Phenotypes Associated with Down-Regulation of \textit{Sl-IAA27} Support Functional Diversity Among Aux/IAA Family Members in Tomato

Carole Bassa$^{1,2}$, Isabelle Mila$^{1,2}$, Mondher Bouzayen$^{1,2}$ and Corinne Audran-Delalande$^{1,2,*}$

$^{1}$Université de Toulouse, INP-ENSA Toulouse, Génomique et Biotechnologie des Fruits, Avenue de l’Agrobiopole BP 32607, Castanet-Tolosan F-31326, France
$^{2}$INRA, Génomique et Biotechnologie des Fruits, 24 Chemin de Borde Rouge, Auzelles, CS 52627 F-31326, France

*Corresponding author: E-mail, delaland@ensat.fr; Fax: +33-534-32-38-73.
(Received May 15, 2012; Accepted June 28, 2012)

The phytohormone auxin is known to regulate several aspects of plant development, and Aux/IAA transcription factors play a pivotal role in auxin signaling. To extend our understanding of the multiple functions of Aux/IAAs further, the present study describes the functional characterization of \textit{SI-IAA27}, a member of the tomato Aux/IAA gene family. \textit{SI-IAA27} displays a distinct behavior compared with most Aux/IAA genes regarding the regulation of its expression by auxin, and the \textit{SI-IAA27}-encoded protein harbors a unique motif of unknown function also present in \textit{SI-IAA9} and remarkably conserved in monocot and dicot species. Tomato transgenic plants underexpressing the \textit{SI-IAA27} gene revealed multiple phenotypes related to vegetative and reproductive growth. Silencing of \textit{SI-IAA27} results in higher auxin sensitivity, altered root development and reduced Chl content in leaves. Both ovule and pollen display a dramatic loss of fertility in \textit{SI-IAA27} down-regulated lines, and the internal anatomy of the flower and the fruit are modified, with an enlarged placenta in smaller fruits. In line with the reduced Chl content in \textit{SI-IAA27} RNA interference (RNAi) leaves, genes involved in Chl synthesis display lower expression at the level of transcript accumulation. Even though \textit{SI-IAA27} is closely related to \textit{SI-IAA9} in terms of sequence homology and the encoded proteins share common structural features, the data indicate that the two genes regulate tomato fruit initiation and development in a distinct manner.

**Keywords:** Auxin • Aux/IAA • Chl • Fruit • Tomato.

**Abbreviations:** AFB, auxin receptor F-box; ARF, auxin response factor; Aux/IAA, auxin/IAA; AuxRE, auxin responsive cis-element; CaMV, Cauliflower mosaic virus; Chl, chlorophyll; EAR, ethylene-responsive element-binding factor-associated amphiphilic repression; GFP, green fluorescent protein; GH3, Gretchen Hagen 3, GUS, β-glucuronidase; MS medium, Murashige and Skoog medium; NAA, naphthalene-1-acetic acid; NLS, nuclear localization signal; NPA, N-1-naphthylphtalamic acid; qRT–PCR, quantitative reverse transcription–PCR; RNAi, RNA interference; SAUR, small auxin up RNA; SCF, SKP1-Cullin-F-box; SGN, Solanaceae Genomics Network; SI-IAA, Solanum lycopersicum auxin/IAA; TIR1, transport inhibitor response1; TPL, toplless; YFP, yellow fluorescent protein.

**Introduction**

The phytohormone auxin (IAA) plays a determinant role in plant development by notably controlling cell division, expansion and differentiation and by regulating organ initiation, embryogenesis and root development (Vanneste and Friml 2009). Auxin is also fundamental for successful fertilization of the flower and for fruit initiation and subsequent growth (Gillaspy et al. 1993, Vivian-Smith and Koltunow 1999, Carmi et al. 2003, Rotino et al. 2005). Components of the auxin signaling pathway have been shown to be involved in repressing fruit initiation until the fertilization cue (Vivian-Smith et al. 2001, Wang et al. 2005, Goetz et al. 2006, Pandolfini et al. 2007). Pioneering studies on Arabidopsis have identified different components of auxin signaling, among which the Aux/IAA family of transcriptional regulators (Abel et al. 1995). Aux/IAA genes constitute one of the three major groups of primary auxin-responsive genes which also include \textit{Small Auxin Up RNA} (SAUR) and \textit{Gretchen Hagen} 3 (GH3) (Theologis et al. 1985, Oeller et al. 1993). Aux/IAA genes encode short-lived proteins that typically share four conserved domains (Reed 2001) and display the ability to function as transcriptional repressors due to a conserved leucine motif (LxLxLx) located in domain I (Tiwari et al. 2004) similar to the so-called EAR (ethylene-responsive element binding factor-associated amphiphilic repression) repression domain (Kagale et al. 2010, Kagale and Rozwadowski 2011). It is well acknowledged now that Aux/IAA genes have a dual activity, one as a transcriptional regulator and another as...
The presence of auxin promotes the association of Aux/IAAs with the SKP1-Cullin-F-box (SCF) complex through binding to the auxin transporter inhibitor response1 (TIR1) or to its paralogs AUXIN RECEPTOR F-BOX (AFB) proteins. The SCF complex targets Aux/IAAs to the proteasome, leading to their rapid degradation (Dharmasiri et al. 2005a, Dharmasiri et al. 2005b, Kepinski and Leyser 2005, Leyser 2006, Tan et al. 2007, Chapman and Estelle 2009). The degradation of Aux/IAAs results in the release of ARFs which can then activate the transcription of target genes via binding to the auxin responsive elements (AuxREs) present in the promoter regions of auxin-regulated genes (Hagen et al. 1991, Ulmasov et al. 1997, Hagen and Guilfoyle 2002). In Arabidopsis, the unraveling of Aux/IAA functions in planta was almost exclusively achieved from the characterization of gain-of-function mutants. Indeed, phenotypes associated with loss-of-function mutations show an important functional redundancy among Aux/IAA family members (Rouse et al. 1998, Tian and Reed 1999, Naggal et al. 2000, Rogg et al. 2001, Fukaki et al. 2002, Tatematsu et al. 2004, Overvoorde et al. 2005, Uehara et al. 2008). In contrast, down-regulation of Aux/IAA genes in tomato proved to be, so far, quite efficient in revealing the physiological significance of a number of Aux/IAAs (Wang et al. 2005, Chaabouni et al. 2009a, Deng et al. 2012). Therefore, this Solanaceae reference species is a suitable system for further deciphering the specific roles of various members of the Aux/IAA gene family made up of 25 genes (Audran-Delalande et al. 2012). Using a reverse dark-grown seedlings in the absence of ethylene, thus supporting the hypothesis that SI-IAA9 and SI-IAA8, these three proteins forming a distinct clade, named clade B, in the tomato Aux/IAA protein family (Audran-Delalande et al. 2012). To shed more light on this clade B, members were compared with those from the same clade in potato (SGN database: Solanaceae Genomics Network, http://www.sgn.cornell), poplar (Kalluri et al. 2007), rice (Jain et al. 2006) (DRTB database http://drtb.cbi.pku.edu.cn/index.php) and maize (Wang et al. 2010b). Alignment of amino acid sequences of clade B Aux/IAAs revealed the presence in IAA9 and IAA27 of a conserved motif (YxGLS) before domain I in all monocot and dicot species investigated. Notably, the YxGLS motif was also found in Arabidopsis At-IAA8 while it is absent in IAA8 proteins from all other species (Fig. 1A). Because this motif is not found in any other Aux/IAA proteins, it provides a specific signature of clade B.

The SI-IAA27 coding sequence is 837 bp long and made up of five exons like its closest Arabidopsis homolog (Fig. 1B). The derived protein contains 278 amino acids. At the nucleotide and amino acid sequence levels, SI-IAA27 displays 67% and 52% identity, respectively, with its putative Arabidopsis ortholog At-IAA27. The four conserved domains (I–IV) characteristic of the Aux/IAA family (Abel et al. 1995) are present in SI-IAA27 protein as well as putative nuclear localization signals (NLSs) with both the bipartite structure of a conserved basic doublet KR between domains I and II and basic amino acids in domain II, and the SV40-type NLS located in domain IV (Fig. 1B). The subcellular localization of the SI-IAA27 protein was assessed by transient expression assay in tobacco protoplasts using a translational fusion between SI-IAA27 and yellow fluorescent protein (YFP) under the control of the 35S promoter of Cauliflower mosaic virus (CaMV). Microscopy analysis showed that in contrast to control protoplasts transformed with YFP alone which displayed fluorescence throughout the cell, the YFP–SI-IAA27 fusion was only localized in the nucleus (Fig. 2A). These data shows that SI-IAA27 was able to direct the fusion protein to the nucleus, consistent with its putative transcriptional regulatory function. To gain insight into the spatial pattern of expression of the SI-IAA27 gene, accumulation of its transcript was assessed in different plant tissues and organs by quantitative reverse transcription–PCR (qRT-PCR) and by analysis of expression of the β-glucuronidase (GUS) reporter gene driven by the SI-IAA27 promoter. These data revealed that...
Sl-IAA27 was expressed in all organs analyzed, with the lowest expression found in leaves (Fig. 2B; Supplementary Fig. S1). In the flower, Sl-IAA27 was expressed in sepals, anthers and in the top of the stamen, but not in petals (Fig. 2B; Supplementary Fig. S1D). The expression was high in the ovary at the anthesis stage (Supplementary Fig. S1F). In mature green fruit, the expression was mainly observed in pericarp, endocarp and columella (Supplementary Fig. S1G), whereas it was down-regulated during ripening with strongly reduced expression in red fruits (Fig. 2B; Supplementary Fig. S1G, H). In hypocotyls and roots, Sl-IAA27 is expressed in vascular tissues (Supplementary Fig. S1A–C).

**Sl-IAA27 transgenic plants showed altered root development and modified auxin sensitivity**

Both RNA interference (RNAi) and overexpression approaches were used to address the physiological significance of Sl-IAA27 protein. Several independent homozygous lines were generated; three Sl-IAA27-overexpressing lines and three Sl-IAA27 RNAi lines were selected for further studies. The Sl-IAA27-overexpressing lines were named Sline1, Sline2 and Sline3, and the Sl-IAA27 RNAi lines were named Rline1, Rline2 and Rline3. Analysis by real time-PCR showed that Sl-IAA27 is overexpressed in the overexpressing transgenic lines with transcript levels 120 times higher than the wild type in Sline1, and up to 50 times higher in Sline2 and Sline3 (Supplementary Fig. S2). In the three RNAi lines, real-time PCR experiments showed substantially lower accumulation of Sl-IAA27 transcripts compared with the wild type. With less than one-third of the mRNA level displayed in the wild type, Rline1 showed the highest level of repression, whereas Rline2 and Rline3 retained half of the normal mRNA levels (Fig. 2C). To check the specificity of the RNAi strategy and rule out any potential interference with other Aux/IAA genes, the expression of clade B members SI-IAA8 and SI-IAA9 was assessed. No reduction in the transcript levels of SI-IAA8 and SI-IAA9 genes was detected, indicating that the altered physiological processes observed in the transgenic lines are primarily due to the down-regulation of Sl-IAA27. Moreover, transcript accumulation of Sl-IAA9, the most closely related Aux/IAA in terms of sequence homology, was even higher in Sl-IAA27 RNAi lines than in wild-type plants (Fig. 2C).

Primary roots were 23–43% more elongated in Sl-IAA27 RNAi lines than in wild-type plants, and the increase in root length correlated with the level of transcript reduction in the three lines (Fig. 3A, B). In addition, Sl-IAA27 RNAi transgenic lines displayed increased lateral root formation (Fig. 3A, C). In contrast, Sl-IAA27-overexpressing lines presented a reduced root length, the main root being up to 50% less elongated than in the wild type and producing no lateral root (Supplementary Fig. S3A, B).

The altered root growth in the transgenic plants suggested that the modification of Sl-IAA27 expression may result in altered auxin sensitivity. The physiological auxin response of Sl-IAA27 RNAi and overexpressing plants was assessed using the auxin transport inhibitor N-1-naphthylphthalamic acid (NPA) known to alter the endogenous auxin gradients notably in roots. Lateral root formation was inhibited in both wild-type and RNAi plants. Primary root growth was reduced in wild-type, Sl-IAA27-overexpressing and RNAi plants with, however, a much lower reduction in the Sl-IAA27 RNAi lines (Fig. 4A, B;
Supplementary Fig. S3C). Due to the difference in NPA sensitivity of Sl-IAA27 RNAi roots, the auxin sensitivity of Sl-IAA27 RNAi plants was further explored by determining the auxin dose–response on elongation of hypocotyl segments. Fig. 4C shows that independently of the auxin concentration used, the Sl-IAA27 RNAi hypocotyls were always more elongated than wild-type hypocotyls. After 24 h of auxin treatment, maximum hypocotyl elongation was obtained with a 10^{-5} M NAA (naphthalene-1-acetic acid) concentration in the transgenic lines; while this was within the range of auxin concentrations used in the experiment, it was not reached for the wild type. These data indicate that down-regulation of Sl-IAA27 confers higher auxin sensitivity to the transgenic plants at the level of hypocotyl elongation.

Because Aux/IAA genes have been reported to be up-regulated by auxin (Abel et al. 1994), the auxin responsiveness of the Sl-IAA27 gene was investigated by qRT–PCR in seedling tissues. The data in Fig. 4D clearly indicate that Sl-IAA27 was down-regulated by auxin treatment. Moreover, using a transient expression experiment in the tobacco protoplast system, the activity of the Sl-IAA27 promoter fused to green fluorescent protein (GFP) showed no auxin-induced regulation compared with the DRS synthetic promoter (Fig. 4E). In silico analysis of 2,000 bp of the Sl-IAA27 promoter sequence using PLACE software (http://www.dna.affrc.go.jp/PLACE/index.html) revealed the presence of a variety of conserved motifs, but none of these putative cis-elements corresponded to the AuxREs known to drive auxin responsiveness. Strikingly, up to 41 conserved motifs putatively involved in light regulation were found, including a GT1 motif (S000198 GRWAAW), one GT1CORE motif (S000125 GGTTAA) and 17 GATA motifs (S000039 GATA).

SI-IAA27 RNAi plants showed a lower Chl content

SI-IAA27 RNAi leaves showed a pale green color more dramatically visible during the first weeks of plant development. In contrast to the green leaves in wild-type plants, transgenic lines presented yellow to light green leaves depending on the level of Sl-IAA27 down-regulation, with a more marked phenotype in Rline1 (Fig. 5A). Chl a and b content measured by spectrophotometry showed a significant reduction in leaves of the transgenic plants compared with wild-type plants, in line with their light green color (Fig. 5C). Notably, the lower level of Chl observed in the RNAi leaves is not related to a diminution of chloroplast number. Indeed, microscopic observation of leaves from the wild type and Rline1 showed that...
**Fig. 3** Altered root growth in *Sl-IAA27* RNAi lines. (A) Root development in the wild type and *Sl-IAA27* RNAi lines assessed in 3-week-old seedlings grown on MS/2 medium. (B) The mean primary root length in the wild type and *Sl-IAA27* down-regulated lines. (C) Lateral root number per cm of root length in the wild type and *Sl-IAA27* RNAi lines. Statistical analysis were realized using the Student’s test, ***P < 0.001; **P < 0.01; *P < 0.05. *Sl-IAA27* RNAi lines = Rline1, Rline2 and Rline3.

**Fig. 4** Altered auxin response in *Sl-IAA27* down-regulated lines. (A) Effect of NPA (5 µM) treatment on root development of 3-week-old wild type (WT) and RNAi tomato lines grown on MS/2 medium. (B) Primary root length of WT and RNAi lines assessed on seedlings treated with NPA. (C) Auxin dose–response of elongation of hypocotyl segments treated with NAA. Hypocotyl segments 9 mm long were cut from 8-day-old seedlings and treated for 24 h with the indicated concentration of NAA. Data represent mean values obtained with three replicates for each of the three *Sl-IAA27* RNAi lines. (D) Auxin regulation of *Sl-IAA27* expression. *Sl-IAA27* transcript accumulation was monitored by qRT–PCR in seedlings treated for 2 h with 20 µM IAA. (E) Auxin responsiveness of the *Sl-IAA27* promoter. GFP fluorescence of protoplasts transformed either with the synthetic promoter DR5 or the *Sl-IAA27* promoter fused to GFP and treated or not with 2,4-D (50 µM). Statistical analyses were realized using the Student’s test, ***P < 0.001; **P < 0.01; *P < 0.05. *Sl-IAA27* RNAi lines = Rline1, Rline2 and Rline3.
chloroplast number was not modified but that Chl fluorescence was dramatically reduced as assessed by fluorescence emission (Fig. 5B). In Sl-IAA27-overexpressing plants, no visible difference was observed in leaf color compared with the wild type. Analysis of Chl a and b content revealed that the Chl content is not modified by the overexpression of Sl-IAA27 in leaves (Supplementary Fig. S4A).

The lower Chl content in Sl-IAA27 RNAi leaves was correlated with the down-regulation of genes involved in key steps of Chl biosynthesis. In particular, transcript accumulation of HEMA1, protoclorophyllide reductase a, b, c and chelatase subunit chlh and chli genes displayed significantly lower levels than the wild type, while those corresponding to aminolevulinate dehydratase and chelatase subunit chld were not affected. Moreover, transcript accumulation of GUN4, a positive regulator of Chl biosynthesis (Davison et al. 2005, Peter and Grimm 2009, Adhikari et al. 2011), was also significantly reduced in the transgenic lines (Fig. 5D; Supplementary Fig. 5S, Supplementary Table S1). Down-regulation of genes related to Chl biosynthesis was still observed in Sl-IAA27 RNAi plants in flowers at the anthesis stage and in young fruits (7 d after anthesis) (Fig. 6A, B). Analysis of the 2,000 bp promoter sequence of the Chl biosynthesis genes analyzed above showed the presence of AuxREs in all corresponding promoters except for aminolevulinate dehydratase, chelatase subunit chli and GUN4, thus suggesting a putative auxin regulation of most of these genes.

To check whether the down-regulation of Chl biosynthesis-related genes results from the higher sensitivity to auxin of Sl-IAA27 RNAi plants, the auxin regulation of these genes was assessed at the transcriptional level in wild-type plants. None of these genes showed down-regulation by auxin and, in fact, HEMA1 and protoclorophyllide a genes rather were up-regulated (Supplementary Fig. 6S). These data suggest that down-regulation of Chl biosynthesis-related genes in the Sl-IAA27 RNAi lines is not directly induced by higher auxin responsiveness of the plants.

**Sl-IAA27 silencing resulted in altered fruit development and reduced fertilization**

The effect of Sl-IAA27 silencing on fruit development was investigated in the three Sl-IAA27 RNAi lines. While the leaves of Sl-IAA27 RNAi plants display a pale green color compared with the wild type, the fruit color is not modified at any stage of fruit development and ripening (Supplementary Fig. S7). However, down-regulation of Sl-IAA27 resulted in alteration of fruit morphology, with the formation of fruits with a modified shape and reduced size. Indeed, Sl-IAA27 RNAi fruits displayed a lower fruit width but normal fruit length (Fig. 6C). Fruit volume was dramatically reduced in Sl-IAA27 RNAi lines compared with wild-type plants (Fig. 6D). Modification of Sl-IAA27 RNAi fruit morphology was correlated with altered flower anatomy. Observation of an Sl-IAA27 RNAi ovary by microscopy
revealed a modified shape with enlarged placenta and lower number of ovules compared with the wild type (Fig. 7A). This altered anatomy structure was maintained in the developed fruits (Fig. 7B). Ovule numbers showed a dramatic reduction in Rline1, with an average of 15 ovules per ovary compared with 24 in the wild type (Fig. 7C). A decrease in seed number was also observed in Sl-IAA27 RNAi lines that was, however, more severe than the reduction in ovule number. Indeed, only 27% of Rline1 ovules developed seeds (Fig. 7C, D). To check whether in addition to the reduction in number, the ovules also have altered fertility in the transgenic lines, cross-fertilization was performed using transgenic lines as the female recipient and the wild type as the pollen donor. The data presented in the upper part of Table 1 clearly show that wild-type pollen is unable to increase the seed number, suggesting that ovule fertility is reduced in Sl-IAA27 RNAi. Moreover, reciprocal crossing using the wild type as the female recipient also revealed a loss of pollen in the transgenic lines (Table 1). Overall, these data indicated that both ovules and pollen display reduced fertility in Sl-IAA27 RNAi. Fruits of Sl-IAA27-overexpressing plants displayed no modification in fruit shape or volume, and presented no reduction in seed number (Supplementary Fig. S4B, C).

**Discussion**

Most of our knowledge of the physiological significance of Aux/IAA genes comes from the characterization of gain-of-function mutants in Arabidopsis (Rouse et al. 1998, Tian and Reed 1999, Nagpal et al. 2000, Rogg et al. 2001, Fukaki et al. 2002, Tatematsu et al. 2004, Uehara et al. 2008). In Solanaceae species and particularly in tomato, a wide range of phenotypes associated with down-regulation of Aux/IAA genes have been reported (Wang et al. 2005, Kloosterman et al. 2006, Chaabouni et al. 2009a, Deng et al. 2012), but the role of most tomato Aux/IAA genes still remains to be elucidated. While aiming to illuminate further the multiple functions of Aux/IAA genes, the present study more specifically deals with the functional characterization of Sl-IAA27 for which information regarding the putative function is lacking, in the Solanaceae as well as for its ortholog in Arabidopsis. Silencing of Sl-IAA27 results in pleiotropic morphological and developmental phenotypes including altered fruit development and Chl accumulation. In these Sl-IAA27 RNAi lines, both ovule and pollen fertility are reduced, indicating that normal expression of Sl-IAA27 is required for flower fertility. Furthermore, Sl-IAA27 down-regulated lines showed a dramatic reduction in ovule formation which, associated with reduced fertility, leads to the production of seedless fruits. Compared with the wild type, the fruit shape and internal fruit structure are also altered, with an enlarged placenta and smaller final fruit size. In Sl-IAA27-overexpressing plants, no phenotypes were observed regarding the Chl content in leaves or fruit development, suggesting that even if the Sl-IAA27 mRNA level is high, its protein level is probably regulated by auxin-mediated degradation, restoring its level to that in the wild type.
Previous studies showed that down-regulation of either Sl-IAA3, Sl-IAA9 or Sl-IAA15 results in modified auxin sensitivity associated with altered leaf structure and modified plant architecture (Wang et al. 2005, Chaabouni et al. 2009a, Deng et al. 2012). In contrast, the higher auxin sensitivity of Sl-IAA27 down-regulated plants did not result in visible alteration of the plant architecture. The transgenic lines display altered primary root growth and lateral root formation. Interestingly, overexpressing and RNAi plants displayed opposite phenotypes, with reduced primary root growth and no lateral root formation in overexpressing lines and increased primary root length with more lateral roots in RNAi lines. This suggests that in roots of overexpressing plants, auxin-dependent degradation of Sl-IAA27 is probably not able to restore protein levels similar to the wild type.

In the presence of the auxin transport inhibitor NPA, Sl-IAA27 RNAi roots displayed no lateral roots, suggesting that NPA is still able to block transverse divisions as in the wild type (Casimiro et al. 2001). Nevertheless, though root elongation was reduced upon NPA treatment in Sl-IAA27 RNAi lines, the root length remains more important than in the wild type subjected to the same treatment. Root elongation is controlled by a specific auxin concentration balance, and auxin is known to play a predominant role in root development mainly through controlling cell division (Friml et al. 2002, Ullah et al. 2003, Ding and Friml 2010, Dubrovsky et al. 2011). In Arabidopsis, characterization of several Aux/IAA mutants (CRANE/IAA18, SHY2/IAA3, AXR2/IAA7 and SLR/IAA14) revealed the function of these genes in regulating the root architecture (Nagpal et al. 2000, Vanneste et al. 2005, Uehara et al. 2008). NPA application was reported to lead to IAA accumulation in the root tip by blocking auxin basipetal transport in roots (Casimiro et al. 2001). The more elongated primary root in Sl-IAA27 RNAi plants upon NPA treatment may result from the difference either in auxin concentration or in auxin sensitivity of transgenic lines compared with the wild type. Analysis of the auxin dose–response of hypocotyl elongation where endogenous auxin was removed showed that transgenic plants were more sensitive to NAA. These data indicate that for a specific auxin concentration, Sl-IAA27 RNAi plants present higher auxin sensitivity, suggesting that Sl-IAA27 protein acts as a repressor of auxin responses. In line with this hypothesis,
previous data showed that Sl-IAA27 represses the auxin-responsive transcription of both synthetic and native auxin-responsive promoters (Audran-Delalande et al. 2012).

In contrast to classical Aux/IAA genes characterized by a rapid induction of their expression in response to auxin (Walker and Key 1982, Theologis et al. 1985), Sl-IAA27 expression is down-regulated by exogenous auxin treatment. At-IAA27 and Sb-IAA21, the Sl-IAA27 homologs from Arabidopsis and Sorghum, respectively, are also down-regulated by auxin (Paponov et al. 2008, Wang et al. 2010a), suggesting that this feature might be conserved across plant species. Previous analysis of the auxin regulation of tomato Aux/IAA genes with short-term treatment has shown that Sl-IAA8 is also down-regulated by auxin and that Sl-IAA9 expression is not significantly responsive to auxin (Audran-Delalande et al. 2012). These data indicate that members of clade B of the Aux/IAA gene family (Sl-IAA8, Sl-IAA9 and Sl-IAA27) behave differently from other members regarding their regulation by auxin. Recently, phylogenetic analysis of the tomato Aux/IAA family revealed that Sl-IAA9 is closely related to Sl-IAA27 (Audran-Delalande et al. 2012), raising the hypothesis that protein structure similarity may result in functional redundancy. Beside domains I–IV which are characteristic of the Aux/IAA family (Abel et al. 1995), our study identified another conserved motif (YxGLS) located before domain I in the N-terminal part. It is noteworthy that this motif, present in both Sl-IAA27 and Sl-IAA9, is not found in any other Sl-IAA and is remarkably conserved in monocot and dicot species, including Arabidopsis, potato, poplar, maize and rice. Whether the conserved YxGLS motif determines some of the functional roles of Sl-IAA27 and Sl-IAA9 remains to be elucidated in planta. Despite their common structural features, the two genes seem to have distinct functions in plant developmental processes, as indicated by the contrasting phenotypes resulting from the down-regulation of Sl-IAA27 and Sl-IAA9. Indeed, down-regulation of Sl-IAA9 leads to altered leaf morphogenesis (Wang et al. 2005) while such a phenotype is absent in Sl-IAA27 underexpressing lines. Also, down-regulation of Sl-IAA27 results in lower fertility of both ovule and pollen, while reduced expression of Sl-IAA9 did not affect this (Wang et al. 2005). Moreover, in contrast to Sl-IAA9 down-regulated plants where fruits display no morphological or size alteration, Sl-IAA27 RNAi fruits present a reduced size and altered shape. Taken together, these data suggest that normal expression of Sl-IAA27 is required for the fertilization process and for fruit development, whereas Sl-IAA9 is essential for triggering fruit initiation (Wang et al. 2005). Even though both Sl-IAA27 and Sl-IAA9 down-regulated lines yield seedless fruit, two different mechanisms seem to underlie this phenotype. This is further supported by the observed up-regulation of Sl-IAA9 expression in Sl-IAA27 down-regulated plants shown in Fig. 2C. Nevertheless, our data suggest a link between the two genes while revealing that the expression of Sl-IAA9 is regulated by Sl-IAA27, either directly or indirectly.

Even though the level of Sl-IAA27 expression is lowest in wild-type leaves, the phenotypes displayed by the RNAi lines indicate that a threshold level of Sl-IAA27 expression is necessary to promote Chl biosynthesis in leaves. On the other hand, the overexpression of Sl-IAA27 does not result in any increase in Chl content in leaves, suggesting that the basal Sl-IAA27 protein level in the wild type is sufficient to sustain maximum activity of Chl biosynthesis genes. This may explain the lack of effect on Chl content in the overexpressing lines. The reduced Chl content of Sl-IAA27 RNAi plants correlates with the down-regulation of many genes involved in Chl biosynthesis in leaves. Moreover, Sl-IAA27 seems to regulate at the transcriptional level the expression of the GUN4 gene which encodes a transcription factor known to activate the Chl biosynthesis pathway. The presence of AuxREs in the promoter of most of these genes suggests that they may potentially undergo direct regulation by auxin signaling. In wild-type plants, the expression of HEMA1 and ProtoA genes is up-regulated upon auxin treatment, while in Sl-IAA27 RNAi plants the expression of these two genes is down-regulated. To explain this apparent contradiction, it is possible to consider that these two genes are under competitive regulation by activator and repressor ARFs. Taking into account that auxin accumulation is known to lead to the degradation of the majority of Aux/IAA proteins which frees both activator and repressor ARFs (Dharmasiri et al. 2005a, Dharmasiri et al. 2005b, Kepinski and Leyser 2005, Leyser 2006, Tan et al. 2007, Chapman and Estelle 2009), it can be hypothesized that in normal situations the expression of genes related to Chl biosynthesis is down-regulated by one or more repressor ARFs that are trapped by Sl-IAA27 protein. These repressor ARFs are released in the absence of Sl-IAA27 in RNAi plants or upon auxin treatment. On the other hand, the release of activator ARFs in the presence of auxin may result in the up-regulation of HEMA1 and ProtoA genes observed in wild-type plants. Taken together, the data suggest a complex and finely tuned mechanism of regulation of auxin responses involving Aux/IAAs and ARFs.

The smaller size of the Sl-IAA27 down-regulated fruits may result from the reduced photosynthetic activity in the leaves. This hypothesis is supported by the prevailing opinion that fruit growth is predominantly supported by photoassimilate supply from source tissues (Lytovchenko et al. 2011). In keeping with the prominent role of photosynthetic activity in the leaves, down-regulation of Chl biosynthesis in a fruit-specific manner indicated that photosynthesis was essential for seed set but did not affect fruit growth (Lytovchenko et al. 2011). It is important to mention, however, that cells in developing fruit contain photosynthetically active chloroplasts and express both nuclear-encoded and plastid-encoded genes for photosynthetic proteins (Piechulla et al. 1987). It has been reported that during early fruit development, photosynthesis itself may provide a significant contribution to sustain the growth of the organ (Obiadalla-Ali et al. 2004). More recently, global transcriptomic profiling of fruit set and early fruit development also revealed a strong activation of photosynthesis-related genes.
were as follows: 14 h day/10 h night cycle, 25°C/20°C grown under standard greenhouse conditions. The conditions may contribute to sustain fruit growth, the relative contribution of the two source tissues to the needs of the fruit in terms of photoassimilate supply remains unclear.

Materials and Methods

Plant materials and growth conditions

Tomato plants (Solanum lycopersicum cv. MicroTom) were grown under standard greenhouse conditions. The conditions were as follows: 14 h day/10 h night cycle, 25°C/20°C day/night temperature, 80% relative humidity, 250 μmol m⁻² s⁻¹ light intensity (intense luminosity). Tomato seeds were first surface sterilized for 5 min in bleach and water, rinsed five times in sterile water and sown in recipient Magenta vessels containing 50 ml of 50% Murashige and Skoog (MS) culture medium and 0.8% (w/v) agar, pH 5.9.

Plant transformation

To generate Sl-IAA27 RNAi plants, the forward 5′-atgtctgtaccatg-3′ and reverse 5′-gttctgagtctcatgg-3′ primers were used to amplify the 155 bp Sl-IAA27 RNAi sequence. This fragment was cloned into the phellsgate12 vector using Gateway cloning. To generate Sl-IAA27-overexpressing plants, the forward 5′-atgtctgtaccatg-3′ and reverse 5′-ctaat-ttttgttcttgccg-3′ primers were used to amplify the 837 bp of Sl-IAA27 coding sequence. This fragment was cloned into the pMDC32 vector using Gateway cloning. To generate plAA27-GUS (the promoter of the Sl-IAA27 gene fused to the GUS reporter gene) plants, the forward 5′-catacaggatatgtgagtg-3′ and reverse 5′-tgctcaacctctc-3′ primers were used to amplify the 2,000 bp Sl-IAA27 promoter sequence. This fragment was cloned into the pMDC162 expression vector by Gateway cloning. Transgenic plants were generated by Agrobacterium tumefaciens-mediated transformation according to Wang et al. (2005). All experiments were carried out using homozygous lines from the F₂ and F₃ generation selected on kanamycin resistance.

Histochemical GUS analysis

For histochemical GUS analysis, tissues of plAA27–GUS lines were put in GUS staining solution (100 mM sodium phosphate buffer, pH 7.2, 10 mM EDTA, 0.1% Triton X-100, 0.3 mg ml⁻¹ X-Gluc); a vacuum was made for 15 min twice. Tissues were then incubated in GUS staining solution at 37°C overnight. Samples were then decolorated using several washes of graded ethanol series as indicated by Wang et al. (2005).

Transient expression using a single cell system

For nuclear localization of the Sl-IAA27 protein, the Sl-IAA27 CDS sequence was cloned by Gateway technology (forward 5′-atgtctgtaccatg-3′ and reverse 5′-gcaagaaccaaaattag-3′ primers) in-frame with YFP into the pEarlyGate104 vector, and expressed under the control of the 35S CaMV promoter. The empty vector pEarleyGate104 was used as control. For the analysis of Sl-IAA27 promoter regulation by auxin, the 2,000 bp of the Sl-IAA27 promoter sequence was fused to the GFP reporter gene by Gateway technology, into the pMDC107 vector. Protoplasts were obtained from suspension-cultured tobacco (Nicotiana tabacum) BY-2 cells and transfected according to the method described previously (Leclercq et al. 2005). YFP localization by confocal microscopy and GFP measurement by flux cytometry were performed as described previously (Audran-Delalande et al. 2012).

Hormonal treatment

For hypocotyl auxin dose–response (0, 1, 10 and 100 μM NAA) and NPA treatment, experiments were carried out as described by Chaabouni et al. (2009a). Auxin treatment of seedlings was performed as previously described by Audran-Delalande et al. (2012).

RNA isolation and real-time PCR analysis

RNA extraction, cDNA synthesis and qRT–PCR analyses were performed as previously described (Pirrello et al. 2006). The primer sequences are listed in Supplementary Table S2.

Determination of Chl content

A 100 mg aliquot of leaves from 3-week-old Sl-IAA27 RNAi and wild-type plants was weighed and ground with 1 ml of 80% acetone. The liquid obtained was centrifuged for 1 min at 10,000 r.p.m. to remove any remaining solid tissue. Samples were analyzed by spectrophotometry at two wavelengths, 645 and 663 nm, using 80% acetone as the blank. The Chl a and b content was determined using the following equations: Chl a = 0.999A₆₆₃ − 0.0989A₆₄₅ and Chl b = −0.328A₆₆₃ + 1.77A₆₄₅.

Microscopy analysis

For chloroplast observation, air was removed from leaves which were then analyzed with a Leica TCS SP2 confocal laser scanning microscope. Images were obtained with a ×40, 1.25 numerical aperture water-immersion objective. Excitation was at 488 nm and the emitted light was captured at 505–535 nm. For flower observation, flowers at the anthesis stage were fixed in ethanol and included in 5% agarose. Flowers were cut every 80 μm using a vibratome (vibratome Leica VT 100S) and then observed with a Leica DM IRBE inverted microscope.
Phenotypic analysis

The phenotypic analyses were performed on fruits of 10 plants from each transgenic line and from the wild-type plants cultivated in the same conditions. The inflorescence formation occurred at the same time in transgenic plants and in the wild type. Phenotypic analyses were performed on the six first fruits developed on each plant, which were similar with regard to their stage of development in transgenic plants and in the wild type (data not shown). In total, 60 fruits of the transgenic lines and the wild type were studied.

The experiment was done twice. Fruit color (Hue angle) was determined using a Chroma Meter CR-310 Minolta. Cross-pollination between SI-IAA27 RNAi and WT flowers was performed as described in Wang et al. (2005).

Sequence data

To identify homologs of tomato clade B, phylogenetic analysis were performed with all Aux/IAA members in maize, rice and poplar genomes, available, respectively, in the Maize GDB genome browser tool (http://gbrowse/maizegdb.org/gbrowse/cgi-bin/gbrowse/maize) and KOME database (http://www.cdna.01.dna.affrc.go.jp/cDNA/), and Populus trichocarpa Genome database (http://www.plantgdb.org/PtrGDB/). Both BLASTN and TBLASTN search were performed on the whole set of potato unigenes in the SGN database (Solanaceae Genomics Network, http://www.sgn.cornell) using Aux/IAA tomato clade B sequences.

Sequence data for the Arabidopsis genes used in this article can be found in the Arabidopsis Genome Initiative data library under the following accession numbers: AtIAA8 (AT2G22670), AtIAA9 (AT5G65670) and AtIAA27 (AT4G29080).

Sequence data for maize genes used in this article can be found in MaizeGDB genome browser tool (http://gbrowse/maizegdb.org/gbrowse/cgi-bin/gbrowse/maize) under the following accession numbers: ZmIAA2 (GRMZM2G077356_PO1), ZmIAA10 (GRMZM2G037369_PO1), ZmIAA14 (GRMZM2G077356_PO1), ZmIAA21 (GRMZM2G147243_PO2), ZmIAA28 (GRMZM2G035465_PO3) and ZmIAA29 (GRMZM2G163848_PO5).

Sequence data for the rice genes used in this article can be found in the KOME database (http://www.cdna.01.dna.affrc.go.jp/cDNA/) under the following accession numbers: OsIAA3 (AK104654), OsIAA5 (AK106121), OsIAA13 (AK059838), OsIAA19 (AK109363) and OsIAA21 (AK121989).

Sequence data for the poplar genes used in this article can be found in the P. trichocarpa genome database under the following accession numbers: PoptrIAA27.1.1 (POPTR_0001s18680.1), PoptrIAA27.2.1 (POPTR_0003s04980.1), PoptrIAA27.3.1 (POPTR_0006s16640.1) and PoptrIAA9 (POPTR_0002s10880.1).

Sequence data for the tomato genes used in this article can be found in Genbank/EMBL data libraries under the following accession numbers: Sl-IAA8 (JN379436), Sl-IAA9 (JN379437) and Sl-IAA27 (JN379450).

Supplementary data

Supplementary data are available at PCP online

Funding

This work was supported by the Laboratoire d’ Excellence (LABEX) [TULIP (ANR-10-LABX-41)]; the French government [grant for the degree of PhD of C.B.].

Acknowledgments

We thank O. Berseille and D. Saint-Martin for tomato culture, and the ‘plateforme de microscope TRI’ (Fédération de Recherches FR3450 Castanet-Tolosan) for microscopy analyses. The authors thank C. Pecher for his technical assistance and expertise in flow cytometry (FacsCalibur, RIO Imaging platform, INSERM UPS UMR 1048, Toulouse). The authors thank F.Regad for his help regarding experimental design and analysis.

References


Deng, W., Yang, Y., Ren, Z., Audran-Delalande, C., Mila, I., Wang, X.
et al. (2012) The tomato SLAAA15 is involved in trichome formation


Dharmasiri, N., Weijers, D., Yamada, M., Hobbie, L., Ehrismann, J.S.,
Jürgens, G. et al. (2005b) Plant development is regulated by a

Ding, Z. and Friml, J. (2010) Auxin regulates distal stem cell differen-
12046–12051.

Dubrovsky, J., Napsucialy-Mendivil, S., Duclercq, J., Cheng, Y.,
Shishkova, S., Ivanchenko, M.G. et al. (2011) Auxin minimum de-
fines a developmental window for lateral root initiation. New PhytoL

Friml, J., Benkova, E., Bilyou, I., Wisniewska, J., Hamann, T., Ljung, K. et al.
(2002) AtPIN4 mediates sink-driven auxin gradients and root pat-

formation is blocked by a gain-of-function mutation in the


AUXIN RESPONSE FACTOR8 is a negative regulator of fruit initi-


genesis, promoters and regulatory factors. Plant Mol. Biol. 49:
373–385.

expression of the soybean GH3 promoter in transgenic tobacco

(2006) Structure and expression analysis of early auxin-responsive
Aux/IAA gene family in rice (Oryza sativa). Funct. Integr. Genomics

Kagale, S., Links, M.G. and Rozwadowski, K. (2010) Genome-wide ana-
sis of ethylene-responsive element binding factor-associated
amphiphilic repression motif-containing transcriptional regulators

responsive element binding factor-associated amphiphilic repres-

Kagale, S. and Rozwadowski, K. (2011) EAR motif-mediated transcrip-
tional repression in plants: an underlying mechanism for epigenetic

Genome-wide analysis of Aux/IAA and ARF gene families in
Populus trichocarpa. BMC Plant Biol. 7: 59.


and characterization of a novel potato Auxin/Indole-3-Acetic
Acid family member (SIAA2) that is involved in petiole hyponasty
and shoot morphogenesis. Plant Physiol. Biochem. 44:
766–775.

Kolotilin, I., Koltai, H., Tadmor, Y., Bar-Or, C., Reuveni, M., Meir, A. et al.
(2007) Transcriptional profiling of high pigment-2dg tomato
mutant links early fruit plastid biogenesis with its overproduction

Leclercq, J., Ranty, B., Sanchez-Ballesta, M., Li, Z., Jones, B., Jauneau, A.
et al. (2005) Molecular and biochemical characterization of LeCRK1,
56: 25–35.

Leyser, O. (2006) Dynamic integration of auxin transport and signal-

Lytovchenko, A., Eickmeier, I., Pons, C., Osorio, S., Szecowka, M.,
Lehmburg, K. et al. (2011) Tomato fruit photosynthesis is seemingly
unimportant in primary metabolism and ripening but plays a con-

Müglen, P., Walker, L.M., Young, J.C., Sonawala, A., Timpte, C.,
Estelle, M. et al. (2000) AXR2 encodes a member of the Aux/IAA

Obidalia-Ali, H., Fernie, A.R., Lytovchenko, A., Kössmann, J. and
Lloyd, J.R. (2004) Inhibition of chloroplastic fructose
1,6-bisphosphatase in tomato fruits leads to decreased fruit size,
but only small changes in carbohydrate metabolism. Planta 219:
533–540.

Structural characterization of the early indoleacetic acid-inducible

Overvoorde, P., Okushima, Y., Alonso, J.M., Chan, A., Chang, C.,
Ecker, J.R. et al. (2005) Functional genomic analysis of the
AUXIN/INDOLE-3-ACETIC ACID gene family members in

Pandolfini, T., Molesini, B. and Spena, A. (2007) Molecular dissection of

Paponov, I., Paponov, M., Teale, W., Menges, M., Charkarbood, S.,
Murray, J.A.H. et al. (2008) Comprehensive transcriptome analysis

Pistor, E. and Grimm, B. (2009) GUN4 is required for posttranslational

Changes in photosynthetic capacity and photosynthetic protein
pattern during tomato fruit ripening. Plant Physiol. 84: 911–917.

Pirrello, J., Jaimes-Miranda, F., Sanchez-Ballesta, M.T., Tournier, B.,
Khali-Abhmad, Q., Regad, F. et al. (2006) SIEF2, a tomato ethylene
response factor involved in ethylene response and seed germin-


in IAA28 suppresses lateral root development. Plant Cell 13:
465–480.

Rotino, G.L., Acciarri, N., Sabatini, E., Mennella, G., Lo Scalzo, R.,
Maestrilli, A. et al. (2005) Open field trial of genetically modified
parthenocarpic tomato: seedlessness and fruit quality. BMC
Biotechnol. 5: 32.

Changes in auxin response from mutations in an AUX/IAA gene.
Science 279: 1371–1373.

auxin-dependent transcriptional repression during Arabidopsis

Tan, X., Calderon-Villalobos, L.A., Sharon, M., Zheng, C.,
Robinson, C.V., Estelle, M. et al. (2007) Mechanism of auxin percep-


