In vivo magnetic resonance spectroscopy (MRS) provides information about metabolite concentrations in tissue. Recently citrate was detected by MRS in subgroups of pediatric brain tumors. Citrate is an intermediate in the tricarboxylic acid (TCA) cycle and accumulates in tissue when the glycolytic rate exceeds the TCA cycle activity, a feature of malignant tumors. Currently, no practical indicators allow clinicians to predict risk for malignant progression of pediatric astrocytomas (World Health Organization [WHO] grade II). Medical records and citrate concentrations measured with in vivo MRS of 29 pediatric astrocytomas were reviewed. This included 6 patients with astrocytomas (WHO II) who had stable disease (indolent LGA) for >2 years, 7 with aggressive grade II astrocytomas (aggressive LGA), 13 with anaplastic astrocytomas (WHO III), and 3 with glioblastoma (WHO IV) with disease progression within 2 years. Citrate was observed in all patients with aggressive LGA, and the mean citrate concentration was significantly higher in this group than among those with indolent LGA (mean ± standard deviation, 4.1 ± 1.1 vs 0.6 ± 0.8 mmol/kg; P < .0001). There was no consistent pattern for citrate in anaplastic astrocytoma and glioblastoma, with citrate prominent in some lesions whereas undetectable in others. It is unclear whether citrate accumulation occurred because of fundamental defects of citrate regulation or was secondary to altered physiological conditions. Nonetheless, prominent citrate identified a subgroup of pediatric grade II astrocytomas destined for aggressive behavior. Citrate was not specific for poor outcome because it was not detectable in all high-grade astrocytomas. In high-grade astrocytoma, tumors with prominent citrate may constitute a metabolic subclass.

Keywords: astrocytoma, citrate, metabolism, MR spectroscopy, pediatrics.

In vivo magnetic resonance spectroscopy (MRS) provides information about the biochemistry of tissue by assessing concentrations or relative concentrations of metabolites and other small low-weight molecules. A recently reported chemical detectable in subgroups of pediatric brain tumors is citrate. Citrate is an intermediate in the tricarboxylic acid (TCA) cycle and accumulates in tissue where the glycolytic rate exceeds the TCA cycle activity, a long-known feature of malignant tumors. Excessive citrate can be used by cells to transport mitochondrial acetyl-CoA carbons to the cytoplasm for the biosynthesis of fatty acids ultimately needed for the de novo synthesis of cell membranes. This is an essential step to support cell divisions and to increase the production of biomass of growing tumors. In vivo citrate levels were reported to be particularly high in pontine gliomas—tumors with apparently inevitable aggressive behavior, despite presenting frequently with histopathological features typical for grade II diffuse astrocytomas.

The overall objective of this retrospective study was to examine the significance of citrate accumulation in pediatric astrocytomas outside the pons. Specifically, the goals were (1) to determine whether high citrate levels separated aggressive from indolent grade II astrocytoma, (2) to examine whether prominent citrate was a specific feature of tumors with poor outcome also observed in high-grade astrocytomas, and (3) to determine whether citrate accumulation correlated with magnetic resonance radiographic features of the lesions.
Material and Methods

Patients

Pediatric patients with biopsy-confirmed astrocytoma who underwent MRS studies were considered for this study. We included only patients who underwent at least 1 MRS study in which the region of interest that was selected for MRS enclosed only tumor tissue without partial volume of surrounding normal appearing tissue as judged by 2 pediatric neuroradiologists. This assessment was performed prior to and independent of the review of clinical information and was extracted from an already existing MRS database. This list of patients (40 subjects) was distributed to the clinicians for a review of the medical records. Subsequent to the review, 9 patients with <2 years of clinical follow-up, unless they died of complications due to progressive disease within that period, were excluded. Also excluded were 2 patients who died of other causes, such as sepsis or organ failure.

Results of only 1 MRS study per patient were analyzed. This was, when available, the MRS study obtained at the time of initial diagnosis. Otherwise, the study closest to initial diagnosis was selected. All pathological assignments were based on initial biopsy findings, and consensus was reached for each tumor. Six patients were identified with diffuse low-grade astrocytoma (World Health Organization [WHO] grade II) and no clinical and radiological progression over ≥2 years of follow-up (indolent LGA). Seven patients were identified with diffuse low-grade astrocytoma (WHO grade II) with progression ≤2 years after diagnosis (aggressive LGA). We also analyzed data obtained from 13 children with anaplastic astrocytoma (AA; WHO grade III) and 3 pediatric patients with glioblastoma (GBM; WHO grade IV) with progression in ≤2 years. All patients with indolent LGA were alive, whereas all with aggressive LGA, AA, and GBM had died of their disease by the time of the completion of this study. Clinical information reviewed for each subject included the time of diagnosis, time to disease progression and death, treatments received, pathology, and surgical reports, as well as the time of MRS studies relative to the time of diagnoses and therapies (Table 1). Criteria for disease progression were new or worsening clinical symptoms, such as problems with swallowing, speech, and walking, during a clinical consultation. In cases in which there were any doubts about disease progression based on these clinical symptoms alone, clinicians ordered a magnetic resonance study, which confirmed a significant growth of a lesion (as per the Children’s Oncology Group guidelines: “20% or more increase in the product of perpendicular diameters of ANY target lesion, or the appearance of one or more new lesions.”12). Progression was also diagnosed when a significant worsening of the disease was noted on MRI and subsequently confirmed by a clinical evaluation (criteria as above). The selection of patients and sorting them into indolent LGA, aggressive LGA, AA, and GBM was performed by 2 authors (M.L. and J.L.F.) and was independent of the fully automated MRS data analysis (this is explained in detail below).

This retrospective study was approved by the Institutional Review Board (IRB). Some of the patients were previously enrolled in prospective research studies and parental consent was obtained. For other patients, MRS data were obtained as part of routine clinical evaluations. For these cases, the IRB permitted the review of already existing data and medical records, and the requirement for parental consent was waived.

MRS Acquisition and Quantitation

All MRS studies were performed on a 1.5T MR system (Signa LX; GE Healthcare). Patients aged ≤5 years were anesthetized with 100–200 μg/min/kg propofol throughout the magnetic resonance study. Single-voxel pointresolved spectroscopy (PRESS) with a short echo time of TE = 35 ms, a repetition time of TR = 1.5 s, and 128 signal averages was used for all acquisitions. With these parameters, the total acquisition time, including scanner adjustments, was less than 5 minutes. All MRS data were acquired prior to contrast injection (Magnevist [gadopentetate dimeglumine], Bayer Healthcare). T2-weighted fast spin-echo, FLAIR, and T1-weighted FLAIR images were acquired in all cases before contrast injection and reviewed to determine the extension of a lesion. The position of the region of interest selected for MRS was documented on 3 MRIs and reviewed to ensure that no partial volume for normal appearing tissue was enclosed. Sizes and shapes of the regions of interest were adjusted to the lesion sizes and typically varied from 5 to 10 cm². Spectra were processed using fully automated LCModel software (Stephen Provencher, Canada, LCModel Version 6.1-4F), and absolute metabolite concentrations (in mmol/kg tissue) of citrate were determined. Also measured were the concentrations of N-acetylaspartate (NAA), creatine, total choline, myo-inositol, glutamate, glutamine, the sum of Glu and Gln (Glx), guanidinoacetate, and lactate, as well as lipid (and possibly underlying macromolecules) intensities at 0.9 ppm (parts per million) and 1.3 ppm (LipMM09 and LipMM13), as described in detail in previous publications.13–15 Often relative concentration ratios instead of absolute concentrations are reported in the literature. To allow comparisons with these studies, we also analyzed concentration ratios relative to creatine level.

Radiographic Appearance

The anatomical location and the contrast enhancement pattern of the lesions were reviewed to determine whether citrate accumulation was limited to tumors in certain brain regions or to tumors with intact/impaired blood-brain-barrier. The apparent diffusion coefficient (ADC), a quantitative measure of the diffusion of water molecules within tissue, was also analyzed. To measure ADC, an echo-planar sequence with an echo time TE of 85 ms, 5–7 mm slice thickness, 20–26 cm field of view, 128 × 128 matrix size, and a b-value of 1000 s/mm² was used. ADC maps were generated from the raw data using a diffusion-weighted imaging (DWI) sequence with the following parameters: TR/TE = 8000/100 ms, echo train length = 16, b-value = 1000 s/mm², 64×64 matrix, slice thickness = 5 mm.
<table>
<thead>
<tr>
<th>Age at diagnosis, years</th>
<th>Sex</th>
<th>Lesion appearance/location</th>
<th>Time of MRS scan</th>
<th>Treatments prior to MRS</th>
<th>Time without/to progression, months</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>11.2</td>
<td>M</td>
<td>Nonenhancing thalamic tumor</td>
<td>25 months after Dx</td>
<td>RT + TEM</td>
<td>&gt;60</td>
<td></td>
</tr>
<tr>
<td>6.4</td>
<td>F</td>
<td>Cerebellar non-enhancing lesion</td>
<td>At initial presentation</td>
<td>None</td>
<td>&gt;49</td>
<td></td>
</tr>
<tr>
<td>4.5</td>
<td>F</td>
<td>Infiltrating non-enhancing bithalamic lesion with involvement of brainstem</td>
<td>3 months after Dx, post biopsy</td>
<td>None</td>
<td>&gt;41</td>
<td>See Fig. 2</td>
</tr>
<tr>
<td>17.8</td>
<td>F</td>
<td>Minimally enhancing pineal region mass</td>
<td>5 years after Dx</td>
<td>RT</td>
<td>&gt;60</td>
<td></td>
</tr>
<tr>
<td>13.3</td>
<td>M</td>
<td>Nonenhancing tectal plate glioma</td>
<td>6 months after Dx, post biopsy</td>
<td>None</td>
<td>&gt;30</td>
<td></td>
</tr>
<tr>
<td>1.1</td>
<td>M</td>
<td>Minimally enhancing intraventricular mass</td>
<td>At initial presentation</td>
<td>None</td>
<td>&gt;24</td>
<td></td>
</tr>
<tr>
<td>10.3 ± 6.5</td>
<td>3M/3F</td>
<td>Grade II diffuse astrocytoma with aggressive behavior</td>
<td></td>
<td></td>
<td>&gt;44 ± 15</td>
<td></td>
</tr>
<tr>
<td>4.1</td>
<td>M</td>
<td>Heterogeneous enhancing, hemorrhagic right frontal mass</td>
<td>At initial presentation</td>
<td>None</td>
<td>24</td>
<td>Recurrent as grade III after 2 yrs</td>
</tr>
<tr>
<td>13.3</td>
<td>F</td>
<td>Frontal lobe cystic/enhancing lesion</td>
<td>15 months after Dx, increased symptoms</td>
<td>Chemotherapy</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>5.0</td>
<td>F</td>
<td>Bithalamic nonenhancing mass</td>
<td>At initial presentation</td>
<td>None</td>
<td>5</td>
<td>See Fig. 3</td>
</tr>
<tr>
<td>8.4</td>
<td>F</td>
<td>Nonenhancing tectal plate glioma</td>
<td>6 months after Dx, post biopsy</td>
<td>None/observation</td>
<td>18</td>
<td>Recurrent as grade III after 18 months</td>
</tr>
<tr>
<td>13.6</td>
<td>M</td>
<td>Patchy enhancing and infiltrating mass at posterior thalamus</td>
<td>11 months after Dx, increased symptoms</td>
<td>Shunt/observation</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>6.0</td>
<td>F</td>
<td>Both enhancing and non-enhancing splenium/callosal lesion</td>
<td>3 months after Dx, increased symptoms</td>
<td>Partial resection/observation</td>
<td>0</td>
<td>Progressed to grade IV</td>
</tr>
<tr>
<td>7.8</td>
<td>F</td>
<td>Heterogeneous mass with solid cystic components at left cerebellar peduncle and brainstem</td>
<td>12 months after Dx, recurrent disease</td>
<td>Partial resection/observation</td>
<td>18</td>
<td></td>
</tr>
<tr>
<td>8.3 ± 3.8</td>
<td>2 M/5 F</td>
<td>Anaplastic astrocytoma (WHO III)</td>
<td></td>
<td></td>
<td>11.9 ± 8.5</td>
<td></td>
</tr>
<tr>
<td>8.1</td>
<td>M</td>
<td>Cystic/solid enhancing bifrontal mass with extensive edema</td>
<td>3 months post Dx, residual tumor</td>
<td>partial resection, RT + IRI + TEM</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>6.1</td>
<td>F</td>
<td>Mildly enhancing thalamic lesion</td>
<td>At initial presentation</td>
<td>None</td>
<td>17</td>
<td></td>
</tr>
<tr>
<td>18.7</td>
<td>F</td>
<td>Minimal enhancing frontal mass</td>
<td>At initial presentation</td>
<td>None</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>14.3</td>
<td>F</td>
<td>Bithalamic tumor with heterogeneous enhancement</td>
<td>3 months after Dx</td>
<td>TEM</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td>17.9</td>
<td>F</td>
<td>Extensive lesion with enhancing areas</td>
<td>2 months after Dx, Post-biopsy</td>
<td>TEM (four days)</td>
<td>1</td>
<td>Gliomatosis cerebri</td>
</tr>
<tr>
<td>14.2</td>
<td>M</td>
<td>Temporal lobe lesion with focal areas of enhancement</td>
<td>At initial presentation</td>
<td>None</td>
<td>7</td>
<td>Gliomatosis cerebri</td>
</tr>
<tr>
<td>15.6</td>
<td>M</td>
<td>Nodular enhancing lesion at fourth ventricle</td>
<td>2 months after Dx</td>
<td>RT (spine) + TEMb</td>
<td>1</td>
<td>History of spinal cord AA</td>
</tr>
<tr>
<td>16.0</td>
<td>M</td>
<td>Non-enhancing thalamic tumor with intraventricular extension</td>
<td>At initial presentation</td>
<td>None</td>
<td>10</td>
<td></td>
</tr>
</tbody>
</table>

Continued
1000 s/mm² was used. The magnetic resonance diffusion scans were conducted before contrast agents were administered. Manufacturer-provided software was used to calculate ADC maps. The maps were then stored on the hospital PACS system (Synapse; Fuji), and the mean ADCs for regions that matched areas selected for the MRS studies were computed.

**Statistical Analyses**

As the primary goal of this study was to specifically test whether citrate was different in aggressive and indolent low-grade astrocytomas, an unpaired 2-tailed Student’s t-test with unequal variance was considered to be the appropriate method. The Student’s t-test was also used to test for other metabolic features that might be different in these 2 groups. The Cramer-Rao lower bounds (CRLBs) were used to determine whether there was evidence of the presence of citrate in spectra of high-grade astrocytomas. CRLBs are objective indicators for the reliability of fit, taking into consideration the noise level of the magnetic resonance signal. A CRLB of >100% indicated that there was no evidence of citrate in the spectrum. To determine whether citrate levels correlated with the ADC, data from all patients were pooled, and the Pearson correlation coefficient was calculated.

**Results**

**Citrate Levels in Aggressive and Indolent Grade II Astrocytomas**

Citrate was significantly higher in grade II astrocytoma with disease progression in ≤2 years than in stable grade II astrocytoma (mean ± standard deviation, 4.1 ± 1.1 vs 0.6 ± 0.8 mmol/kg; \( P < .0001 \), by Student t test). The concentration ratio of citrate relative to creatine was also significantly higher in aggressive LGA versus indolent LGA (0.7 ± 0.3 vs 0.1 ± 0.1; \( P < .001 \)) (Figs 1–3). In addition, NAA was significantly reduced in aggressive LGA versus indolent LGA, but at a lower level of significance (0.7 ± 0.7 vs 2.9 ± 1.1 mmol/kg; \( P < .01 \)). No other metabolic features were significantly different (Table 2).

**Citrate in High-grade Astrocytomas**

Prominent citrate was not a specific feature of high-grade astrocytomas with poor outcome (Fig. 4). In anaplastic astrocytomas, citrate levels varied considerably in individual patients (2.7 ± 2.7 mmol/kg; range, 0.0–10.1 mmol/kg). The tumor with highest level of citrate included in this study was in a patient with anaplastic astrocytoma. However, citrate was undetectable in 3 patients with anaplastic astrocytomas. Data from only 3 patients with GBM at diagnosis were available for this study. The averaged spectrum from these patients showed no evidence for citrate (Fig. 5).
When all data from all patients were pooled, there was a weak but significant positive correlation between citrate level and the tissue ADC ($r^2 = 0.6; P < .001$, by Pearson correlation). Citrate accumulated in enhancing and nonenhancing lesions, and there was no apparent correlation with lesion location (Table 1).

Other Metabolic Features of Pediatric Astrocytomas

All metabolite concentrations and ratios measured with our MRS method for indolent LGA, aggressive LGA, and AA are summarized in Table 2. These metabolic features were analyzed using analysis of variance and the Fisher multiple comparison test. The analyses did not reveal any significant additional findings and are thus not described in detail. In addition, when indolent LGA and aggressive LGA were pooled and compared with anaplastic astrocytoma, there were no significant differences observed. Some of the differences between low-grade and high-grade astrocytomas reported in the literature, such as generally higher mean total choline, lactate, and lipid levels in high-grade astrocytomas, were observed but did not reach significance.

Discussion

Prominent Citrate in Aggressive LGA

Citrate is a key metabolite in the TCA cycle and can accumulate in tissues with increased glycolytic rates in the absence of equally up-regulated TCA cycle activity, a well-known feature of malignant tumors (i.e., the Warburg effect). Our study identified prominent citrate as a metabolic feature that distinguished aggressive pediatric astrocytomas from stable astrocytoma. This included patients for whom MRS was performed before clinical or radiological manifestation of aggressive disease. Previously, it was reported that citrate was abnormally elevated in pontine gliomas. For these tumors, the detection of citrate at diagnosis adds little to the already poor prognosis. On the other hand, stratification of astrocytomas outside the pons is a significant problem in pediatric neuro-oncology, and early identification of astrocytomas that are destined for malignant progression has a significant impact on patient management.

For each patient, we analyzed only 1 MRS study performed at the time of diagnosis or close to diagnosis. This avoided emphasizing results from individual more frequently examined patients. Additional analyses (not reported in detail) showed that pooling all studies or analyzing only the latest available study for each patient would not alter any of the reported findings.

Variability of Citrate in AA and Glioblastomas

Prominent citrate was not a specific feature of tumors with poor prognoses. Indeed, among the 13 patients with AA included in this study, there was no evidence for citrate in three patients. On the other hand, of all patients, the tumor with the highest citrate concentration was among those with AA. Because all of the MRS studies of AA were performed at initial diagnosis or within the first 3 months after diagnosis, it is unlikely that the timing of the study had an impact on this finding. A more likely scenario is that the considerable
metabolic heterogeneity of AA is a reflection of histological and genetic subclasses of these tumors. Tumors with prominent citrate may constitute a metabolic subclass among AA. The averaged GBM spectrum was not obtained at diagnosis. The patient subsequently underwent shunt surgery, and a biopsy of the lesion was performed (grade II diffuse astrocytoma). MRI at three months demonstrated reduced size of the ventricles and resolved edema. In the spectrum obtained at that time of the tumor, N-acetyl-aspartate (NAA), creatine (Cr), choline (Cho), and myo-inositol (mI) were readily detectable. There was no evidence for citrate. Shown are the unfiltered raw data (thin grey line) and the fit to the data used for quantitation (black line). The box on the FLAIR MR image (B) indicates the region of interest from where the spectrum was acquired. This patient is alive and doing well 5 years after initial diagnosis.

Radiographic Features of Astrocytomas with High Citrate Levels

There was no apparent association between citrate levels and contrast enhancement or the location of a tumor. However, there was a positive correlation between citrate concentrations and the ADCs. Because diffusion imaging is sensitive to the extracellular space, this observation indicated a correlation of citrate concentrations with the amount of interstitial fluid and swelling. There is a correlation between ADC and the T2 relaxation time of tissue. This means that tumors with...
higher citrate concentrations were generally brighter on T2-weighted MRIs. The finding of a positive correlation between ADC and citrate may be counterintuitive, because one might expect lesions with higher cellularity (ie, lower ADC) to be more aggressive. However, this observation of an inverse correlation between ADC and aggressiveness was made in adults with glioblastoma, and it is not generally applicable in pediatrics. For example, pediatric pontine gliomas are characterized by high initial ADC values (and high citrate concentrations), despite having the worst outcomes of all tumors in pediatric neuro-oncology.

What Causes Citrate Accumulation?

It is unclear which mechanisms caused increased citrate in some of the tumors. Recently, mutations of the isocitrate dehydrogenase 1 gene (IDH1) have been detected in human gliomas. IDH1 mutations are considered to be early events in the progression of a tumor, preceding other genetic mutations associated with gliomas, and have been linked with upregulation of processes (eg, angiogenesis) that promote tumor growth. Although the exact impact of IDH1 mutations is unclear, there is evidence that they can cause an inhibition of the catalysis of isocitrate to α-ketoglutarate. Because the conversion of citrate to isocitrate is reversible, citrate may potentially accumulate in tissue with impaired IDH1 activity. IDH1 mutations were most frequently observed in secondary GBM (73%–88%), evolving from low-grade astrocytomas. IDH1 mutations were also frequently observed in grade II diffuse astrocytoma (59%–88%), and anaplastic astrocytoma (50%–78%), whereas there is a very low incidence in primary GBM (3%–16%) and in pediatric GBM (0%–7%). In this context, we noted that, for 3 of our aggressive LGAs, transformation to higher-grade astrocytoma was confirmed by biopsies performed at disease progression. No data were available for the remaining patients in this group.

If IDH1 mutations cause or correlate with citrate accumulation, one would expect studies that report the in vivo detection of citrate in adults with secondary GBM, grade II and III astrocytomas but not for primary GBM. There have been, to the best of our knowledge, no such reports. There are 3 possible explanations for the lack of detection of citrate in astrocytomas in adults. First, there might be no such correlation. Second, IDH1 mutations might not be

<table>
<thead>
<tr>
<th>Table 2. Absolute concentrations (mmol/kg tissue) and metabolite concentration ratios relative to creatine level (mean ± standard deviation) of pediatric astrocytomas</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>N</strong></td>
</tr>
<tr>
<td>-------</td>
</tr>
<tr>
<td>All LGA</td>
</tr>
<tr>
<td>Indolent LGA</td>
</tr>
<tr>
<td>Astrocytomas with malignant progression</td>
</tr>
<tr>
<td>Aggressive LGA</td>
</tr>
<tr>
<td>AA</td>
</tr>
</tbody>
</table>

For aggressive versus indolent LGA, *P < .01, **P < .001, and ***P < .0001.

aLactate concentrations are reported in mmol/L.
bGLX = [glutamate] + [glutamine].
cAbsolute intensity (arbitrary units).
dIn mm²/s × 10⁻³.
sufficient to cause citrate accumulation in adults with astrocytoma. Other mechanisms, such as increased activity of ATP citrate lyase, the enzyme that breaks down cytosolic citric, could compensate for any effect IDH1 mutations might have on citrate levels. Third, citrate might have been overlooked in previous MRS studies, because its detection is technically demanding and investigators have not looked for citrate. It is noteworthy that, in a publication by Howe et al, who used a similar short echo time single-voxel MRS approach as used in our studies, a figure shows peaks consistent with citrate for adult grade II and grade III astrocytoma, but not for GBM. Ultimately, studies that correlate IDH1 mutations with in vivo citrate levels are required to confirm a possible link.

The metabolism of tumors may also be altered due to physiological conditions. Increased interstitial fluid pressure (IFP) is a feature of many tumors and is an independent predictor of inferior outcomes, because it may prevent the uptake of macromolecular anti-cancer agents. However, the mechanism that would relate increased IFP and increased citrate is unclear. Another abnormal physiological condition postulated for tumors is hypoxia. Under hypoxic conditions, tumors increase glycolytic energy production and decrease mitochondrial function, which may result in the observed accumulation of citrate. Hypoxic tumors have been associated with poor prognoses, because they show increased angiogenesis and local invasion, as well as increased formation of distant CNS metastases. They are more likely to transition to phenotypes that are more malignant. Hypoxic tumors are also considered more resistant to therapy.

**Other Metabolic Features of Indolent LGA and Aggressive LGA**

NAA was also significantly differed between aggressive LGA and indolent LGA, albeit at a much lower level.
than citrate. Because NAA is present in high concentrations only in normal neurons and axons, low NAA in aggressive LGA likely indicated a small partial volume of normal tissue within those tumors. Among other chemicals detectable by MRS, choline, lactate, and lipids have been of particular interest in tumor spectroscopy. Choline-containing compounds are involved in cell membrane synthesis and breakdown and are abnormal in all forms of cancer. Lactate is an indicator of abnormal glycolytic function, but it can also accumulate in necrotic tissue, whereas lipid levels are believed to be elevated in aggressive, fast-growing tumors that are outgrowing their blood supply and causing necrosis in insufficiently perfused tumor tissue. These metabolic features were not significantly different in aggressive LGA and indolent LGA in our study, which focused on magnetic resonance spectra acquired at an early stage of the disease. However, there is ample evidence in the literature that prominent choline, lactate, and lipids in spectra are indicators for malignant tumors. In our study, these quantities progressively increased in aggressive LGA, when serial studies were available (cf. Fig. 3) consistent with what has been observed in previous publications, whereas no significant changes were observed for indolent LGA. These measures are valuable for monitoring disease status, because they can often detect progression prior to clinical or radiological progression.

Limitations
The number of patients included in this study was small. In addition, data were retrospectively evaluated, so there is a higher risk of introducing a patient selection bias, although efforts were made to minimize this effect. Single-voxel MRS was used because this methodology provided robust and good-quality spectra in a timely fashion. On the other hand, single-voxel MRS is less practical for assessment of the heterogeneity of lesions, planning of focal treatments (such as radiation or surgical resection), and identification of areas of response to therapy and disease progression. We have conducted a limited number of multi-voxel studies of large bi-thalamic low-grade astrocytomas (not reported in detail) and noted, consistent with the positive correlation of citrate levels with ADC, that citrate levels are higher in the more edematous-appearing parts (ie, high ADC) of the lesion. Although these preliminary observations cannot be generalized at this stage, this is nevertheless a clear indication of the possibility of sampling errors in individual patients.

Conclusions
An outcome-based comparison of grade II astrocytomas revealed that lesions with aggressive behavior within 2 years after diagnosis had significantly higher levels of citrate than did stable tumors. Prominent citrate may be an early sensitive indicator for pediatric grade II astrocytomas destined for aggressive behavior and, thus, useful for noninvasive patient stratification. Citrate was not specific for poor outcome, because it was not detectable in a significant portion of the high-grade astrocytomas. Among high-grade astrocytomas, tumors with prominent citrate may constitute a metabolic subclass.

Acknowledgments
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Conflict of interest statement. None declared.

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