

# Supplementary Data

## Enterocyte glycosylation is responsive to changes in extracellular conditions: Implications for membrane functions

Dayoung Park<sup>2,5</sup>, Gege Xu<sup>2,5</sup>, Mariana Barboza<sup>2,3</sup>, Ishita M. Shah<sup>4</sup>, Maurice Wong<sup>2</sup>, Helen Raybould<sup>3</sup>, David A. Mills<sup>4</sup>, Carlito B. Lebrilla<sup>1,2</sup>

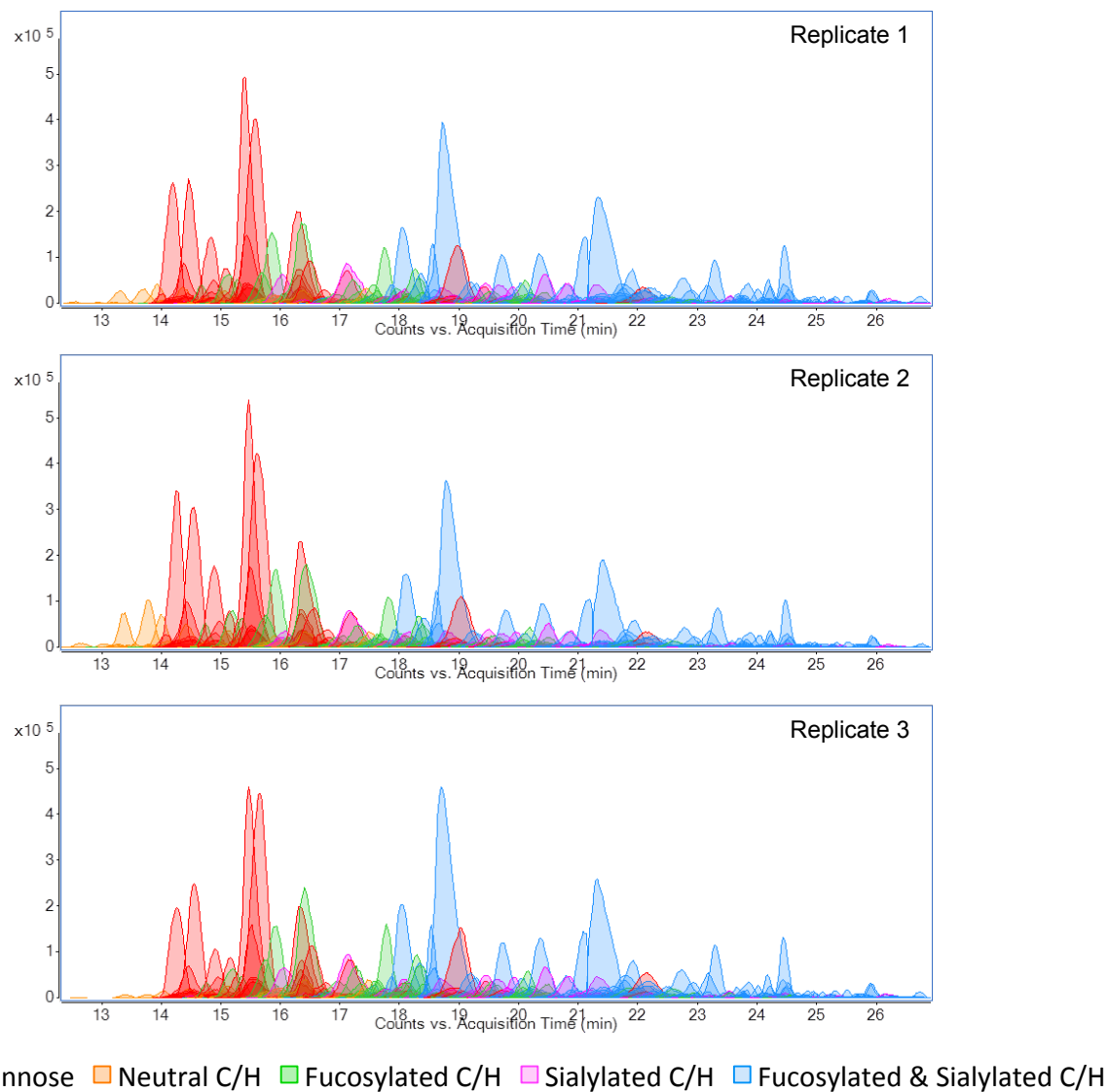
<sup>2</sup>Department of Chemistry, University of California, Davis, CA 95616 USA

<sup>3</sup>Department of Anatomy, Physiology and Cell Biology, University of California, Davis, CA 95616 USA

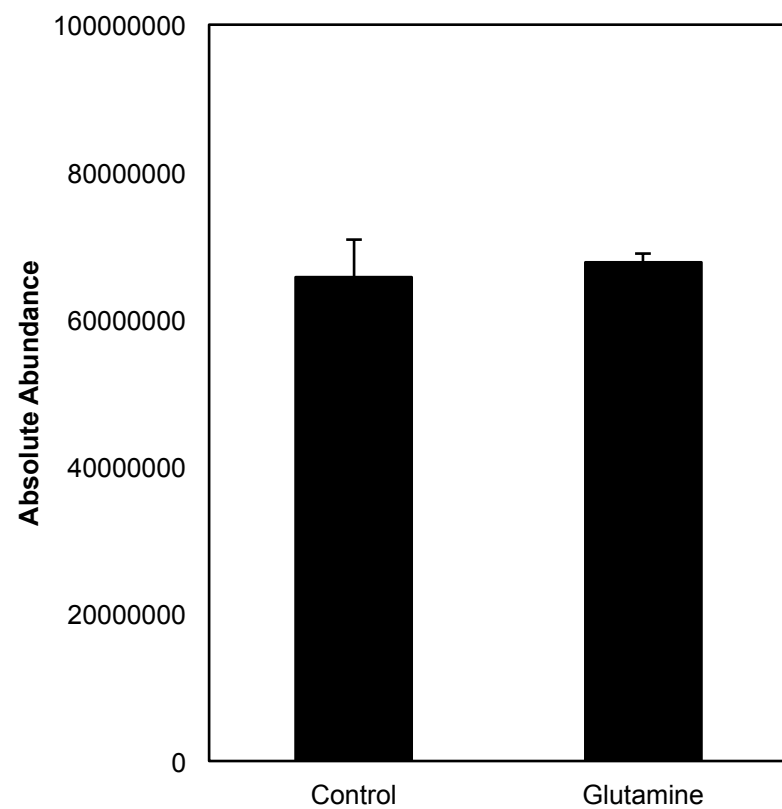
<sup>4</sup>Department of Food Science and Technology, University of California, Davis, CA 95616 USA

<sup>5</sup>These authors contributed equally to this work

<sup>1</sup>To whom correspondence should be addressed: Tel/Fax: +1-530-752-6364/+1-530-752-8995;  
E-mail: [cblebrilla@ucdavis.edu](mailto:cblebrilla@ucdavis.edu) (C.B.L.)

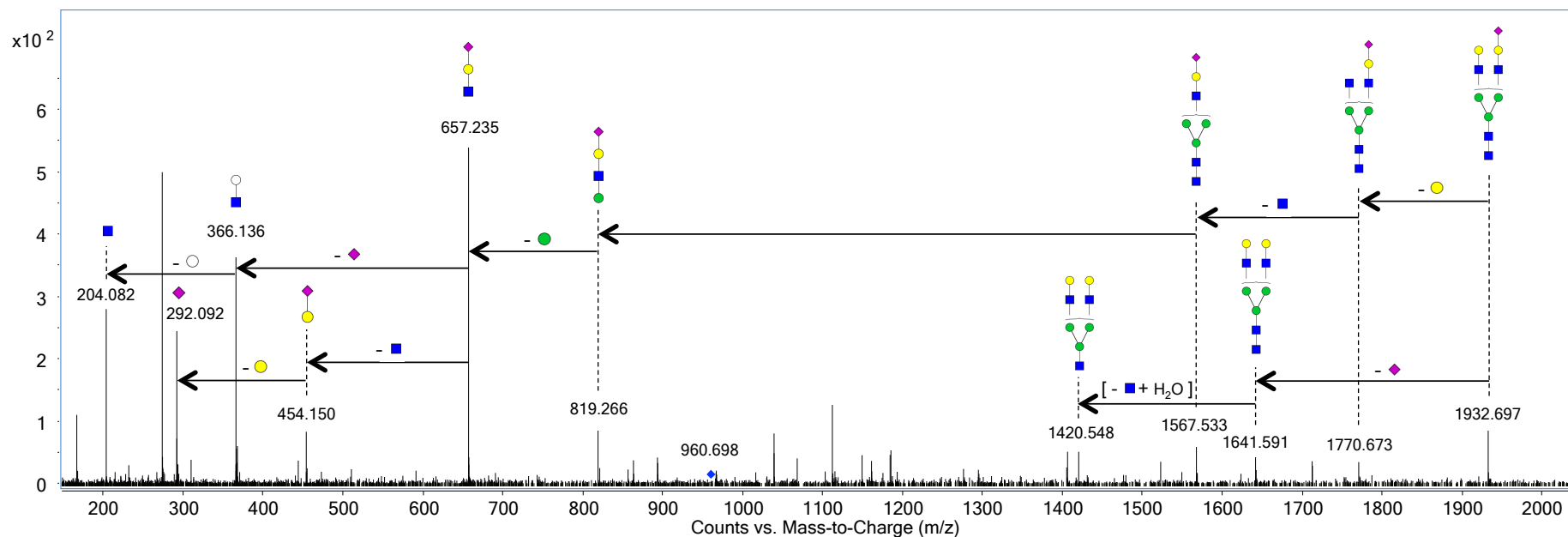
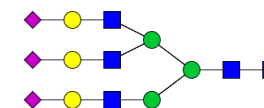


**Figure S1.** Stacked chromatograms of three biological replicates of identified Caco-2 cell surface N-glycans treated with high galactose (25 mM). Each peak represents a glycan compound and is colored according to type as indicated by the legend where C = Complex, H = Hybrid. Separation was performed using PGC-LC.



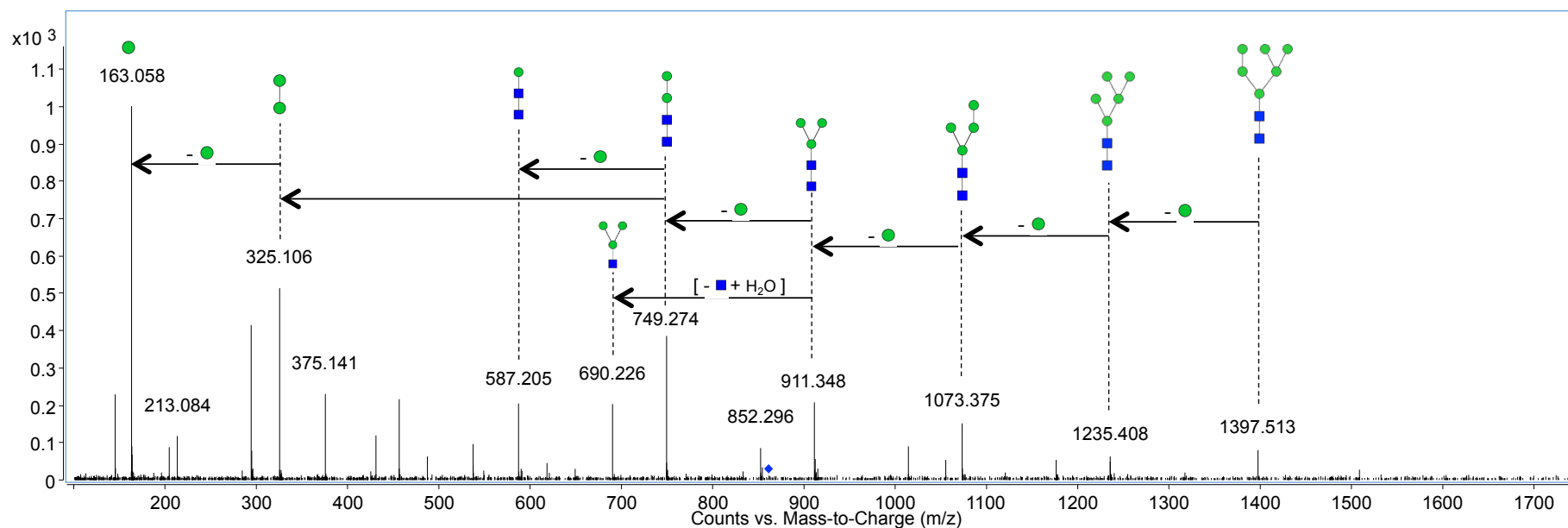
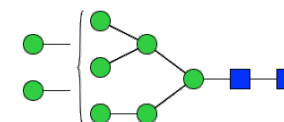
**Figure S2.** Summed absolute abundances of all sialylated structures profiled on Caco-2 cells without and with glutamine supplementation (n = 3).

◆ Precursor  $m/z = 960.698$  ( $z=3$ )



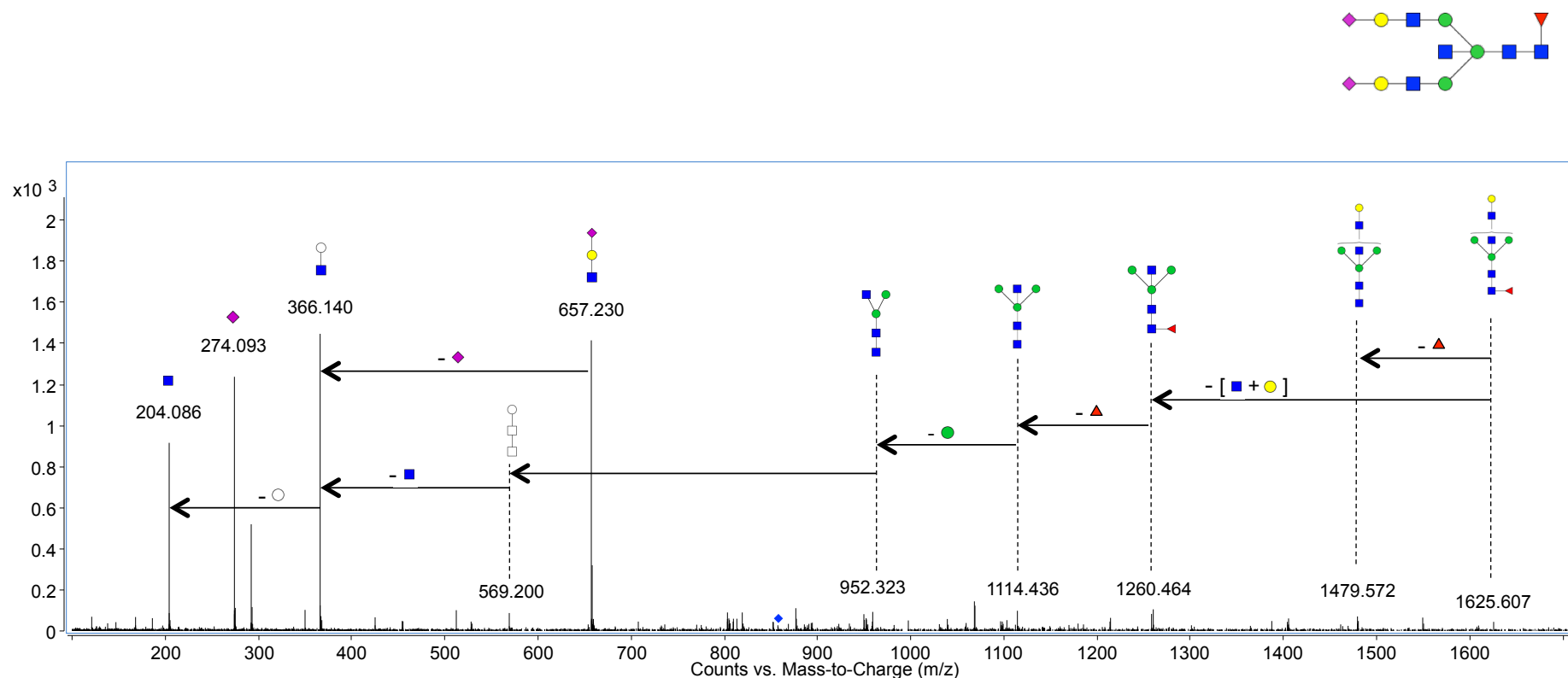
**Figure S3.** CID fragmentation spectra of N-glycan structures derived from Caco-2 as specified by  $m/z$  and symbolic representation. Sequential losses are annotated inset. Open symbols are used to indicate fragments with more than one possible position (e.g., chitobiose/antennae) and/or composition (e.g., Man/Gal). Monosaccharide symbols follow the SNFG (Symbol Nomenclature for Glycans) system details at NCBI (Varki, A., Cummings, R.D., et al. 2015).

◆ Precursor  $m/z = 861.306$  ( $z=2$ )



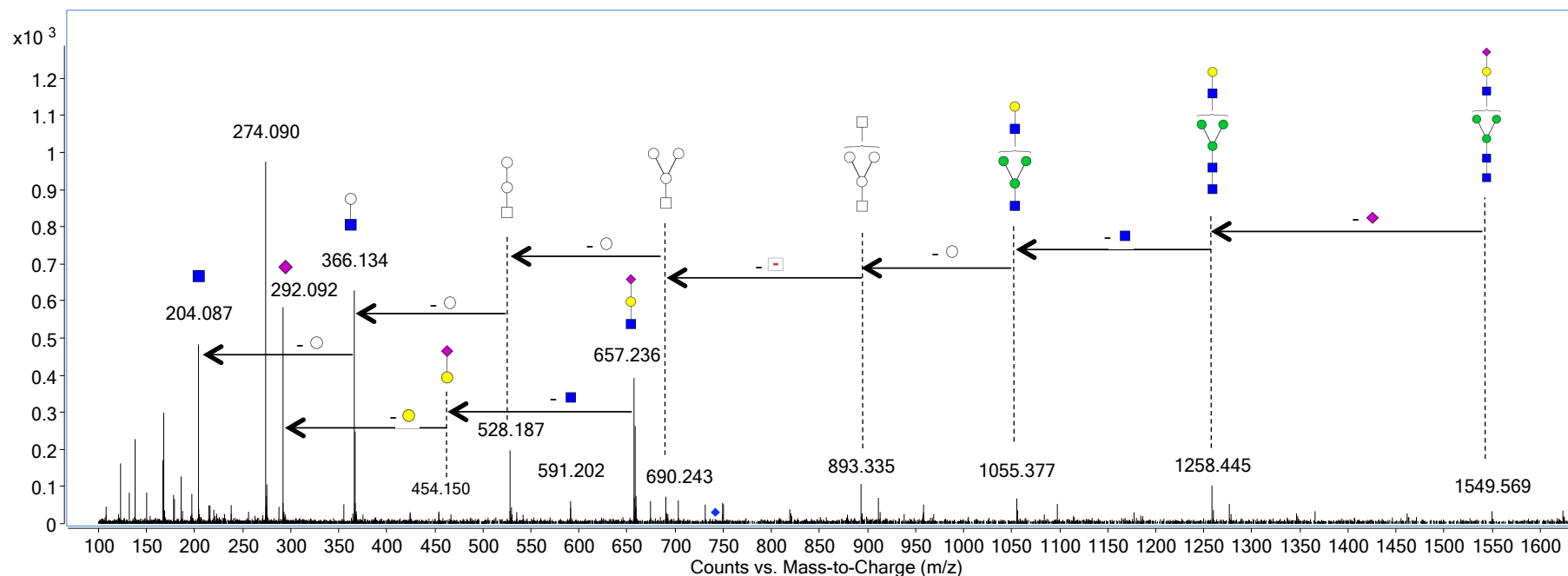
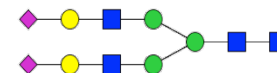
**Figure S3.** CID fragmentation spectra of N-glycan structures derived from Caco-2 as specified by  $m/z$  and symbolic representation. Sequential losses are annotated inset. Open symbols are used to indicate fragments with more than one possible position (e.g., chitobiose/antennae) and/or composition (e.g., Man/Gal). Monosaccharide symbols follow the SNFG (Symbol Nomenclature for Glycans) system details at NCBI (Varki, A., Cummings, R.D., et al. 2015).

◆ Precursor  $m/z = 858.315$  ( $z=3$ )

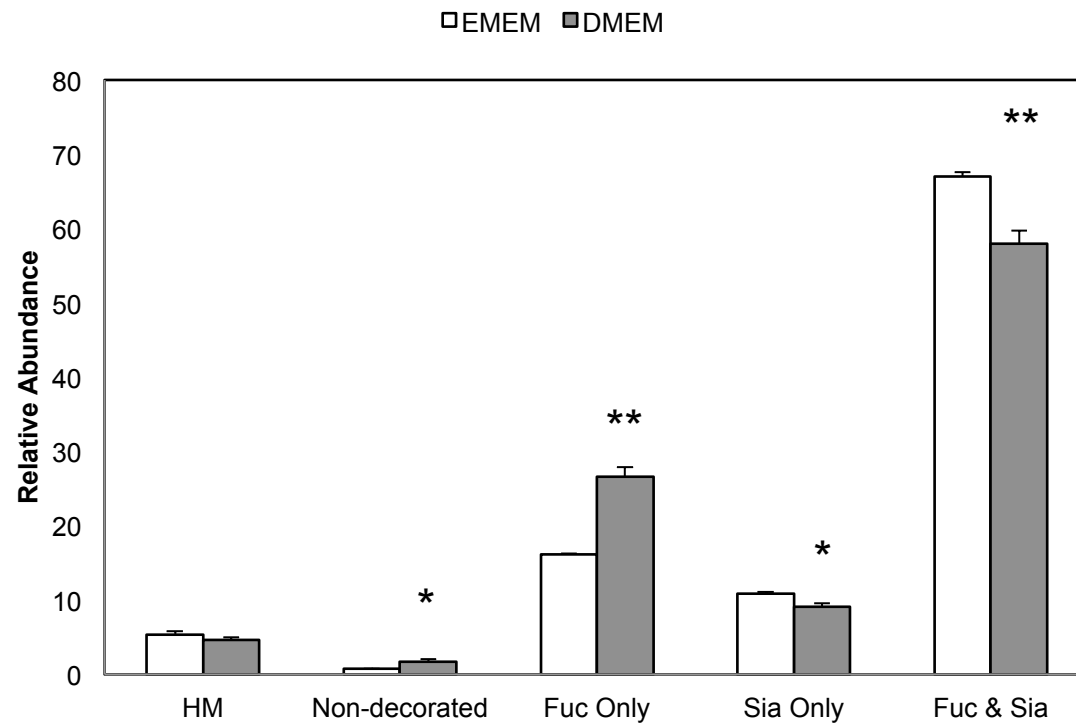


**Figure S3.** CID fragmentation spectra of N-glycan structures derived from Caco-2 as specified by  $m/z$  and symbolic representation. Sequential losses are annotated inset. Open symbols are used to indicate fragments with more than one possible position (e.g., chitobiose/antennae) and/or composition (e.g., Man/Gal). Monosaccharide symbols follow the SNFG (Symbol Nomenclature for Glycans) system details at NCBI (Varki, A., Cummings, R.D., et al. 2015).

◆ Precursor  $m/z = 741.935$  ( $z=3$ )

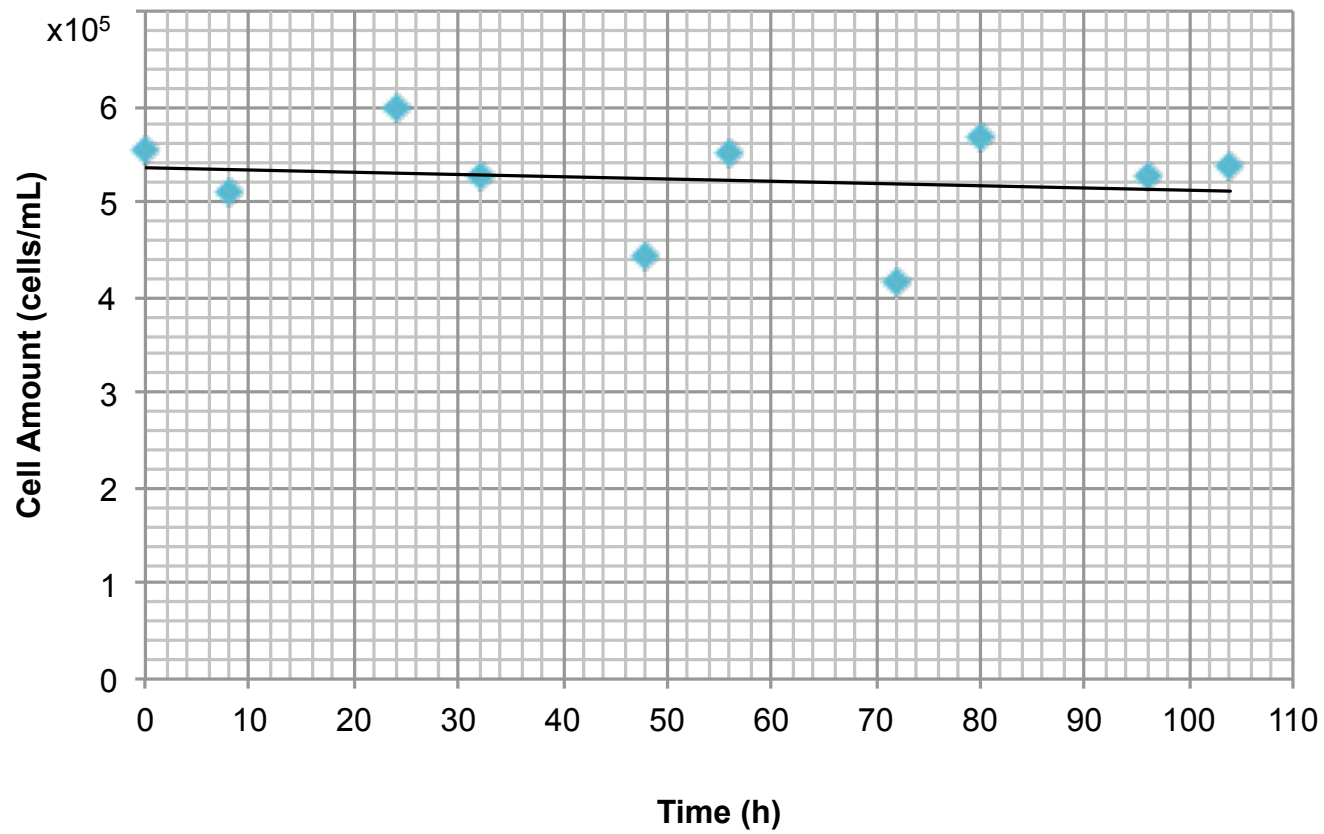


**Figure S3.** CID fragmentation spectra of N-glycan structures derived from Caco-2 as specified by  $m/z$  and symbolic representation. Sequential losses are annotated inset. Open symbols are used to indicate fragments with more than one possible position (e.g., chitobiose/antennae) and/or composition (e.g., Man/Gal). Monosaccharide symbols follow the SNFG (Symbol Nomenclature for Glycans) system details at NCBI (Varki, A., Cummings, R.D., et al. 2015).

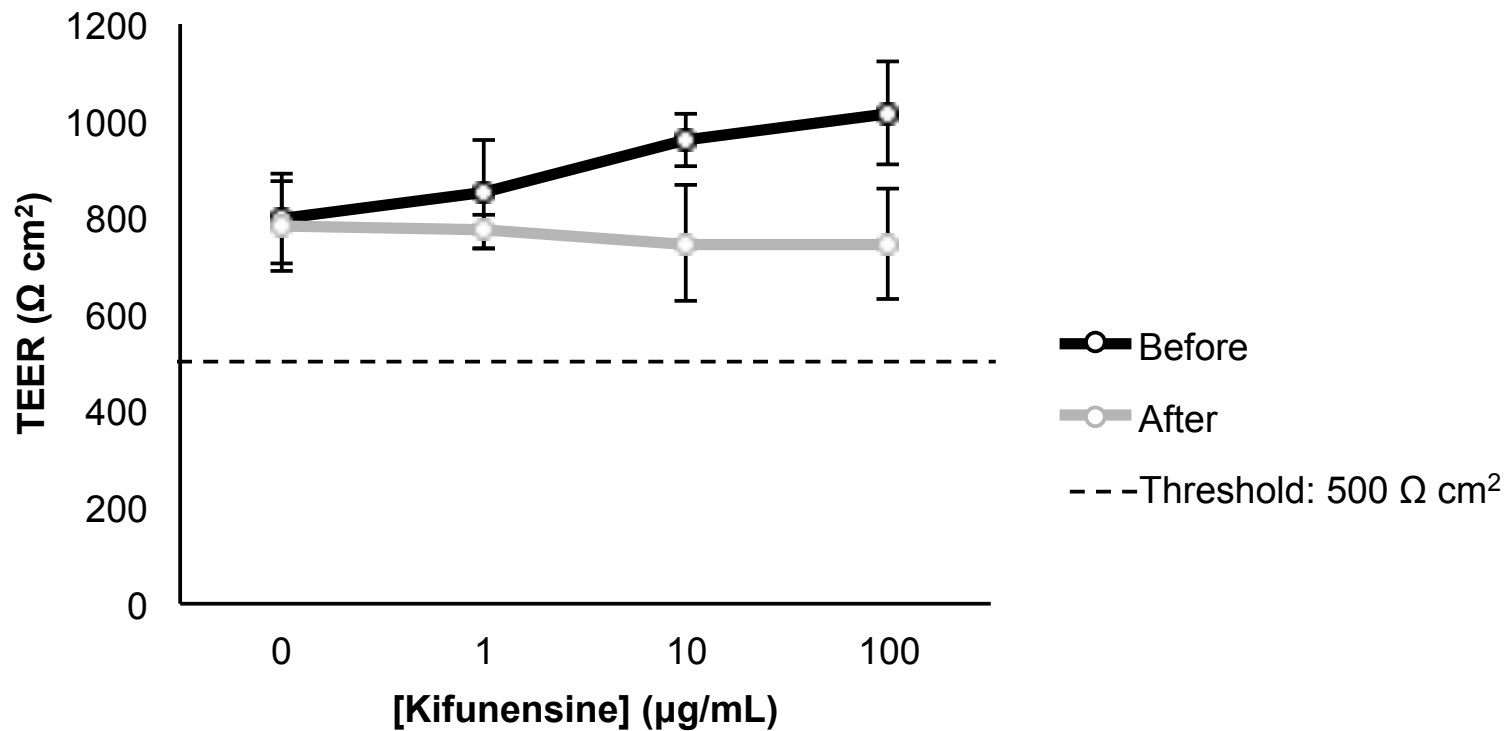


**Figure S4.** Summed relative abundances of cell surface N-glycans identified in Caco-2 cells grown in EMEM versus DMEM basal media. Data are compared by glycan types, where HM = High Mannose, Fuc = Fucosylated, and Sia = Sialylated. Error bars represent standard deviation. Asterisks indicate statistical significant changes (EMEM to DMEM), where \* $p < 0.05$  and \*\* $p < 0.01$ .

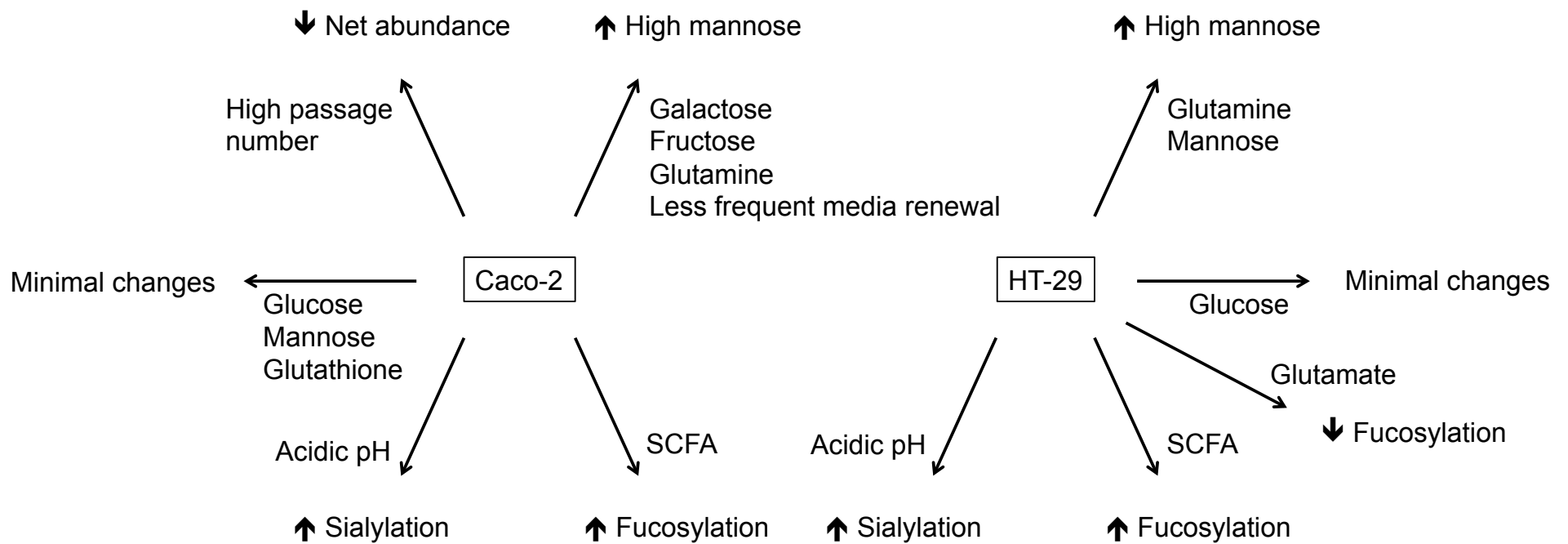




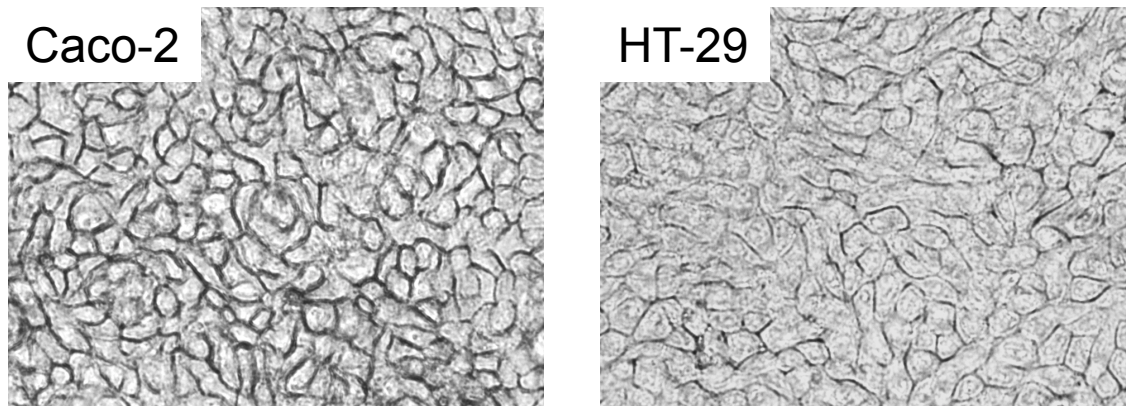
**Figure S5.** Number of viable Caco-2 cells over time from the initial introduction of kifunensine to 104 h post-treatment. The regression line is plotted inset.



**Figure S6.** Transepithelial electrical resistance (TEER) measurements in Caco-2 monolayers grown on permeable supports as a function of the amount of kifunensine added. A threshold of 500 Ω cm<sup>2</sup> was used to ensure monolayer integrity.



**Figure S7.** Summary of major changes observed in the cell surface N-glycan profiles of Caco-2 and HT-29 with metabolite and growth manipulations.



<b>Species</b>	Homo sapiens	Homo sapiens
<b>Origin</b>	Caucasian Male 72 years	Caucasian Female 44 years
<b>Tissue</b>	Colon	Colon
<b>Cell Type</b>	Epithelial	Epithelial
<b>Differentiation</b>	Spontaneous Enterocyte-like	Not spontaneous Enterocyte-like or goblet-like
<b>Growth</b>	Adherent	Adherent
<b>Base Medium</b>	Eagle's Minimum Essential Medium	McCoy's 5A Medium
<b>Passage Number (used in this study)</b>	P16 – P31	P15 – P29

**Figure S8.** Characteristics and representative microscope images of fully differentiated Caco-2 and partially differentiated HT-29 cells in culture. Images were collected at 40X magnification on a Leica DMI3000B. To follow the differentiation process, focus position was adjusted to visualize the formation of distinct outlines of cell junctions.