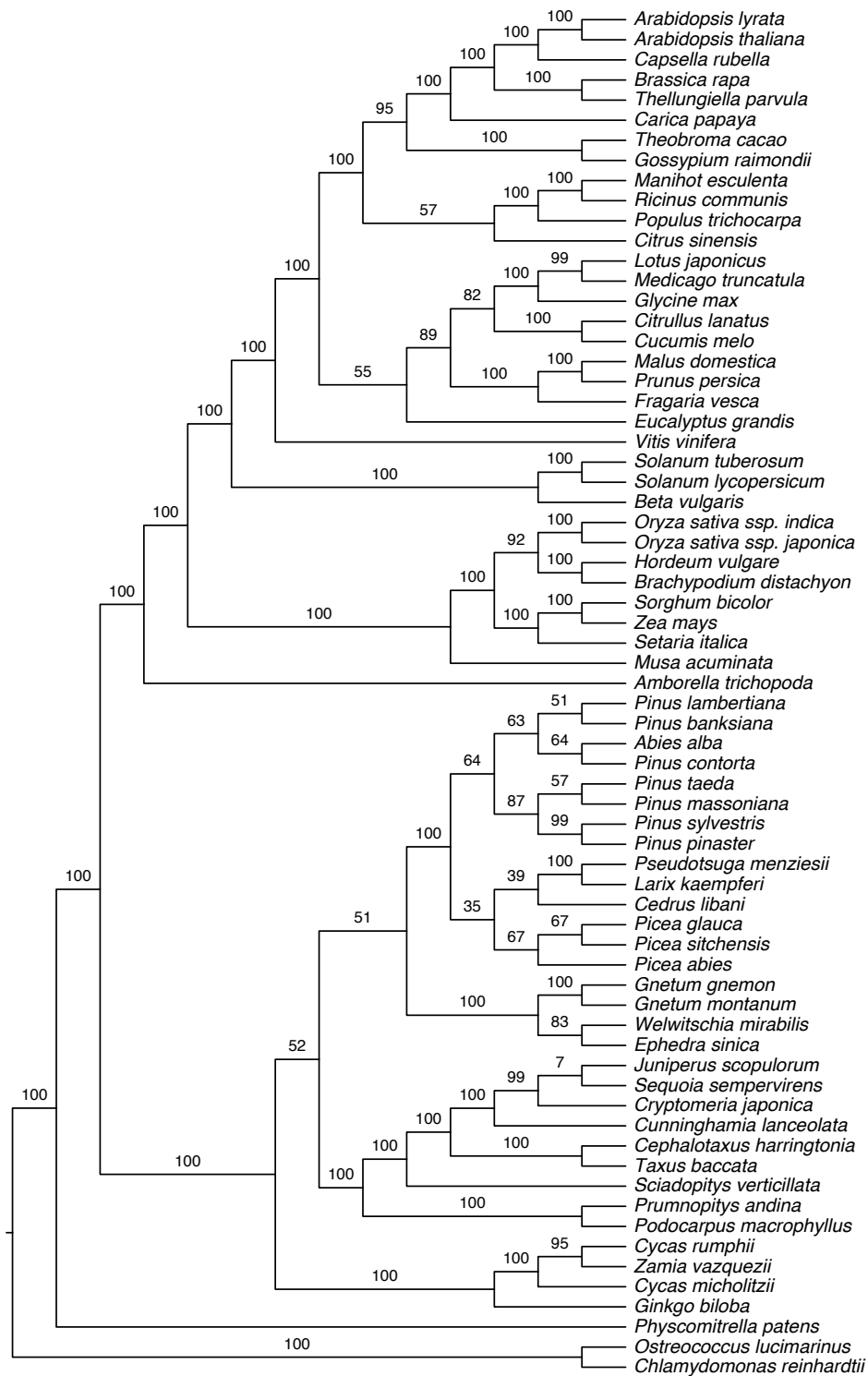
Supplementary Figure 6. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants with 3rd codon positions removed and with position 1 and 2 used as separate partitions. Numbers on branches represent bootstrap values.



Supplementary Figure 7. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants only with 3rd codon positions. Numbers on branches represent bootstrap values.



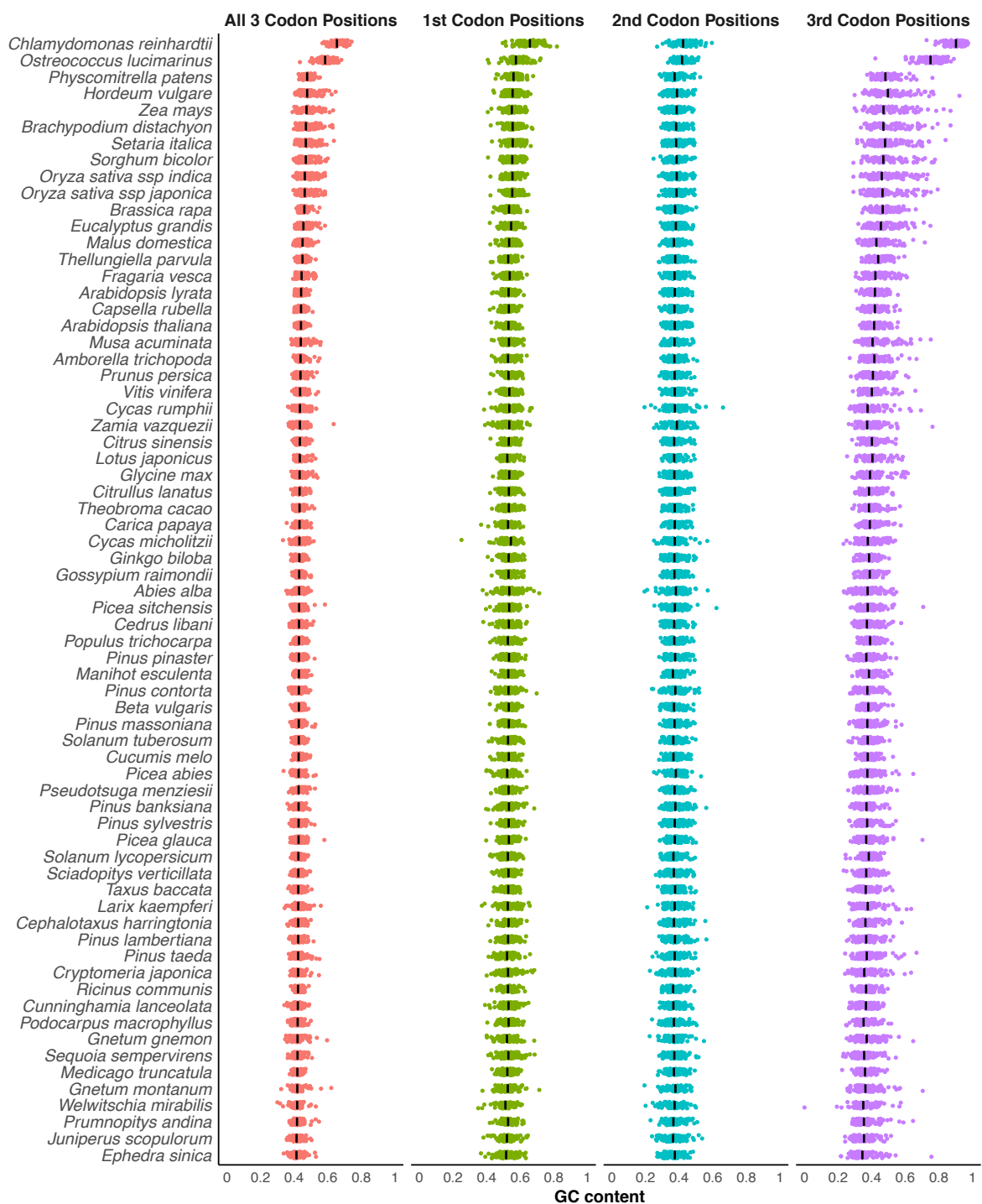
Supplementary Figure 8. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants only with 1st codon positions. Numbers on branches represent bootstrap values.



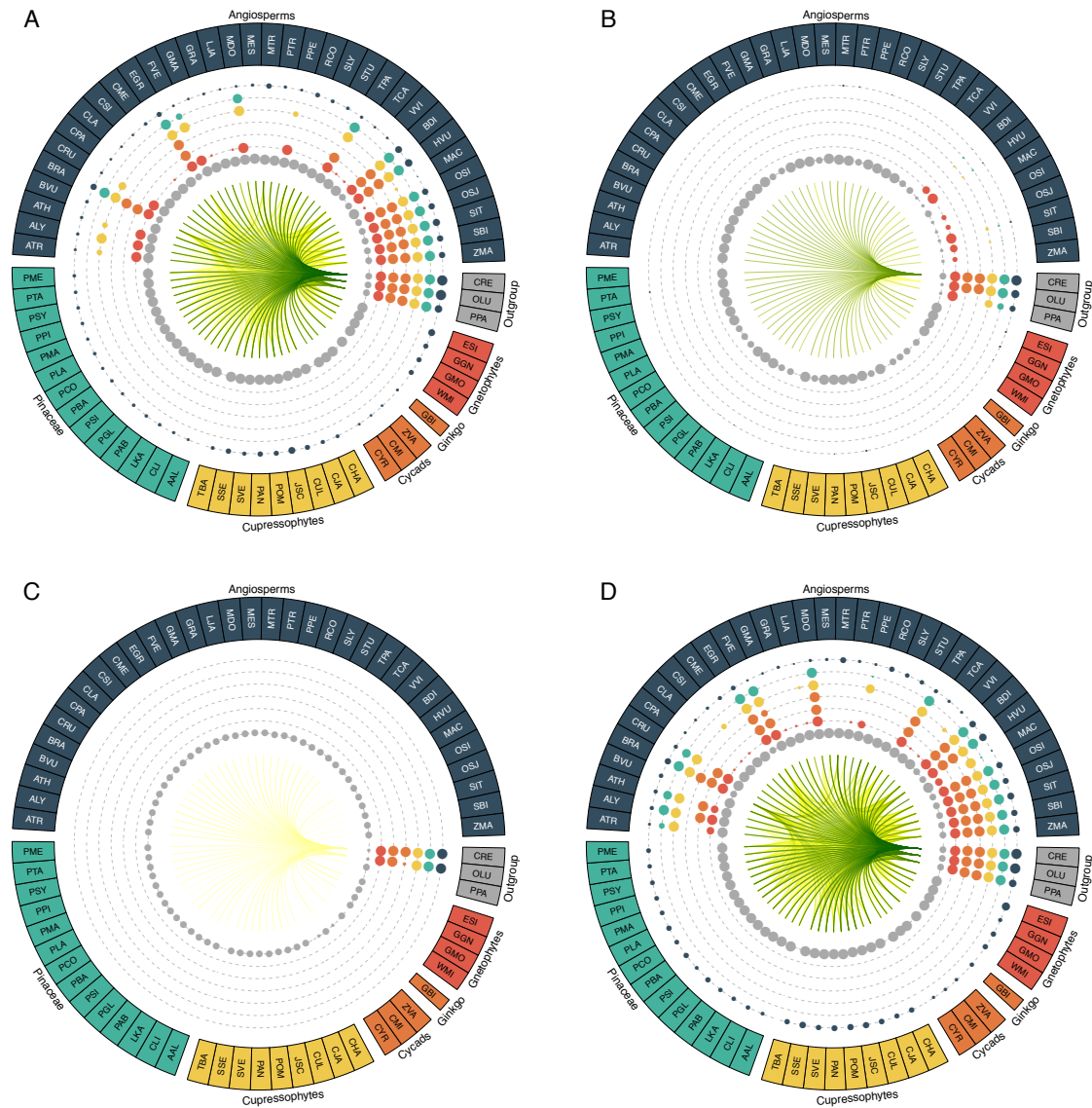
Supplementary Figure 9. Maximum likelihood tree inferred from a concatenated alignment of 106 single-copy genes in seed plants only with 2nd codon positions. Numbers on branches represent bootstrap values.



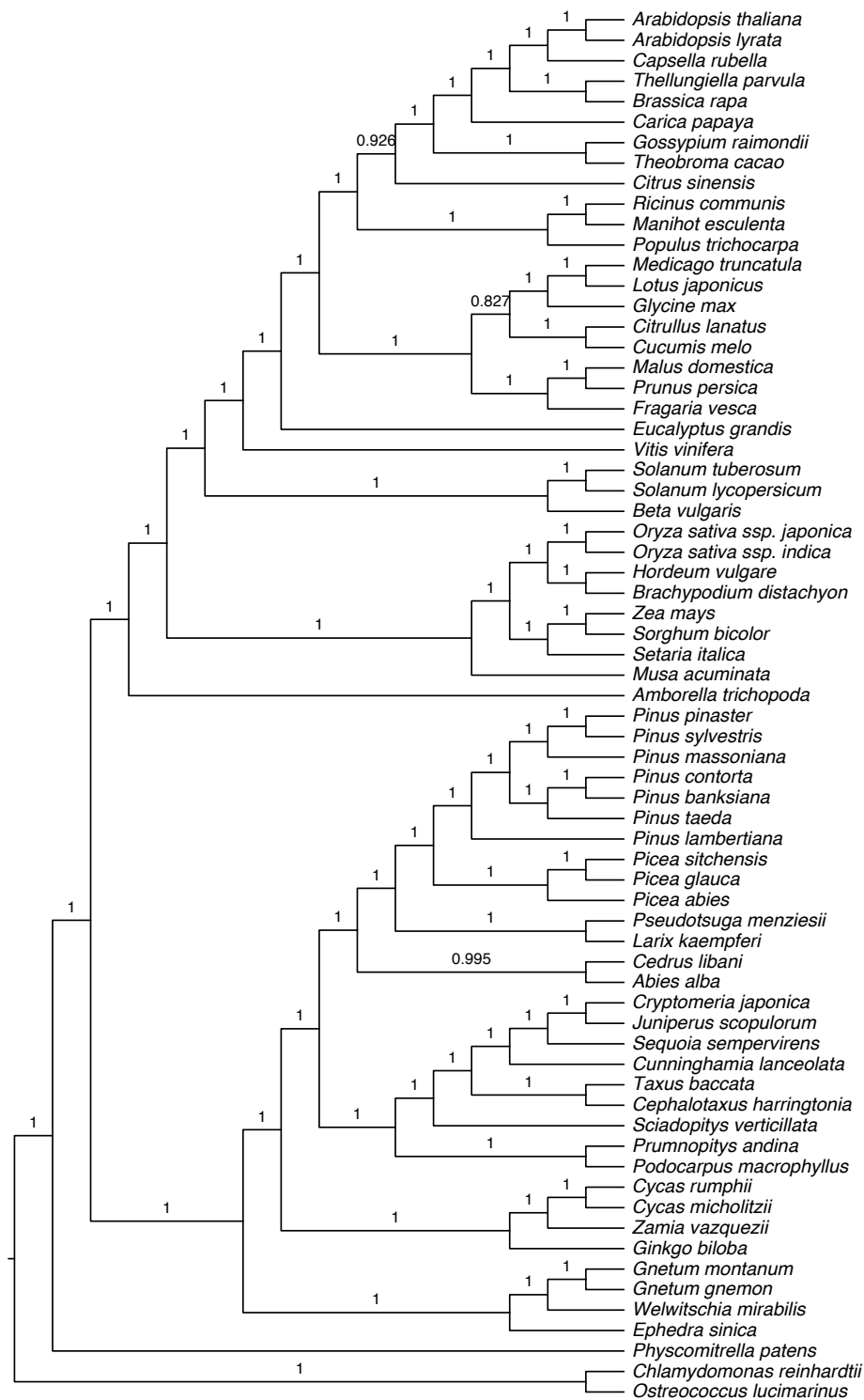
Supplementary Figure 10. GC content of the 106 phylogenetic markers at 1st, 2nd, and 3rd codon positions for the species studied.



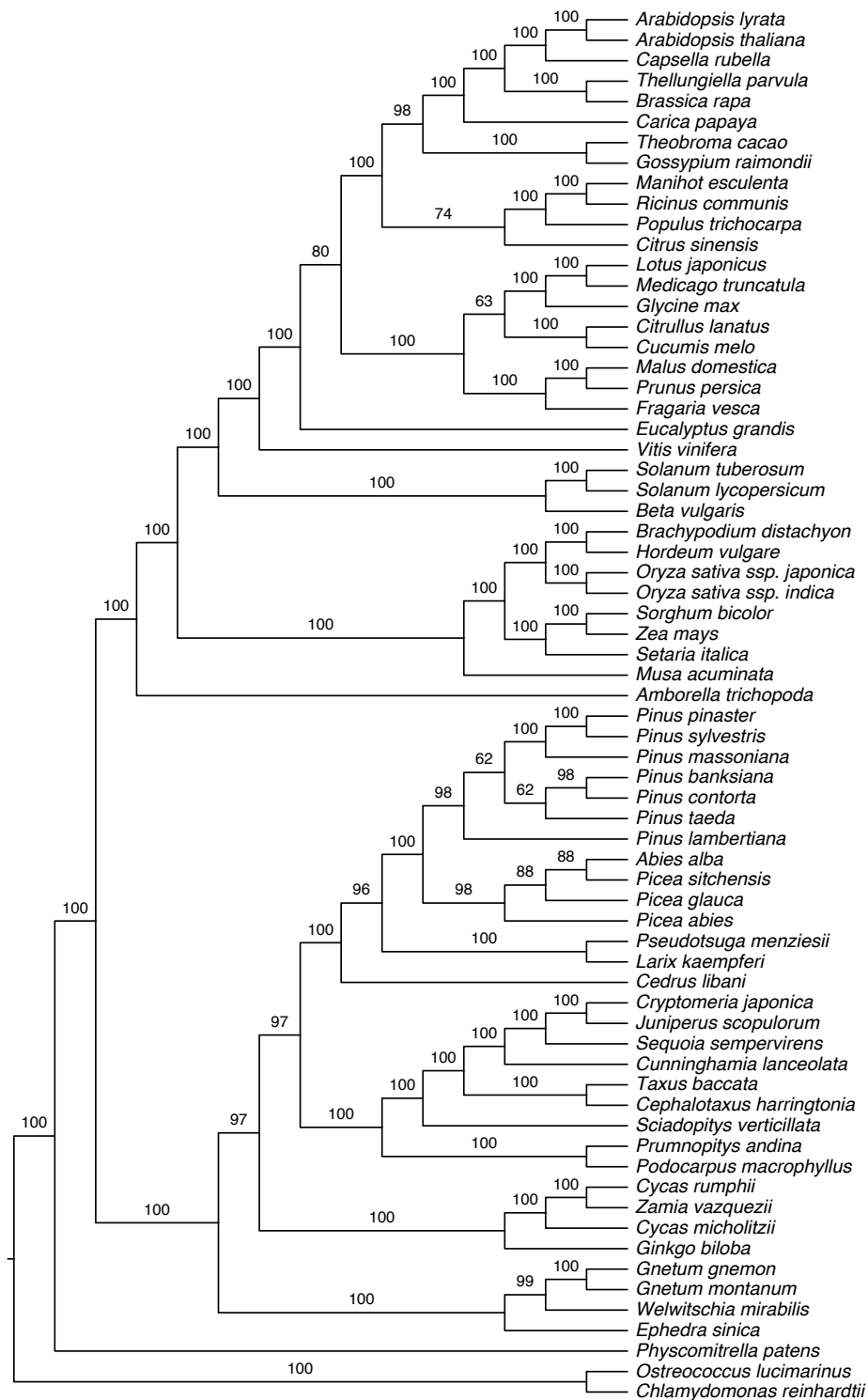
Supplementary Figure 11. GC content of the 106 phylogenetic markers at 1st, 2nd, and 3rd codon positions for the species studied after removing sites that encode the same amino acids.



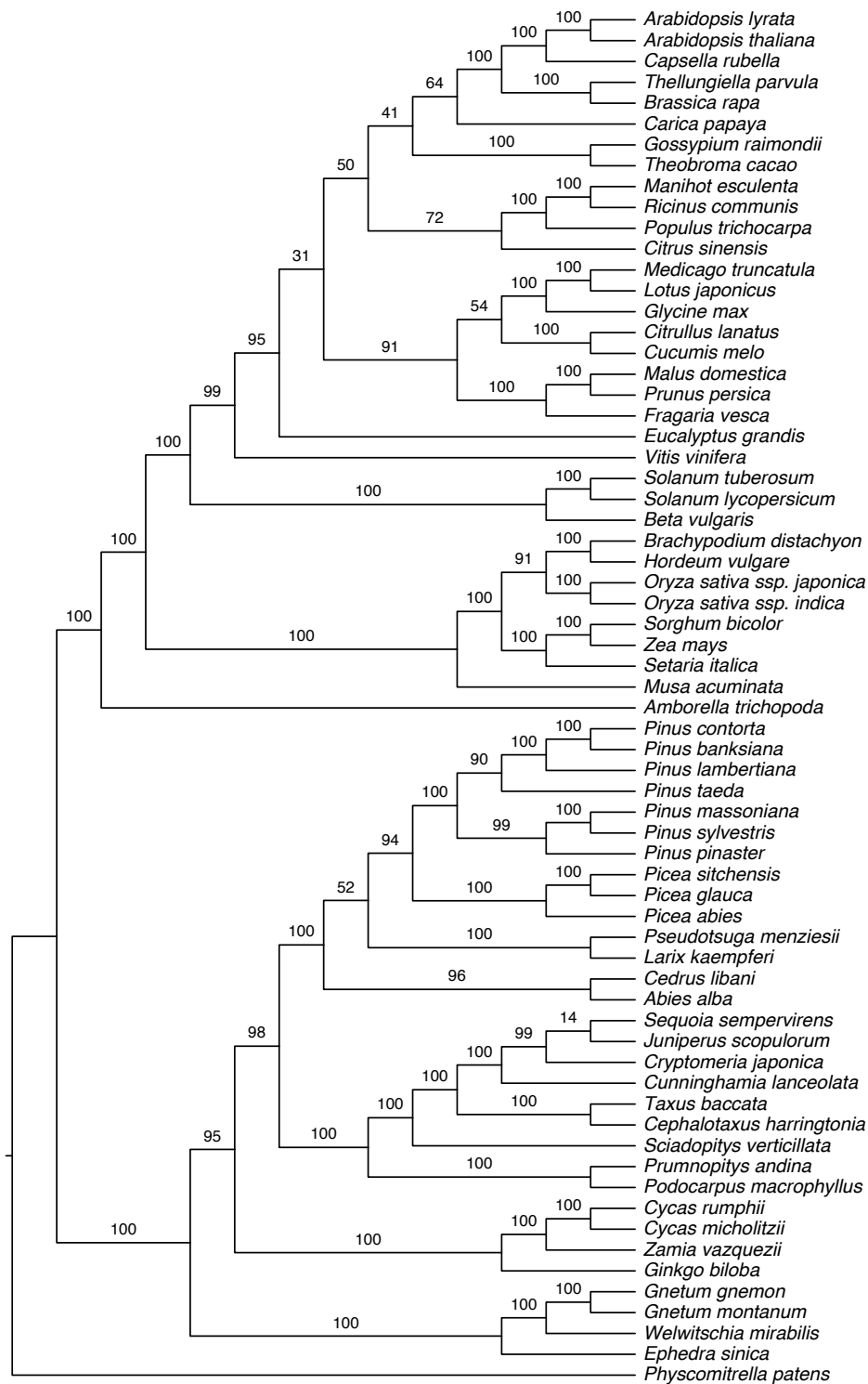
Supplementary Figure 12. Comparison of GC content in the concatenated alignment (A) and at each codon position (B, C, and D) from 106 genes in 68 species after removing sites that encode the same amino acids. Dot size correlates with the number of species in each lineage (group) that have a significantly different GC% (Wilcox test, $P < 1 \times 10^{-3}$) with the species compared with (colors of dots correspond to the compared lineages). Lines connecting any two species represent significant difference in GC content, with most significant in green and weakest in yellow (1×10^{-3}). The full names for the species can be found in Supplementary Table 3.



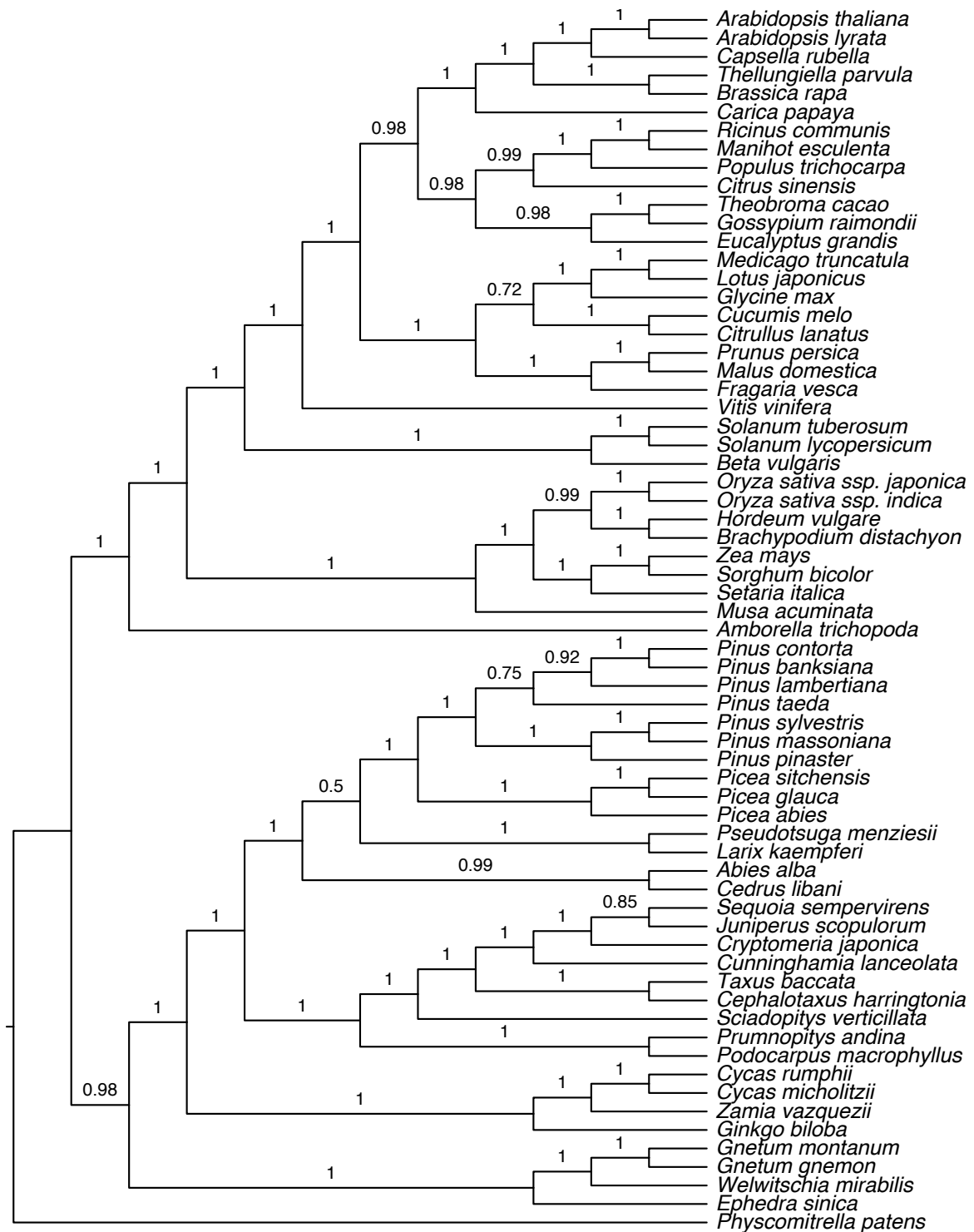
Supplementary Figure 13. Maximum likelihood tree inferred from a concatenated codon alignment of 106 single-copy genes using the Goldman and Yang (GY) model. Numbers on branches represent support values from the SH-like approximate likelihood-ratio test.



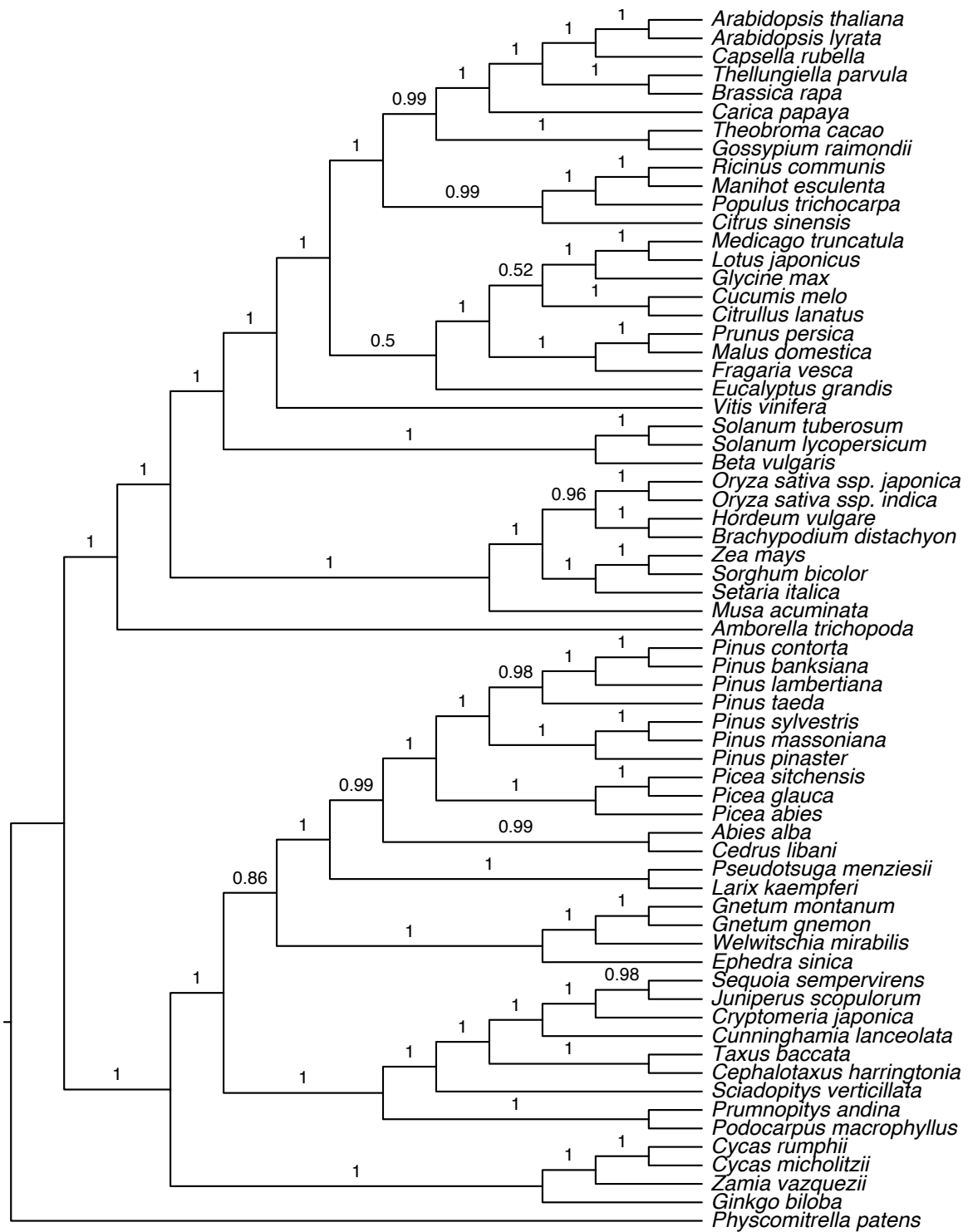
Supplementary Figure 14. Maximum likelihood tree inferred from a concatenated amino acid alignment of 106 single-copy genes using the JTT+I+GAMMA+F model. Numbers on branches represent bootstrap values.



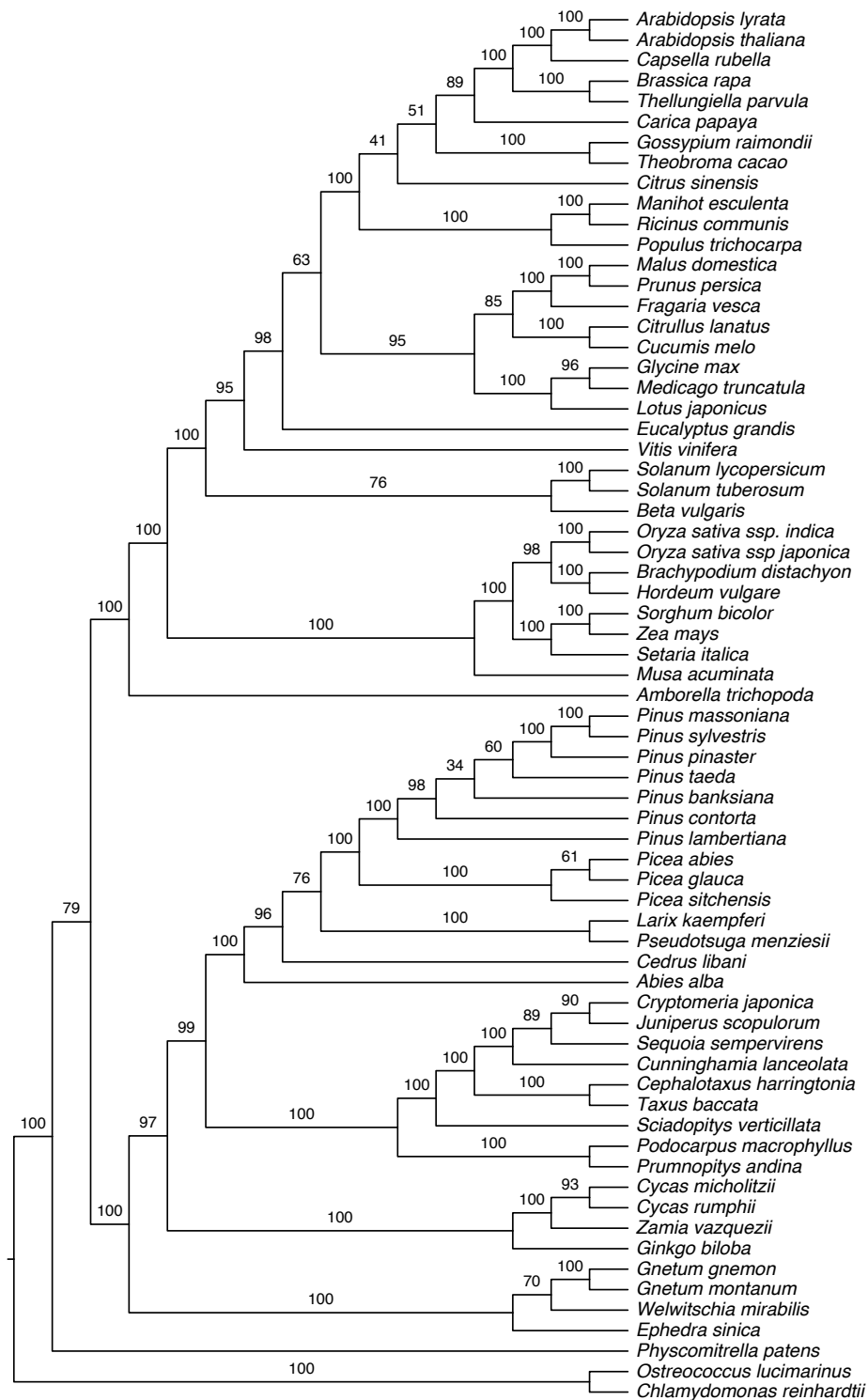
Supplementary Figure 15. Maximum likelihood tree inferred from a reduced concatenated amino acid alignment of 106 single-copy genes using the JTT+I+GAMMA+F model. Numbers on branches represent bootstrap values.



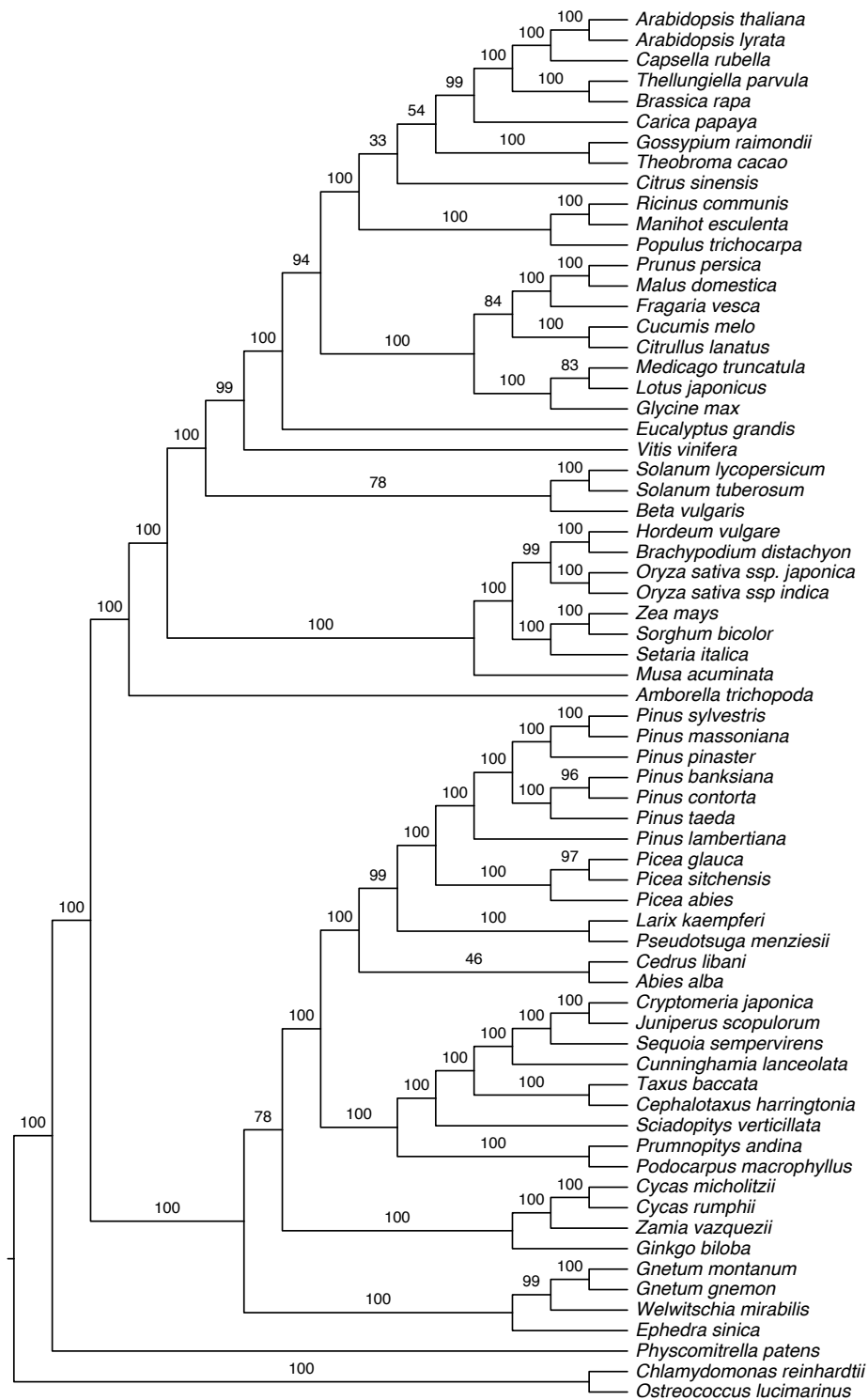
Supplementary Figure 16. Bayesian phylogenetic tree based on the reduced amino acid concatenation of 106 single-copy genes under CAT model. Numbers indicate posterior probabilities.



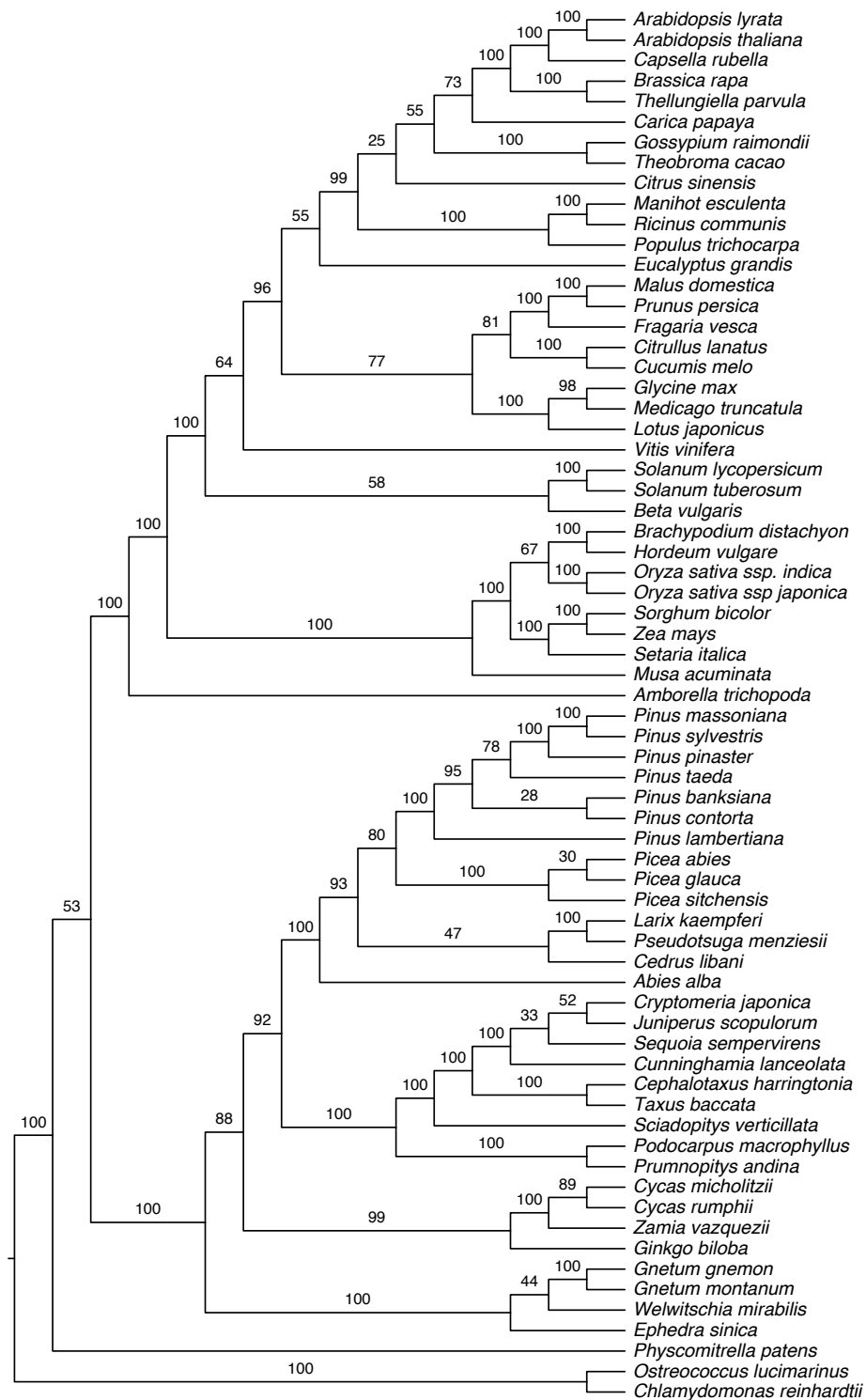
Supplementary Figure 17. Bayesian phylogenetic tree based on the reduced amino acid concatenation of 106 single-copy genes under CAT+GTR model. Numbers indicate posterior probabilities.



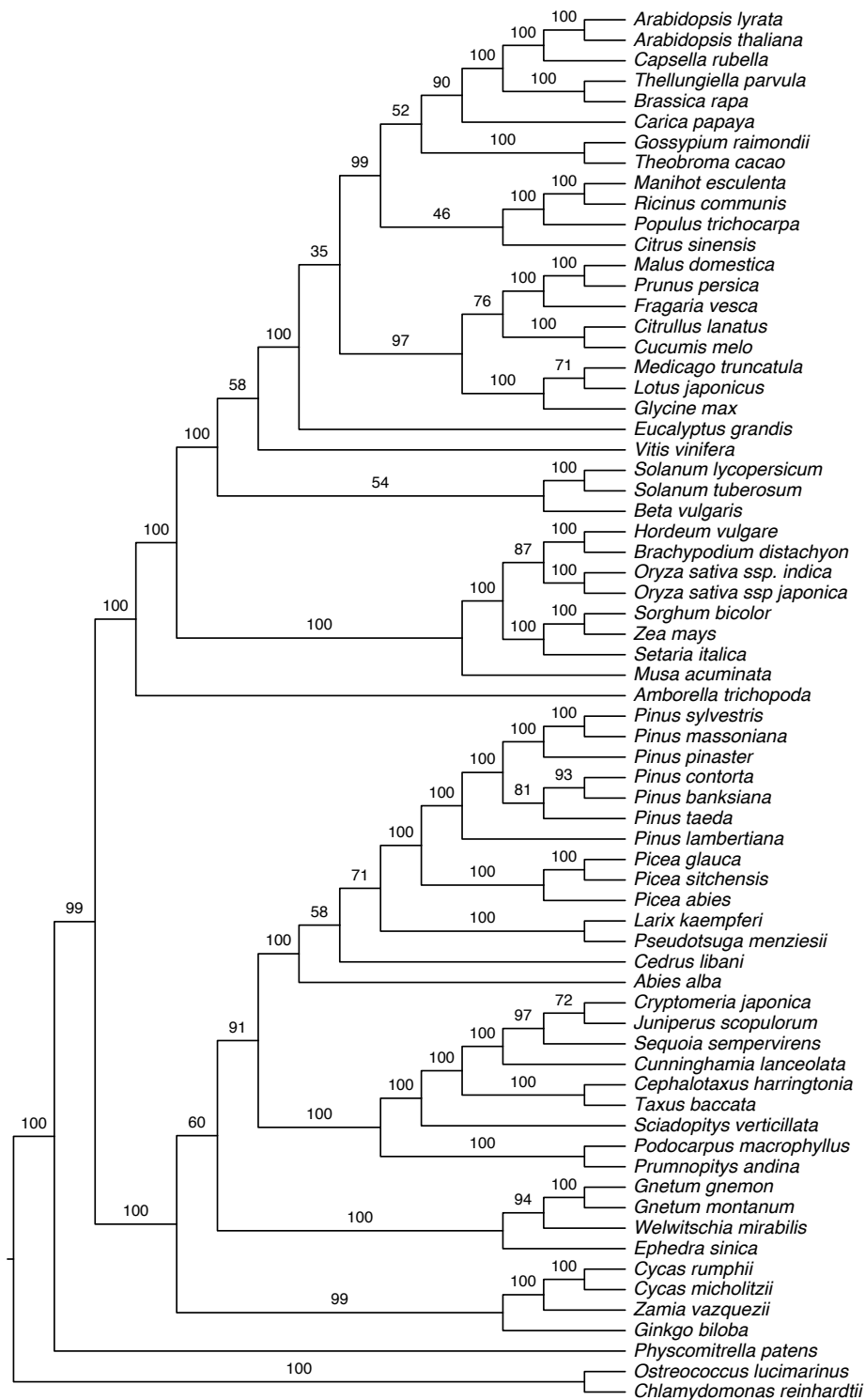
Supplementary Figure 18. Coalescent based tree inferred from gene trees of 106 single-copy genes by STAR. Numbers on branches represent bootstrap values.



Supplementary Figure 19. Coalescent based tree inferred from gene trees of 106 single-copy genes by ASTRA-II. Numbers on branches represent bootstrap values.



Supplementary Figure 20. Coalescent based tree inferred from gene trees of 106 single-copy genes by STAR. Gene trees were built without considering 3rd codon positions. Numbers on branches represent bootstrap values.



Supplementary Figure 21. Coalescent based tree inferred from gene trees of 106 single-copy genes by ASTRAL-II. Gene trees were built without considering 3rd codon positions. Numbers on branches represent bootstrap values.

Supplementary Table 1. Statistics of the sequencing reads in *Pinus pinaster* and *Pinus sylvestris*

	Tissue	Raw Data	# Reads for Assembly	# Bases for Assembly	% Reads for Assembly
<i>Pinus pinaster</i>	Shoot Apical Meristem	548,328	489,417	193,513,935	89.26%
	Shoot Apical Meristem	511,488	450,986	246,681,796	88.17%
	Cortex Root	595,575	388,906	144,683,636	65.30%
	Cortex Hypocotyl	552,098	445,605	238,646,186	80.71%
	Pith Hypocotyl	238,995	177,643	34,516,172	74.33%
	Pith Hypocotyl	141,112	110,838	31,040,865	78.55%
	Vascular Developing Root	596,858	511,873	351,624,842	85.76%
	Cortex Developing Root	466,708	380,969	251,267,660	81.63%
	Root Apical Meristem	475,279	422,455	153,747,617	88.89%
	Root Apical Meristem	605,083	535,391	338,062,907	88.48%
	Vascular Root	297,094	173,094	78,592,365	58.26%
	Vascular Root	448,120	262,767	129,947,819	58.64%
	Vascular Cotyledon	726,853	663,607	424,191,066	91.30%
	Mesophyll Cotyledon	776,974	720,010	460,528,808	92.67%
	Pith Hypocotyl	631,059	573,629	353,808,071	90.90%
	Vascular New Needle	747,050	691,860	472,097,944	92.61%
	Vascular Hypocotyl	678,337	607,026	363,679,917	89.49%
	Developing Needle	747,508	702,247	460,245,151	93.95%
	Mesophyll New Needle	682,094	601,778	328,943,737	88.23%
	Vascular Root	712,551	673,643	335,482,084	94.54%
	PPIN_454_Reads_Reg1.RL6.sff	493,631	394,947	151,121,677	80.01%
	PPIN_454_Reads_Reg2.RL6.sff	724,677	562,481	212,858,345	77.62%
	Total	12,397,472	10,541,172	5,755,282,600	85.03%
<i>Pinus sylvestris</i>	Early Embryo (E1)	603,508	539,840	250,762,894	89.45%
	Cleavage (E2)	625,195	573,611	279,768,670	91.75%
	Dominant Embryo (E3DO)	749,430	711,685	506,198,526	94.96%
	Megagametophyte (E3SU)	745,590	708,247	499,030,847	94.99%
	Dominant Embryo (E4)	817,722	780,954	552,917,241	95.50%
	Subordinate Embryos (M1)	758,018	725,345	512,694,678	95.69%
	Megagametophyte (M2)	712,754	671,451	465,102,991	94.21%
	Megagametophyte (M3)	800,707	768,722	521,997,230	96.01%
	Megagametophyte (M4)	789,571	754,048	516,959,353	95.50%
	Total	6,602,495	6,233,903	4,105,432,430	94.42%

Supplementary Table 2. Integrated transcriptomes for gymnosperms from public databases

Species	TreeGenes*	PlantGDB**	oneKP	# Transcripts	# Proteins
<i>Abies alba</i>	25,419	-	-	20,796	17,279
<i>Cedrus libani</i>	-	-	70,595	65,491	37,730
<i>Cephalotaxus harringtonia</i>	13,997	-	-	13,927	16,578
<i>Cryptomeria japonica</i>	347	24,299	-	21,922	19,688
<i>Cunninghamia lanceolata</i>	62,140	-	68,622	70,761	48,057
<i>Cycas micholitzii</i>	-	-	54,202	51,130	28,901
<i>Cycas rumphii</i>	-	10,901	22,908	21,844	12,619
<i>Ephedra sinica</i>	-	-	57,134	51,088	26,873
<i>Ginkgo biloba</i>	-	10,210	48,343	45,631	30,404
<i>Gnetum gnemon</i>	10,221	6,193	-	12,994	14,889
<i>Gnetum montanum</i>	-	-	70,316	65,123	32,549
<i>Juniperus scopulorum</i>	-	-	61,299	57,857	35,004
<i>Larix kaempferi</i>	57,484	-	-	53,274	40,692
<i>Pinus banksiana</i>	16	13,040	-	12,036	13,421
<i>Pinus contorta</i>	32	13,570	-	12,605	14,984
<i>Pinus lambertiana</i>	19,509	-	-	19,292	20,561
<i>Pinus massoniana</i>	69,738	-	-	55,525	46,423
<i>Podocarpus macrophyllus</i>	12,266	-	-	12,209	14,296
<i>Prumnopitys andina</i>	-	-	45,616	42,269	30,894
<i>Pseudotsuga menziesii</i>	393,638	9,857	-	246,282	149,717
<i>Sciadopitys verticillata</i>	11,955	-	51,723	40,310	30,035
<i>Sequoia sempervirens</i>	11,517	-	-	11,462	13,461
<i>Taxus baccata</i>	10,554	-	54,249	44,059	32,062
<i>Welwitschia mirabilis</i>	-	-	6,606	6,261	6,052
<i>Zamia vazquezii</i>	-	7,657	50,336	33,979	24,619

* TreeGenes includes ESTs, cDNAs, TSAs, and 454 assemblies

***Cycas rumphii*, *Ginkgo biloba*, *Gnetum gnemon*, and *Zamia vazquezii* are directly from PlantGDB

Supplementary Table 3. The short name and clade of species used in the current study.

Species	Short name	Clade
<i>Cephalotaxus harringtonia</i>	CHA	Cupressophytes
<i>Cryptomeria japonica</i>	CJA	Cupressophytes
<i>Cunninghamia lanceolata</i>	CUL	Cupressophytes
<i>Juniperus scopulorum</i>	JSC	Cupressophytes
<i>Podocarpus macrophyllus</i>	POM	Cupressophytes
<i>Prumnopitys andina</i>	PAN	Cupressophytes
<i>Sciadopitys verticillata</i>	SVE	Cupressophytes
<i>Sequoia sempervirens</i>	SSE	Cupressophytes
<i>Taxus baccata</i>	TBA	Cupressophytes
<i>Zamia vazquezii</i>	ZVA	Cycads
<i>Cycas micholitzii</i>	CMI	Cycads
<i>Cycas rumphii</i>	CYR	Cycads
<i>Amborella trichopoda</i>	ATR	Angiosperms
<i>Arabidopsis lyrata</i>	ALY	Angiosperms
<i>Arabidopsis thaliana</i>	ATH	Angiosperms
<i>Beta vulgaris</i>	BVU	Angiosperms
<i>Brassica rapa</i>	BRA	Angiosperms
<i>Capsella rubella</i>	CRU	Angiosperms
<i>Carica papaya</i>	CPA	Angiosperms
<i>Citrullus lanatus</i>	CLA	Angiosperms
<i>Citrus sinensis</i>	CSI	Angiosperms
<i>Cucumis melo</i>	CME	Angiosperms
<i>Eucalyptus grandis</i>	EGR	Angiosperms
<i>Fragaria vesca</i>	FVE	Angiosperms
<i>Glycine max</i>	GMA	Angiosperms
<i>Gossypium raimondii</i>	GRA	Angiosperms
<i>Lotus japonicus</i>	LJA	Angiosperms
<i>Malus domestica</i>	MDO	Angiosperms
<i>Manihot esculenta</i>	MES	Angiosperms
<i>Medicago truncatula</i>	MTR	Angiosperms
<i>Populus trichocarpa</i>	PTR	Angiosperms
<i>Prunus persica</i>	PPE	Angiosperms
<i>Ricinus communis</i>	RCO	Angiosperms
<i>Solanum lycopersicum</i>	SLY	Angiosperms
<i>Solanum tuberosum</i>	STU	Angiosperms
<i>Thellungiella parvula</i>	TPA	Angiosperms
<i>Theobroma cacao</i>	TCA	Angiosperms
<i>Vitis vinifera</i>	VVI	Angiosperms
<i>Ginkgo biloba</i>	GBI	Ginkgo
<i>Ephedra sinica</i>	ESI	Gnetophytes

Species	Short name	Clade
<i>Gnetum gnemon</i>	GGN	Gnetophytes
<i>Gnetum montanum</i>	GMO	Gnetophytes
<i>Welwitschia mirabilis</i>	WMI	Gnetophytes
<i>Brachypodium distachyon</i>	BDI	Angiosperms
<i>Hordeum vulgare</i>	HVU	Angiosperms
<i>Musa acuminata</i>	MAC	Angiosperms
<i>Oryza sativa ssp indica</i>	OSI	Angiosperms
<i>Oryza sativa ssp japonica</i>	OSJ	Angiosperms
<i>Setaria italica</i>	SIT	Angiosperms
<i>Sorghum bicolor</i>	SBI	Angiosperms
<i>Zea mays</i>	ZMA	Angiosperms
<i>Chlamydomonas reinhardtii</i>	CRE	Outgroup
<i>Ostreococcus lucimarinus</i>	OLU	Outgroup
<i>Physcomitrella patens</i>	PPA	Outgroup
<i>Abies alba</i>	AAL	Pinaceae
<i>Cedrus libani</i>	CLI	Pinaceae
<i>Larix kaempferi</i>	LKA	Pinaceae
<i>Picea abies</i>	PAB	Pinaceae
<i>Picea glauca</i>	PGL	Pinaceae
<i>Picea sitchensis</i>	PSI	Pinaceae
<i>Pinus banksiana</i>	PBA	Pinaceae
<i>Pinus contorta</i>	PCO	Pinaceae
<i>Pinus lambertiana</i>	PLA	Pinaceae
<i>Pinus massoniana</i>	PMA	Pinaceae
<i>Pinus pinaster</i>	PPI	Pinaceae
<i>Pinus sylvestris</i>	PSY	Pinaceae
<i>Pinus taeda</i>	PTA	Pinaceae
<i>Pseudotsuga menziesii</i>	PME	Pinaceae