siRNA Selection Server: an automated siRNA oligonucleotide prediction server

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ABSTRACT

The Whitehead siRNA (short interfering RNA) Selection Web Server (http://jura.wi.mit.edu/bioc/siRNA) automates the design of short oligonucleotides that can specifically ‘knock down’ expression of target genes. These short sequences are about 21 nt in length, and when synthesized as double stranded RNA and introduced into cell culture, can reduce or eliminate the function of the target gene. Depending on the length of a gene, there are potentially numerous combinations of possible 21mers. Some experimental evidence has already shown that not all 21mers in a gene have the same effectiveness at silencing gene function. Our tool incorporates published design rules and presents the scientist with information about uniqueness of the 21mers within the genome, thermodynamic stability of the double stranded RNA duplex, GC content, presence of SNPs and other features that may contribute to the effectiveness of a siRNA.

INTRODUCTION

One way to study the function of a gene is to reduce or eliminate its expression in a cell. An emerging technology to study the role of an individual gene is RNAi and its effector molecule siRNA—short interfering RNA (1,2). A properly selected short double stranded RNA (~21 nt) targeted to a specific sequence can silence the expression of the gene. Some experimental evidence has already shown that not all 21mers in a gene have the same effectiveness at silencing gene function [see, for example (1,3–6)]. We have used a number of bioinformatics approaches to help select optimal siRNAs. The computational tools we build are useful to scientists wishing to silence specific genes, specific gene families, or even specific biological pathways within a genome.

ACCESS TO SERVER

In order to access our server, we require user registration. This permits us to limit the number of searches per day for an individual investigator. Our current limit is 15 searches per day. The use of this site is provided free of charge to the research community.

INPUT TO THE SERVER

Figure 1 shows the input form that guides the biologist through the design of siRNAs. You can enter either a cDNA sequence in raw or FASTA format, or a GI or GenBank accession

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**siRNA Selection Program**

- * Enter your sequence in **Raw or FASTA format** below,

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OR enter GI or Accession number

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- *Choose the siRNA pattern:

<table>
<thead>
<tr>
<th>Recommended patterns</th>
<th>custom</th>
</tr>
</thead>
<tbody>
<tr>
<td>☑️ AAN19TT</td>
<td>☑️</td>
</tr>
<tr>
<td>☑️ NANN21</td>
<td>Enter pattern up to 23 bases</td>
</tr>
</tbody>
</table>

- Filter criteria:
  - *GC percentage: from [30] to [70]
  - *exceed a run of 4 or more T or A in a row
  - *exceed a run of 4 or more Gs in a row
  - *include less than 7 consecutive GC in a row.
  - ☑️ equal %(+/- 10 %) for all 4 bases.

- *End your siRNAs with [U.U]  

[Search]  [reset]

**Note:** *: required parameters.

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**Figure 1.** Data input web page to begin design of siRNAs for your target sequence.

number. Next you need to choose a pattern for the composition of the siRNA. There are several commonly used siRNA sequence patterns (for example, AAN19TT; for more detail see http://www.rockefeller.edu/labheads/tuschl/sirna.html). Our website provides biologists the flexibility to use these pre-defined siRNA patterns or input their own pattern. The full NC-IUB nomenclature can be used to represent the sequence. In addition, a modified regular expression pattern can be constructed.

Several filters related to base composition are also available in the design process. Sequence characteristics such as GC percentage, base variations and the number of repetitive bases in a row are available for selection. These filters are based on experimental results observed by Tuschi and others. A run of
four or more Ts or As should be excluded under some circumstances because four or five Ts in a row is the transcription terminator signal for pol III. If it is desired to design hairpin RNA expression vectors that are expressed from pol III promoters (U6, H1, or tRNA promoter), pol III terminator signals must be excluded from the sense or anti-sense strand. Similarly four or more Gs in a row should be excluded because oligoG-containing RNAs may form tetraplexes and are difficult to chemically synthesize with some, but not all types of RNA chemistry. GC rich sequences form more stable duplexes than those that are AT rich, thus more than seven G/C pairs in a row would be suboptimal.

The final step in design is to select the terminal bases for the siRNA. To construct the siRNAs, we first find all 23mers with your pattern. For the sense strand, we consider the 21mer from positions 3 to 23 of the candidate siRNAs for further analysis. For the anti-sense strand, the 21mer is the reverse complement of positions 1 to 21 of the sense strand. The two bases at the 3' end of the 21mers are replaced by your choice of terminal bases. If you choose NN, the original two nucleotides are kept intact.

SELECTING OLIGOS FOR FURTHER ANALYSIS

The next screen (Figure 2) shows the potential siRNAs based on your input sequence, pattern and filtering choices. Although the hits are initially sorted by position within the target sequence, the results can be sorted by various criteria. The interesting information includes the position of the siRNA within the input sequence, the pattern the siRNA matches, the percentage GC content and the thermodynamic values based on stability of the 5' ends of the sense and anti-sense siRNAs. Recent experimental evidence indicates that the two strands of a siRNA duplex do not enter into the RNAi pathway equally. Rather, the less stable 5' end (either sense or anti-sense) in the siRNA duplex directs that strand to enter into the RISC complex and has more effect silencing the target gene (7–10). To calculate stability of the duplex, we examine the free energy at its two ends using the nearest neighbor method (11). For each siRNA, a model helix is made from each end of the duplex with five bases of the 5' end of the first strand, four Ns, then seven bases of the 3' end of the second strand. For each model, thermodynamic parameters for four nearest
neighbors and one 3’-dangling nucleotide are summed. The energy unit is K/mol at 1 M NaCl, pH 7 and 37 °C. Using the stability information one may wish to select siRNA duplexes that are less stable at the 5’ end of the anti-sense siRNA.

Because the objective of using siRNAs is to silence a specific gene in a mammalian cell, the base-pairing region for a siRNA must be selected carefully to avoid similarity to an unrelated mRNA. Therefore, our program BLASTs (12) each siRNA candidate against the human or mouse UniGene (13) database, and parses the output into an HTML table for users. Furthermore, we have developed an algorithm for mapping the siRNA to the genomic sequences and indicate to the user if the siRNA is at an exon–exon junction or if the siRNA contains any SNPs. The positions of these junctions and any SNPs are calculated in the following manner. Each week every UniGene entry is BLATted (14) against the current genome build. The position of the siRNA in the genome is calculated by mapping its position on the best-hit sequence (see below) with the genomic positions of the best hit. This position is used to calculate the exon–exon boundaries. Starting your design with either a GenBank accession number or a sequence, SNP and 3’-UTR/coding/5’-UTR locations (see below) are calculated by mapping the siRNA position on the best-hit sequence with the GenBank entry of the best hit.

There are two options for getting your final results—you can receive the URL for your results by email or you can wait for the results to appear in the browser. If you are submitting more than a dozen sequences for further analysis, it is best to receive the results by email.

THE RESULTS PAGE

The best hit to your input sequence is found by BLASTing your query sequence against the UniGene database (Figure 3). After the BLAST search is completed, the results are displayed in tabular form with links to explore the BLAST results (Figure 4). By default, the oligos in the table are sorted by the position of the siRNA within the target sequence. With the Select box on the left menu bar, you can re-sort the siRNAs by any one of the following criteria: query position, type (siRNA pattern), GC%, thermodynamic values, exon—exon boundary, the number of SNPs and BLAST result.

The positions of the candidate oligos in the target sequence are colour coded to represent the region of the gene in which they fall. For the best UniGene hit in the candidate oligos, green indicates that the oligo is upstream of the CDS. Red indicates that the oligo is within the CDS and blue indicates that the oligo is downstream of the CDS. If the best GenBank hit is an NM entry, then the green region is the 5’-UTR and the blue indicates that the oligo is in the 3’-UTR.

DOWNLOADING DATA

There are two options for downloading the results of your siRNA design. You can choose to download all of the information in the result table or just the information about the siRNAs. Both options produce tab-delimited files that can easily be read into a spreadsheet.

Figure 3. Result page after a BLAST search is done on candidate oligos.
**FUTURE DEVELOPMENTS**

We are continuing to develop our website to include additional capabilities. One such feature is to allow the user to design siRNA of various lengths rather than only 21mers. In addition we will be adding a batch capability so that multiple sequences can be input at once. Our site will soon provide assistance in predicting short hairpin siRNAs. Also, as new rules are discovered by our group and others, we will incorporate them into our siRNA design server.

**SUMMARY**

In summary, we wish to emphasize that our siRNA design server was built in collaboration with laboratory scientists. Our main goal is to provide the most accurate siRNA results and flexibility for the end user so that resulting information can be effectively evaluated.

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**REFERENCES**


