File S1

SUPPLEMENTARY MATERIALS AND METHODS

RNA isolation and qRT-PCR

Prior to RNA isolation, wild type and *skpo-1* mutant animals were grown on *cdc-25.1* dsRNA expressing *E. coli* HT115 from the egg to L4 larval stage. L4 animals were then exposed to either *cdc-25.1* RNAi or *E. faecalis* OG1RF for 18 hours. The RNA was isolated using Trizol (Invitrogen) according to the manufacturer. RNA samples were treated with DNase I to eliminate contaminating DNA by the Turbo DNA free kit (Applied Biosystem) according to the manufacturer. qRT-PCR was performed as described (VAN DER HOEVEN *et al.* 2011). Primers used are listed in Table S6 (*act-1* served as the reference gene).

Bacterial Colonization

The CFU analysis was conducted in a manner similar to past work (GARSIN *et al.* 2001). Briefly, L4 wild type and *skpo-1* mutant animals grown on *E. coli* OP50 were exposed on 100 mm plates containing BHI-agar with gentamycin (10 μg/mL) seeded with 100 μl of *E. faecalis* OG1RF for either 12 or 36 hours at 25˚ C. Worms were washed 3 times with M9 buffer at 1.4 rpm. Worms were then washed twice with 25 mM tetramisole hydrochloride to prevent ingestion of the antibiotic treatment. Worms were incubated at room temperature for 60 minutes in 25 mM tetramisole hydrochloride supplemented with ampicillin and kanamycin, both at 1 mg/mL, to kill surface-attached *E. faecalis.* Worms were collected at 1.4 rpm and washed twice with 25 mM tetramisole hydrochloride prior to grinding. Ten worms in 10 μl were transferred to 200 μl of M9 and ground for 1 minute using a motorized pestle (Kontes cordless cat# K749540-0000 and pestles cat# K749521-1590). Serial dilutions were performed and 100 μl of each dilution plated onto 100 mm BHI gentamycin 10 μg/mL plates for 24 hours at 37˚ C.